

SCIENTIFIC REPORT OF EFSA

Compendium of botanicals reported to contain naturally occurring substances of possible concern for human health when used in food and food supplements¹

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ABSTRACT

In April 2009, EFSA published on its website a Compendium of botanicals reported to contain toxic, addictive, psychotropic or other substances of concern. The purpose of the Compendium is to assist risk assessors responsible for the evaluation of specific ingredients in food supplements, in more easily identifying the compound(s) of concern on which to focus the assessment. The Scientific Committee worked on a second version of that Compendium between January 2010 and February 2012, considering botanicals that appear on a negative list or subject to restricted use (e.g. max. level or certain parts allowed only) in at least one European Member State. Two annexes have been added compared to the first version; the first one lists botanicals for which not enough information on possible substances of concern could be found, or for which the information present could not be verified. The second one lists botanicals for which, although some data were available, the Scientific Committee could not identify substances of concern, or other reasons for the inclusion in the compendium. This new “Compendium of botanicals reported to contain naturally occurring substances of possible concern for human health” replaces the first version published in 2009; it lists in alphabetical order botanicals without any judgment on whether they are suitable or not suitable for food applications in Europe; it has no legal or regulatory force pertaining to the legal classification of products or substances.

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KEY WORDS

Compendium, botanicals, food supplements, hazard identification, compounds of concern, adverse effects

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SUMMARY

Since August 2005, the Scientific Committee (SC) of EFSA has been developing a compendium (in table format) of compounds present in botanicals, that can be of concern for human health. This work has been undertaken in cooperation with the Advisory Forum Representatives of the European Member States. A first version of the Compendium entitled “Compendium of botanicals reported to contain toxic, addictive, psychotropic or other substances of possible health concern” was published on the EFSA website in April 2009. At that time, the SC underlined that the compendium of botanicals is a living document and should be updated on a regular basis by EFSA.

As a follow up, the SC considered botanicals appearing on a negative list or subject to restricted use (e.g. max. level or certain parts allowed only) in at least one European Member State. A literature search was performed to get information on compound(s) and/or possible health effects that would have motivated the insertion of the botanicals in the above-mentioned lists. In cases where not enough information on possible substances of concern could be found, or for which the information present could not be verified, the botanical species have been transferred to an attached “insufficient information” list (Annex A). In cases where some data were available, but the Scientific Committee could not identify substances of concern, or other reasons for the inclusion in the compendium, the botanical species were then transferred to another attached list (Annex B).

The resulting “Compendium of botanicals reported to contain naturally occurring substances of possible concern for human health” replaces the previous version published on the EFSA website in 2009. The Compendium is intended to help with the safety assessment of botanicals and botanical preparations intended for use as food supplements, by facilitating hazard identification. The Compendium aims at flagging plants or part of plants or compounds of possible concern for human health naturally present in the listed botanicals and that therefore require specific attention while assessing the safety of products containing such botanical(s). It is underlined that the presence of a substance of concern in a given botanical does not necessarily mean that this substance will also be present in the botanical preparation and, if present, that it is at a dosage causing a health concern. The absence of a given botanical species in this Compendium cannot be interpreted as this species is devoid of compounds hazardous for human health. In the same way, not mentioning a specific part of plant does not imply absence of substance(s) of concern in this part. The compendium has no legal or regulatory force pertaining to the legal classification of products or substances.

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BACKGROUND AS PROVIDED BY EFSA

In September 2009, the Scientific Committee of EFSA published a guidance document for the safety assessment of botanicals and botanical preparations intended for use as ingredients in food supplements. The opinion specifies what data are needed to carry out such safety assessments. It also suggests a two-tiered scientific approach depending on the existing level of knowledge on a given botanical and the substance(s) it contains. Moreover, working together with EU Member States, EFSA has also established a large database by compiling the available literature data and other information on a large number of botanicals and botanical preparations which have been reported to contain substances that may be of health concern when used in food or food supplements.

The Compendium comprises around 900 botanical entries, identifying for each of them the scientific name, the most common synonyms, the plant part containing compound(s) of concern, the chemical(s) of concern, specific remarks and references of relevance for a safety assessment.

The purpose of the Compendium is to assist risk assessors responsible for the evaluation of specific ingredients in food supplements, in more easily identifying the compound(s) of concern on which to focus the assessment, making then use of the above-mentioned guidance document to assess whether the considered botanical preparation is safe.

The Scientific Committee and Advisory Forum of EFSA, as well as the representatives of the stakeholders and Member States Competent Authorities who participated in the workshop organised in Athens on 24 November 2009 to debate such issues expressed their appreciation for the results achieved so far by EFSA. They underlined the importance of developing further the Compendium to include more botanical entries and regularly updating the provided information with most recent data. An additional recommendation made to EFSA was to incorporate in the Compendium information on botanicals and botanical preparations that have no history of use in the European Union but that are reported to have a history of traditional use in third countries.

TERMS OF REFERENCE AS PROVIDED BY EFSA

The Scientific Committee is requested by the European Food Safety Authority to carry out a bi-annual review of the Compendium of botanicals reported to contain toxic, addictive, psychotropic or other substances of concern. To this end, the Scientific Committee is requested to:

- Include in the Compendium the missing botanical species containing compounds of possible concern for human health and currently being used as ingredients in food supplements in the European Union;
- Update, where necessary, the information on botanical species already included in the Compendium;
- Develop and test a practical approach to identify, classify and include in the Compendium botanicals and botanical preparations that have no history of use in the European Union but could enter the European market at some point because of having such history of traditional use in ultra-peripheral regions of the European Union, i.e. overseas territories of the European Member States, or in third countries.

1. Preamble

In June 2004, the Scientific Committee (SC) of EFSA published a discussion paper on botanicals and botanical preparations widely used in food supplements and related products. Concerns about quality and safety issues were expressed, as well as the need for a better characterisation of the range of products on the market and for harmonising risk assessment and consumer information approaches. The discussion paper was brought to the attention of the members of the Advisory Forum, who confirmed the importance of the issues addressed by the paper for their countries. EFSA therefore mandated its Scientific Committee in August 2005 to develop guidance for the safety assessment of botanicals and botanical preparations, as well as a compendium (in table format) of compounds present in botanicals, that can be of concern for human health. A first version of the guidance document and the compendium were published on the EFSA website in June 2008.

As from May 2008, work was undertaken by the EFSA Scientific Committee (SC), in cooperation with the Advisory Forum Representatives of the European Member States, to develop further the compendium, based on a compilation of lists of botanicals made by Member States competent authorities, or international organisations such as the Council of Europe (see the “sources of information” section of this document). When information was available, the Scientific Committee identified and characterised the compound(s) of possible concern for human health. The “compendium of botanicals reported to contain toxic, addictive, psychotropic or other substances of possible health concern” was published on the EFSA website in April 2009 (EFSA Journal 2009; 7(9):281). The SC underlined that the compendium of botanicals is a living document and should be updated on a regular basis by EFSA.

As a follow up, the SC extended its work to the analysis of official positive and negative lists available in European Member States. The SC used in particular the overview prepared by the Association of the European Self Medication Industry (AESGP, 2007) and focussed its work on botanicals appearing on a negative list or subject to restricted use (e.g. max. level or certain parts allowed only) in at least one European Member State. For new entries, i.e. not listed yet in the EFSA compendium, a literature search was performed to get information on compound(s) and/or possible health effects that would have motivated the insertion of the botanicals in the above-mentioned lists. Due to some limitations in data accessibility, e.g. availability of full articles, language issues, users of the compendium should check the completeness and relevance of these data for their assessment. Different sources like textbooks, peer-reviewed scientific articles and different databases were checked to find the most recent data. In cases where not enough information on possible substances of concern or adverse effects could be found, or for which the information present could not be verified, the botanical species have been transferred to an attached “insufficient information” list (Annex A). In cases where some data were available, but the Scientific Committee could not identify substances of concern, or other reasons for the inclusion in the compendium, the botanical species were then transferred to another attached list (Annex B). Considering the risk assessment approach described in the guidance for the safety assessment of botanicals and botanical preparations, it is underlined that Annex B cannot be considered as a list of “safe botanicals” for use in food supplements, since the Compendium identifies possible hazards in a non-exhaustive way and no risk assessment was performed. Both the compendium and Annex B are of particular use for Tier 1 of the safety assessment framework for specific botanical preparations, as described in the guidance adopted by the Scientific Committee in 2006⁴. Botanicals mentioned in Annex A would then be candidates for a direct evaluation under Tier 2, following provision of additional data needed for the assessment. The search for information from literature for the newly added botanical species ended in October 2011. It should be underlined that the working group could not update the information for most of the botanical species listed in the first version of the compendium, although new data may have become available since 2008.

⁴ See <http://www.efsa.europa.eu/en/efsajournal/pub/1249.htm>

The present “Compendium of botanicals reported to contain naturally occurring substances of possible concern for human health” replaces the previous version published on the EFSA website in 2009.

2. Legal disclaimer

This compendium lists in alphabetical order botanicals without any judgment on whether they are suitable or not suitable for food applications in Europe. The compendium is part of a preparatory work undertaken by EFSA to harmonise the methodology for assessing the safety of botanicals and botanical preparations used in food. The compendium has no legal or regulatory force and may not be used as support or evidence in any disagreement or dispute pertaining to the legal classification of products or substances.

3. Purpose of the Compendium

The Compendium is intended to help with the safety assessment of botanicals and botanical preparations intended for use as food supplements, by facilitating hazard identification. The Compendium aims at flagging plants or part of plants or compounds of possible concern for human health naturally present in the listed botanicals and that therefore require specific attention while assessing the safety of products containing such botanical(s). It is underlined that the presence of a substance of concern in a given botanical does not necessarily mean that this substance will also be present in the botanical preparation and, if present, that it is at a dosage causing a health concern. This depends largely on the plant part used, the preparation method and the conditions of use. For some of the compounds flagged in the compendium, health-based guidance values (e.g. ADIs) have been established, but are not mentioned in this compendium. This compendium does not address possible synergies or antagonisms between botanical substances, nor possible interactions with other products that would need to be taken into account when assessing safety, as described in the EFSA guidance for the safety assessment of botanicals and botanical preparations.

4. Structure of the Compendium

The Compendium contains specific information organised in 6 columns.

In the first column the scientific name can be found which is based primarily on the taxonomy database of Kew taken as reference⁵. If not found in the Kew database, the ARSGRIN is used⁶. Commonly used synonyms are also mentioned into brackets. The whole genus is mentioned in this column when evidence is available that several species of the genus contain the same group of molecules of concern. In such case, the botanical species considered in the national lists are then brought under the appropriate spp.

Some European Member States considered fungi in their lists of plants, although they belong to another kingdom than botanicals. It was decided to extend the scope of the compendium so that it covers also fungi. The Index Fungorum (www.indexfungorum.org) was used as the main source of information for scientific names and families.

In the second column the family name is given. In many cases, botanicals from a given family contain similar groups of compounds. Therefore, knowing the family name may provide indications of the

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See http://apps.kew.org/wcsp/prepareChecklist.do;jsessionid=671510C7F22D4F19BEC4567F8810F73D?checklist=selected_families%40%40179280620101519590

⁶ See <http://www.ars-grin.gov/cgi-bin/npgs/html/taxgenform.pl>

possible presence of similar compounds in other species not included in the compendium. “The Plant List” database (www.theplantlist.org) was used as the main source of information. When the scientific name of the botanical was not an officially accepted one (“unresolved”), the ARSGRIN database was consulted.

The third column contains the plant parts in which the compounds of concern were reported to be present.

The fourth column lists the main compounds of concern and the chemical class to which they belong. When information on the amount present is available, this information is also mentioned. This column is left blank in cases no substances of concern could be identified in spite of available information on adverse effects. It is not the intention of the compendium to list all biologically active substances present in a given botanical; as indicated in the title, the Compendium focuses on substances of concern and adverse effects reported in the literature.

The fifth column deals with information concerning adverse health effect(s) found in the literature but that cannot be associated to the compound(s) of concern listed in the fourth column. In some cases, information on composition is also provided.

The last column contains selected reference(s) retrieved from literature searches for the data given, and/or standard reference text books providing monographs or more general scientific information for the botanicals considered.

The absence of a given botanical species in this Compendium cannot be interpreted as this species is devoid of compounds hazardous for human health. In the same way, not mentioning a specific part of plant does not imply absence of substance(s) of concern in this part. The main reasons for a botanical species not to appear in the compendium are the following:

- It did not appear in any of the national lists considered.
- Where no or insufficient data were available, the botanical was then transferred to Annex A.
- Where data were available and no indication of the presence of substances of concern or adverse effects could be found; the botanical was then transferred to Annex B.

5. Recommendations

The Scientific Committee underlines that the Compendium is a living document and should be updated on a regular basis by EFSA. The Compendium is therefore open for additional contributions and comments.

The Scientific Committee recommends as a follow-up activity that a systematic literature review is performed for the botanical species listed in annexes A and B.

Sources of information used to compile the list of botanical species to review

Country	Reference
Austria	List of Botanicals not admitted or restricted in food in Austria: (Codex Unterkommission Nahrungsergänzungsmittel) 9/7/2005
Belgium	Decree of 29 August 1997 on the manufacturing and placing on the market of foodstuffs which consist of plants or to which plants are added. List 1 - Plants that cannot be used in or as foodstuffs List 2 - Edible mushrooms List 3 - Plants that can be used in food supplements
Bulgaria	Decree on food supplements - Annex 4
Croatia	Regulation on foods to meet special nutritional requirements - Annex VIII
Czech Republic	Regulation on the requirements for food supplements and the addition of nutrients to foodstuff - Annex IV
Czech Republic	Recommendations of herbals which should only be used in food supplements under certain restrictions - State Institute of Drug Control
Denmark	Danish list concerning toxicological evaluation of plants in food supplements; The list contains plants considered as unacceptable, plants with a restriction on daily use (max. level), and plants that are evaluated at a daily dose ("Drogelister" (2000) and later update March 2011)
Denmark	The departmental order of the Danish Ministry of Health no. 698 (31. August 1993) List of euphorbiants. (Latest update 11. April 2007)
Estonia	Decree 59/2005 on establishing a list of plants for pharmaceutical use - Positive and negative lists of plants which may or may not be used in food supplements
Finland	Decision 1179/2006 on a list of medicines - Annex 2 negative list of herbal ingredients which cannot be used in food supplements
France	French Pharmacopoeia (10 th edition): List A of medicinal plants with a traditional use and List B of medicinal plants with a traditional use but whose possible undesirable effects exceed expected beneficial therapeutic effect.
Hungary	Horacsek M. 2005. Food Supplements and special-purpose foods. Komplementer Medicina. Vol 1-2, pp. 32-37 - List of herbal ingredients whose use in food supplements is permitted
Iceland	Medicines Control Agency - List of hebal ingredients A) for food use, B) for medicinal use, C) needing a case-by-case assessment, N) a natural medicine
Italy	Italian Ministry of Health - Plants not suitable for use in food supplement manufacturing - Positive list of herbal substances which may be used in the manufacture of foodstuffs
Latvia	Regulation on the labelling of food supplements - Annex II list of herbals whose use is restricted in food supplements - Annex III list of herbals whose use is restricted to certain levels and parts of plants
Netherlands	Dutch Regulation implementing the Law 19 January 2001 on Goods and identifying pyrrolizidine alkaloids containing plants (for which a maximum limit of 1 µg/kg or per litre is imposed) (E1) and plants not to be used in herbostic products (E2)
Norway	Regulation 1565/1999 on medicinal product classification. Herbal substances are classified as H) general food use, L) medicine, LR) prescription only medicine
Poland	Office for Registration of medicinal products, medicinal devices and biocidal products - two lists of herbals which may or may not be used in food supplements
Romania	Ordinance 244/2005 on herbals and partially processed herbals used in food supplements - contains a list of plants unsuitable for human consumption, and a list of plants which may be used in food supplements
Slovenia	Rules for the classification of herbals Nr. 133/03 - Contains a list of herbals classified as H) can be used in foodstuffs, Z) for the prevention and treatment of disease, ZR) prescription needed, ND, prohibited because of their toxic potential
Spain	Spanish Regulation (Ministerio de Sanidad y Consumo Orden SCO/ 190/2004) concerning plants for which public sale is forbidden or limited because of toxicity
Sweden	National Food Administration - List of plants considered as not suitable in foods
United Kingdom	Medicines and Healthcare Products Regulatory Agency - Indicative list of herbals which have a reported medicinal, food or cosmetic use.

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of possible concern for human health when used in food and food supplements

EU/International Organisation	Reference
AESGP	The Regulatory Framework for Food Supplements in Europe
Council of Europe	Plants assessed as flavourings by the Council of Europe in 2000 and 2004 belonging to Category 3 or 4 (restrictions recommended for use) (H1 and H2 respectively) or as Category 5 (restrictions recommended and further data required) (H3) or Category 6 (considered not appropriate for human consumption) (H4)
Council of Europe	Active principles (constituents of toxicological concern) contained in natural sources of flavourings. Council of Europe, 2004
EMEA/EMA	Plants containing toxic substances (CPMP / EMEA, 1992)
EMEA/EMA	Plants assessed as medicinal products by the EMEA/HMPC since its inception, and previously by the Working Party on Herbal Medicinal Products between 1998 and 2004
EMEA/EMA	Final Public Statement on the use of herbal medicinal products containing estragole, Committee on Herbal Medicinal Products (HMPC), London 23 November 2005
EMEA/EMA	Final Public Statement on the use of herbal medicinal products containing methyleugenol, Committee on Herbal Medicinal Products (HMPC), London 23 November 2005
EMEA/EMA	Final Public Statement on the risk associated with the use of herbal products containing Aristolochia species, Committee on Herbal Medicinal Products (HMPC), London 23 November 2005
EMEA/EMA	Final Public Statement on the use of herbal medicinal products containing pulegone and menthofuran, Committee on Herbal Medicinal Products (HMPC), London 23 November 2005
EMEA/EMA	Final Public Statement on the use of herbal medicinal products containing asarone, Committee on Herbal Medicinal Products (HMPC), London 23 November 2005
ESCP	Plants assessed as medicinal products by ESCOP (2003)
EuroFIR-NETTOX	Pilegaard K, Eriksen FD, Soerensen M, Gry J. (2007) EuroFIR-NETTOX Plant List. European Food Information Resource Consortium (EuroFIR). ISBN 0 907 667 570
WHO	Plants assessed as medicinal products by WHO in 1999 (Vol. 1), 2002 (Vol. 2) and 2005 (Vol. 3)

COMPENDIUM OF BOTANICALS REPORTED TO CONTAIN NATURALLY OCCURRING SUBSTANCES OF POSSIBLE CONCERN FOR HUMAN HEALTH WHEN USED IN FOOD AND FOOD SUPPLEMENTS

This compendium lists in alphabetical order botanicals without any judgment on whether these are suitable or not suitable for food applications in Europe. This compendium is part of a preliminary work undertaken by EFSA to harmonise the methodology across its panels for assessing the safety of botanicals and botanical preparations used in food and food supplements. Without prejudice to the existing legal framework, such compendium has no legal status and may not be used as support or evidence in any disagreement or dispute pertaining to the legal classification of products or substances. This compendium is a living document and is therefore open for additional contributions and comments.

Botanical name	Family	Parts of plants of possible concern	Chemical of concern	Remarks on toxic/adverse effect(s) not known to be related to the identified chemical(s) of concern	Specific References
<i>Abus precatorius</i> L.	Fabaceae (Leguminosae)	Seed	Glycoproteins (lectins), e.g. abuin		Frome D., Pfander H.J. and Anton R. 2009. Plantes à risques. Ed. Tec et Doc-Lavoisier, ISBN: 978-2-7430-0907-1
<i>Acacia</i> spp.	Fabaceae (Leguminosae)	Bark, leaf and seed	Genus in which species may contain dimethyltryptamine derivatives and cyanogenic glycosides (e.g. prunasin, sambubitrin, acetapellin).		Siegler D.S. and Ehinger J.E. 1987. Cyanogenic Glycosides in Amygdalaceae of Mexico and Central America. Southwest. Nat. 32(4): 489-503
<i>Acichia abrotanoides</i> Vis.	Asteraceae (Compositae)	Aerial part	Essential oil: bicyclic monoterpenes: beta-thujone (16.8%), pinoasone (15.5%), camphor (14%), and monodipentene etheroxide: 1,8-cineole (11.2%)		Bloch C. et al. 1988. On the composition of <i>Acichia abrotanoides</i> (Vis.) Vis. essential oil. Flavour Frag. J. 3(3): 101-104
<i>Acichia fragrantissima</i> Sol.Bip.	Asteraceae (Compositae)	Aerial part	Essential oil: bicyclic monoterpenes: thujones		Elgamal M.H.A. et al. 1991. Constituents of <i>Acichia fragrantissima</i> . Fitoterapia. 62(4): 382
<i>Acichia millefolium</i> L.	Asteraceae (Compositae)	Aerial part	Essential oil from fresh plant: bicyclic monoterpenes: alpha-thujone (0.28%), beta-thujone (1.60%), camphor (2.93%) and monodipentene etheroxide: 1,8-cineole (2.24%), Essential oil from dried plant: alpha-thujone (0.40%), beta-thujone (3.21%), camphor (4.43%), 1,8-cineole (4.54%), Essential oil from flower: alpha-thujone (1.02%), beta-thujone (0.59%), camphor (17.8%), 1,8-cineole (3.70-9.6%), Essential oil from leaf: alpha-thujone (0.50%), beta-thujone (0.25%), camphor (16.80%), 1,8-cineole (0.09%)		Council of Europe. 2000. Natural Sources of Flavonurings. Rep No.1, ISBN: 978-92-871-4324-2
<i>Acockenhiera</i> spp.	Apocynaceae	Whole plant	Genus in which species may contain cardenolide glycosides: e.g. ouabain		Kokwaro J.O. 1976. Medicinal plants of East Africa. East Africa Literature Bureau, General Printers Ltd. Nairobi, Kenya
<i>Aconitum</i> spp.	Ranunculaceae	Whole plant	Genus in which species may contain diterpene alkaloids: e.g. aconitine, hypaconitine, mesaconitine.		Omino EA and Kokwaro J.O. 1993. Ethnobotany of Apocynaceae species in Kenya. J. Ethnopharmacol. 40(3): 167-180.
<i>Acorus calamus</i> L.	Acetaceae	Leaf and rhizome	Phenylpropanoids: e.g. methylchavicol in rhizome (1%) and in unspecified quantities in other parts. Essential oil from the rhizome: phenylpropanoids: e.g. beta-asarone (Z-isasarone).		Frome D., Pfander H.J. and Anton R. 2009. Plantes à risques. Ed. Tec et Doc-Lavoisier, ISBN: 978-2-7430-0907-1
<i>Acorus calamus</i> L. var. <i>calamus</i>	Acetaceae	Leaf and rhizome	Triplic plant: phenylpropanoids: e.g. beta-asarone (50-65% in the essential oil from the leaf, 9-15% in the essential oil from the rhizome).		Duke J.A. 1992. Handbook of Phytochemical Constituents of GRAS Herbs and Other Economic Plants. CRC Press, Inc. Boca Raton, FL. Nico Vermeulen-The Complete Encyclopedic of Condenser Plants. Rebo International, Netherlands, 1998. ISBN 90-366-1584-4
<i>Acorus calamus</i> L. var. <i>varangustatus</i>	Acetaceae	Leaf and rhizome	Tetraploid plant: phenylpropanoids: e.g. beta-asarone (85-95% in the essential oil from the fresh rhizome, 4-8.3% in the dried rhizome)		EMA-HMPC. 2005. Public statement on the use of herbal medicinal products containing asarone. EME/AT/HMPC/139215/2005
<i>Acorus gramineus</i> Sol.	Acetaceae	Leaf and rhizome	Essential oil from the rhizome: (0.5-0.9%) with phenylpropanoids: cis- and trans- isasarones, methylisogerol, cis-methylisogerol and safrole, calicumoxalate lactides		EMA-HMPC. 2005. Public statement on the use of herbal medicinal products containing asarone. EME/AT/HMPC/139215/2005
<i>Actaea spicata</i> L.	Ranunculaceae	Whole plant	Benzylisquinoline alkaloids: e.g. magnoflorine, coryduberine		Hegmauer R. 1992. Chemotaxonomie der Pflanzen. Vol. 10. Birkhäuser Verlag, ISBN: 3-7643-2576-X.
<i>Adenia</i> spp.	Passifloraceae	Root and seed	Genus in which species may contain lectins		Council of Europe. 2008. Natural sources of Flavonurings. Report No. 3. Council of Europe Publication, ISBN: 978-92-871-5822-3.
<i>Adenium</i> spp.	Apocynaceae	Root and stem (latex), seed	Genus in which species may contain cardenolide glycosides: e.g. edujine		Pelosi E. et al. 2005. Flabonoids-inactivating proteins and other lectins from <i>Adenia</i> (Passifloraceae). Toxicon. 46(6): 656-663
<i>Adiantum vesica</i> Ness.					Barbieri L. et al. 1984. Volkensin, the toxin of <i>Adenia volkensii</i> (Kiyamifil plant). FEBS Letters. 171, (1, 2): 277-279
<i>Secusulfida adhatoda</i> L.					Schneizer GH. and Fakim AG. 2003. Medicinal plants 1. Plant resources of tropical Africa 11 (1). PROTA Foundation. Backhuys Publishers/CTA Wageningen, Netherlands.
<i>Adonis</i> spp.	Ranunculaceae	Whole plant	Genus in which species may contain cardenolide glycosides: e.g. adonitoxin		Frome D., Pfander H.J. et Anton R.2009. Plantes à risques. Ed. Tec et Doc-Lavoisier, ISBN: 978-2-7430-0907-1
<i>Adiantum vesica</i> Ness.					Brunelton J. 2005. Plantes toxiques, 3ème édition, Ed. Tec et Doc-Lavoisier, ISBN: 2-7430-098-7
<i>Aegle marmelos</i> (L.) Corrêa	Rutaceae	Leaf	Quinoline alkaloids: e.g. aegeline, skimmianine.	Ethnolic extract of leaf shows dose-dependent decrease of testosterone levels, spermatogenesis and fertility in rats.	Charuan A. et al. 2007. Suppression of fertility in male albino rats following the administration of 50% ethanolic extract of <i>Aegle marmelos</i> . Contraception. 76: 474-481
<i>Aegle marmelos</i> (L.) Corrêa	Rutaceae	Leaf	Quinoline alkaloids: e.g. aegeline, skimmianine.	A dry aqueous extract, given perorally to mice in doses of 1 g/kg body weight (N=7) for 15 days reduced the serum levels of the thyroid hormone T3 but not T4 compared to control mice receiving vehicle.	Charuan A. et al. 2008. Reversible changes in the antileptin induced by <i>Aegle marmelos</i> in male albino rats. Syst. Biol. Reprod. Med. 54: 240-246
<i>Aegle marmelos</i> (L.) Corrêa	Rutaceae	Leaf	Quinoline alkaloids: e.g. aegeline, skimmianine.	poisonous potential is questionable as no toxicity is found in mice and quinine pigs. Probably toxic information comes from confusion with <i>Conium maculatum</i> or plants infested with the rust fungus <i>Puccinia aethusa</i> with consequent production of larger amounts of the toxins	Kar A. et al. 2002. Relative efficacy of three medicinal plant extracts in the alteration of thyroid hormone concentrations in male mice. J. Ethnopharmacol. 81: 281-285.
<i>Aegle marmelos</i> (L.) Corrêa	Rutaceae	Leaf	Quinoline alkaloids: e.g. aegeline, skimmianine.		Yedav NP. and Chandia CS. 2009. Phytochemical and pharmacological profile of leaves of <i>Aegle marmelos</i> (Linn). Pharma Review. 7(42): 144-149.
<i>Aitonomum angustifolium</i> (Som.) K.Schum.	Zingiberaceae	Seed	Essential oil: monodipentene etheroxide: 1,8-cineole (4%)		Frome D., Pfander H. J. and Alford I. 2005. Poisonous plant. Blackwell, ISBN: 1-874545-94-4
<i>Aitonomum angustifolium</i> (Som.) K.Schum.	Zingiberaceae	Seed	Essential oil: monodipentene etheroxide: 1,8-cineole (4%)		Frome D., Pfander H. J. and Alford I. 2005. Poisonous plant. Blackwell, ISBN: 1-874545-94-4
<i>Aitonomum melegueta</i> K.Schum. (<i>Aitonomum melegueta</i> Krosch.)	Zingiberaceae	Fruit and seed	Piperidine alkaloids: e.g. piperine		Brunelton J. (1996). Plantes toxiques - Vegetaux dangereux pour l'homme et les animaux. Tec&Doc, ISBN: 2-7430-169-0
<i>Aiphanthus</i> spp.	Amariyllidaceae	Leaf and rhizome			Igwé SA. et al. 1999. Ocular toxicity of <i>Aitonomum melegueta</i> (alligator pepper) on healthy igbos of Nigeria. J. Ethnopharmacol. 65: 203-206
<i>Aiphanthus</i> spp.	Amariyllidaceae	Leaf and rhizome			Kemthong P. et al. 2002. Effect of <i>Aitonomum melegueta</i> and <i>Piper guineense</i> on sexual behaviour of male rats. <i>Behav. Pharmacol.</i> 13: 243-247.
<i>Aiphanthus</i> spp.	Amariyllidaceae	Leaf and rhizome			Lachman-White D.A., Adams CD. and Troz LO. 1992. A guide to the medicinal plants of Coastal Guyana. Commonwealth Science Council. Technical Publications Series 225. London, UK.
<i>Aiphanthus</i> spp.	Amariyllidaceae	Leaf and rhizome			Fuller TC and McClintock E. 1986. Poisonous plants of California. Univ. California Press, Berkeley, Calif., USA
<i>Aiphanthus</i> spp.	Amariyllidaceae	Leaf and rhizome			Van Wyk BE, Van Oudshoorn B and Gericke N. 1997. Medicinal plants of South Africa. Briza, Pretoria.

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<i>Agastache</i> spp. (Labiatae)	Lamiaceae	Whole plant	Genus in which species may contain in their essential oil phenylpropanoids: e.g. methylchavicol and/or methyl Eugenol and/or monoterpenes: e.g. pulegone	<i>A. rugosa</i> : 83%-96% methyl Eugenol and 5-chemotypes: T1: methylchavicol, T2: methyl Eugenol, T3: methyl Eugenol and limonene, T4: menthones, T5: menthone and eudipone. <i>A. foeniculum</i> : 43%-74% methylchavicol	Charles DJ et al. 1991. Characterisation of essential oil of <i>Agastache</i> species. <i>J Agr Food Chem.</i> 39(11), 1946-1949.
<i>Agathosma coronilla</i> Bartl. & Wendl.	Rutaceae	Leaf	Essential oil: 2% (summer) - 5% (winter) with 50% phenylpropanoids: e.g. methylchavicol and anethole.		
<i>Agrostemma githago</i> L.	Caryophyllaceae	Seed	Triterpenoid saponins: e.g. githagin (7%), agrostemmic acid		
<i>Albanthus alissima</i> (Mill.) Swingle	Simarubaceae	Whole plant	Indolomonoterpene alkaloids: e.g. cathine- β -one and beta-carboline derivatives		
<i>Albizia julibrissin</i> Durazz.	Leguminosae (Fabaceae)	Seed	Genus in which species may contain saponins and diterpene derivatives: e.g. phorbol esters	Unknown neurotoxin in the seed	
<i>Aleurites</i> spp.	Euphorbiaceae	Whole plant	Genus in which species may contain saponins and diterpene derivatives: e.g. phorbol esters		
<i>Alisma plantago-aquatica</i> L.	Alismataceae	Whole plant	Genus in which species may contain unsaturated pyridizidine alkaloids	Toxicity associated with all plant parts; compound(s) are unknown	
<i>Alisma spp.</i>	Boraginaceae	Root	Genus in which species may contain unsaturated pyridizidine alkaloids		
<i>Alismata cathartica</i> L.	Apocynaceae	Whole plant	Iridoid lactones: e.g. alismaridin	Purgative effect (laxac)	
<i>Alcea</i> spp.	Asparagaceae (Agavaceae)	Leaf	Genus in which species may contain hydroxyanthracene derivatives: C-glycosides of 1,8-dihydroxy anthrones: e.g. alchins	Alchins are present only in the juice obtained from the pericycle cells and adjacent leaf parenchyma	
<i>Alhania galingga</i> (L.) Willd.	Zingiberaceae	Rhizome	Essential oil: phenylpropanoids: e.g. methyl Eugenol in unspecified quantities		
<i>Alhania officinarum</i> Hance	Zingiberaceae	Rhizome	Essential oil: monoterpene etheroxide: 1,8-cineole (65%)		
<i>Alistona</i> spp.	Apocynaceae	Bark and leaf	Genus in which species may contain monoterpenoid indole alkaloids: e.g. alstonine, alstonidine, plicnine.		
<i>Anagyris</i> spp.	Ameyllidaceae	Bulb	Genus in which species may contain isoquinoline alkaloids: e.g. lycorine, ambelicine, caranine.		
<i>Anagyris majus</i> L.	Apiaceae (Umbelliferae)	Fruit and leaf	Furanocoumarins: e.g. 5-methoxypsoralen.		
<i>Annona versipaga</i> Lam.	Apiaceae (Umbelliferae)	Aerial part	Furochromones: e.g. khellin, versipagine.		
<i>Anagyris communis</i> L. (<i>Platanus anagyalis</i> Batsch, P. <i>dulcis</i> (Mill.) D. A. Webb)	Rosaceae	Seed	Cyanogenic glycosides: e.g. purasin corresponding to 300-3400 mg HCN/Kg		
<i>Anabasis aphylla</i> L.	Amaranthaceae (Chenopodiaceae)	Aerial part	Pyridine alkaloids: e.g. arabosine, N-methylarabosine, arabosamine and isonicotidine.		
<i>Anacardium occidentale</i> L.	Anacardiaceae	Leaf and pericarp	Alcenylyphenols: e.g. anacardic acids, cardanol.		
<i>Anacyclus officinarum</i> Hayne See <i>Anacyclus azyrthium</i> (L.) Lab.					
<i>Anacyclus pyrethrum</i> (L.) Lag. (<i>Anacyclus officinarum</i> Hayne)	Compositae (Asteraceae)	Root	Akylaridites: e.g. pellitorine	Seeds caused miscarriages in pregnant albino rats when fed for 10 days after copulation at a daily dose of 175 mg/kg bw. Skeletal and visceral malformations were commonly observed in the fetuses	
<i>Anadenanthera</i> spp.	Leguminosae (Fabaceae)	Bark and seed	Genus in which species may contain indolamines derived from tryptamines: e.g. bufotenine and beta-carbolines		
<i>Anagallis arvensis</i> L.	Primulaceae	Whole plant	Tetracyclic terpenic saponins: e.g. anagalline and oxygenated tetracyclic terpenes: e.g. arvensins, cucurbitacinones E, B, D and I.		
<i>Anagyris foetida</i> L.	Leguminosae (Fabaceae)	Leaf	Quinolizidine alkaloids: e.g. cystine and anagyrrine		

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<i>Amorpha paniculata</i> C. DCbr. (A. caerulea (L.) Wright & Arn.)	Menispermaceae	Fruit and seed	Sequitriene lactones, e.g. picrotoxin, picrodalin.		Frome D, Pfander HJ and Anton R. 2009. <i>Plantas a rasques</i> . Ed. Tec et Doc-Lavoisier. ISBN: 978-2-7430-0907-1
<i>Archusa</i> spp.	Borraginaceae	Flower and leaf	Genus in which species may contain unsaturated pyridizidine alkaloids, e.g. Nyoosamine		Shibano T et al. 2005. Pyridizidine alkaloids from <i>Archusa strigos</i> and their antileishman activity. <i>Phytochemistry</i> . 66(13): 1593-1600. Broch-Due AI et al. 1980. Alkaloids of <i>Archusa officinalis</i> L. Identification of the pyridizidine alkaloid typosamine. <i>Acta Chemica Scandinavica</i> . Series B: Organic Chemistry and Biochemistry. B34(1): 75-7.
<i>Andra araroba</i> Aquilar See <i>Vatica rosepis araroba</i> (Aquilar)					
<i>Andra nemis</i> (W.Wright) Kuntz	Leguminosae (Fabaceae)	Bark	Isocoumarin alkaloids, e.g. berberine isoflavonoids derivatives, e.g. biochanin-A, calycosin and genistein		Duke JA, Bogershuizen-Godwin MJ and Ohtsuen AR. 2009. <i>Dukes Handbook of Medicinal Plants of Latin America</i> . CRC Press Taylor & Francis. ISBN: 13: 978-1-4200-4316-7
<i>Andropogon paniculata</i> (Burm.f.) Nees (<i>Justicia paniculata</i> Burm. f.)	Acanthaceae	Aerial part	Diterpene lactones and derivatives from dried aerial part: e.g. andrographolide (2.5-4.4%), dehydroandrographolide (1.4-2.1%), neoandrographolide (1.4-1.9%) and deoxyandrographolide-19-beta D glucoside (0.7-1.8%)	Abortifacient effect reported in studies with rabbits and mice (WHO 2002).	Akhtaria MA and Munglani P. 2000. Aspects of the male reproductive toxicological anti-fertility property of andrographolide in albino rats: effect on the testis and the cauda epididymal spermatozoa. <i>Phytoter. Res.</i> 14: 432-435. Akhtaria MA et al. 1990. Anti-fertility effect of <i>Andropogon paniculata</i> (Nees) in male albino rats. <i>Indian J. Exp. Biol.</i> 28: 421-426. Paratapanich C et al. 2007. HPLC determination of active diterpene lactones from <i>Andropogon paniculata</i> Nees planned in various seasons and regions in Thailand. <i>Thai J. Pharm. Sci.</i> 31: 91-99. World Health Organization. 2002. WHO monographs on selected medicinal plants. Geneva. ISBN 92.4.154537.2
<i>Andromeda</i> spp.	Ericaceae	Flower, fruit and leaf	Genus in which species may contain diterpenes, e.g. grayanotoxin (andromedotoxin)		Frome D, Pfander HJ and Anton R. 2009. <i>Plantas a rasques</i> . Ed. Tec et Doc-Lavoisier. ISBN: 978-2-7430-0907-1
<i>Andropogon citratus</i> DC. See <i>Cymbopogon citratus</i> (DC.) Stapf					Frome D, Pfander HJ and Anton R. 2009. <i>Plantas a rasques</i> . Ed. Tec et Doc-Lavoisier. ISBN: 978-2-7430-0907-1
<i>Antennaria</i> spp.	Ranunculaceae	Aerial part	Genus in which species may contain lactones, e.g. protoanemonins	protoanemonin only present in fresh herb	Frome D, Pfander HJ and Anton R. 2009. <i>Plantas a rasques</i> . Ed. Tec et Doc-Lavoisier. ISBN: 978-2-7430-0907-1
<i>Antennium graveolens</i> L.	Apiaceae (Umbelliferae)	Whole plant	Essential oil: phenylpropanoids, e.g. methylchavicol		Pfater KV. 2004. <i>The Handbook of Herb and Spices</i> vol. 2. CRC Press. ISBN: 0849312175
<i>Argemone</i> spp.	Apocynaceae (Umbelliferae)	Fruit and root	Genus in which species may contain coumarins, e.g. anethangelin, prangolin, oxipeucedalin hydrate, ostsofol and ostfol.		Sobong I, Andersson C, Gny J. 1996. Furcoumarins in Plant Food - exposure, biological properties, risk assessment and recommendations. <i>Terrahol</i> 1996:600. Ed. Nordic Council of Ministers. Copenhagen, ISBN 92 943 0. Available at: http://www.norden.org/sv/publikationer/publikationer/1996-600at_downloadpublicatlonfile
<i>Antanionum lewini</i> Hennings See <i>Lophophora williamsii</i> (Salin)-Dockrill M.Coutt.					
<i>Amorpha</i> spp.	Annonaceae	Whole plant	Genus in which species may contain acetogenins in the seed, e.g. amonacin, isocoumarin alkaloids in the bark, leaf, fruit and stem, e.g. amonetine, and monoterpene etheroxide in the fruit. 1,5-dicoum	<i>A. muricata</i> : total alkaloids 0.65 g/kg in leaf, 19.7 g/kg in root bark, 2.5 g/kg in stem bark, 0.5 g/kg. Bark is rich in cyanogenic glycosides, leaves contain small amounts and fruits only traces.	Caparros-Lefebvre D and Echeza A. 1999. Possible relation of atypical parkinsonism in the French west Indies with consumption of tropical plants: a case-control study. <i>Lancet</i> 354: 281-283. Hassal JA, et al. 1997. Isocoumarin derivatives isolated from the fruit of <i>Amorpha muricata</i> as 5-HT _{1A} receptor agonists in rats: unexplained antidepressive (lead) products. <i>J. Pharm. Pharmacol.</i> 49(11): 1145-1149. Council of Europe. 2000. <i>Natural sources of flavonoids</i> . Report No. 1. Council of Europe Publishing. ISBN 978-92-871-4324-2. Law C, et al. 2004. Nine new cycloac monoterpenylflavonol Ammonaceous acetogenins from <i>Amorpha montana</i> . <i>Planta Medica</i> . 70(10): 946-959. Pilegaard K., Eriksen F.D., Steensen M., Gny J. (2007) EGFR/RET/TOX plant list. European Food Information Resource Consortium (EuroFIR). ISBN: 0 907 667 570.
<i>Anthraxanthum odoratum</i> L.	Poaceae (Gramineae)	Aerial part	Coumarins: e.g. coumarin (5% of the dried part)		Council of Europe. 2000. <i>Natural sources of flavonoids</i> . Report No. 1. Council of Europe Publishing. ISBN 978-92-871-4324-2
<i>Antiaris toxicaria</i> (Pers.) Lessch.	Moraceae	Bark and leaf	Cardenolide glycosides, e.g. antiarins, toxicarosides B and C; (turanocoumarins		Carter CA et al. 1997. Toxicaroside A: A new cardenolide isolated from <i>Antiaris toxicaria</i> latex-derived dact poison. Assignment of the H ⁺ and ¹³ C-NMR shifts for an anthracenyl anthoxone. <i>Tetrahedron</i> 53(50): 16959-16968.
<i>Apocynum</i> spp.	Apocynaceae	Whole plant	Genus in which species may contain cardenolide glycosides and aglycones, e.g. oumarin, strophanthin.		EMEA. Committee for veterinary medicinal products. 1997. <i>Apocynum cannabinum</i> summary report. EMEA/MRL/569/99-Final
<i>Aquilegia vulgaris</i> L.	Ranunculaceae	Whole plant	Cyanogenic glycosides		Giffin M, Hobbs C, Utison R and Goldberg A. ISBN 0-8493-1675-8 Hegnauer R. 1990. <i>Chemotaxonomie der Pflanzen</i> . Birkhauser-Verlag. ISBN 3-7643-2576X.
<i>Arctostaphylos uva-ursi</i> (L.) Spreng.	Ericaceae	Leaf	Quinone glycosides, e.g. arbutin (5% -15%), methylarbutin (up to 4%)		Frome D, Pfander H.J. and Anton R. 2009. <i>Plantas a rasques</i> . Tec et Doc. ed. ISBN 978-2-7430-0907-1
<i>Arcaea catechu</i> L.	Arecaceae (Palmae)	Seed	Piperidine alkaloids, e.g. arecoline, arecaldine		British Herbal Compendium. 1993. Vol. 1: A handbook of scientific information on widely used plant drugs. Editor: P. Bradley. ISBN 978-0903032904
<i>Argemone mexicana</i> L.	Papaveraceae	Whole plant	Isocoumarin alkaloids, e.g. protopine, allocryptopine, sanguinarine		Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1188-8
<i>Argyranthemum frutescens</i> (L.) Sca.Bip. (<i>Chrysanthemum frutescens</i> L.)	Compositae (Asteraceae)	Aerial part	Acetylenic compounds, e.g. frutescinal isovalerate		Verna SK et al. 2001. <i>Argemone mexicana</i> poisoning: autopsy findings of two cases. <i>Forens. Sci. Int.</i> 115: 135-141

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<i>Argyrea</i> spp.	Convolvulaceae	Seed	Genus in which species may contain ergoline alkaloids	There is still discussion whether the ergoline alkaloids found are due to biochemical synthesis in the plant or due to fungal production	Steiner U. et al. 2006. Molecular characterization of a seed transmitted clavicipitaceous fungus occurring on dicotyledonous plants (Convolvulaceae). <i>Phyta</i> 22(4)(3):533-544.
<i>Artemisa</i> spp.	Araceae	Whole plant	Genus in which species may contain calcium oxalate raphides and some species saprom glycosides e.g. arolin		Ylian Y. 2002. Chinese herbal medicines: comparison and characteristics. Churchill-Livingstone, London. ISBN 044307-186-7
<i>Artemisia</i> spp.	Aristolochiaceae	Whole plant	Genus in which species may contain nitric phenanthrene derivatives, e.g. aristolochic acids, aristolactams		Froine D, Pflander HJ and Arton R. 2009. Plantes à risques. Ed. Tec et Doc-Lavoisier. ISBN : 978-2-7430-0907-1
<i>Artemisia chamissonis</i> Less.	Compositae (Asteraceae)	Whole plant	Sesquiterpene lactones (1.5%), and their esters, e.g. helenalin, amfifolins, chamissonolides	Helenalin reported to be the causative agent for oral toxicity	Burtonel J. 2005. Plantes toxiques (Végétaux dangereux pour l'homme et les animaux). Ed. Tec & Doc. Lavoisier, Paris. 3ème édition. ISBN : 2-7430-0806-7
<i>Artemisia montana</i> L.	Compositae (Asteraceae)	Whole plant	Sesquiterpene lactones and esters (0.2-0.5%) e.g.: helenalin and derivatives	Helenalin reported to be the causative agent for oral toxicity	Ed. Tec & Doc. Lavoisier, Paris. 3ème édition. ISBN : 2-7430-0806-7
<i>Artemisia abrotanum</i> L.	Compositae (Asteraceae)	Aerial part	Essential oil: bicyclic monoterpenes, e.g. alpha-thujone; monoterpene etheroxide: 1,8-cineole; phenylpropanoids: e.g. methyl Eugenol. Essential oil from leaf (1.4%); bicyclic monoterpenes: e.g. thujones (up to 70%), monoterpene etheroxide: 1,8-cineole (up to 60%).		Burtonel J. 2005. Plantes toxiques (Végétaux dangereux pour l'homme et les animaux). Ed. Tec & Doc. Lavoisier, Paris. 3ème édition. ISBN : 2-7430-0806-7
<i>Artemisia absinthium</i> L. (<i>Asium officinale</i> Brot., <i>Artemisia vulgaris</i> Lam.)	Compositae (Asteraceae)	Aerial part	Essential oil of (Z)-epoxy-ocimene chemotype: bicyclic monoterpenes, e.g. alpha-thujone (up to 0.30%), beta-thujone (up to 7.76%), camphor (0.19-3.30%), essential oil of abrotanum (0.58-0.71%), camphor (up to 0.31%), Essential oil of (Z)-santoniol acetate chemotype: alpha-thujone (1.32%), beta-thujone (18.72%), camphor (0.18%), Essential oil of beta-thujone chemotype: alpha-thujone (0.53-2.76%), beta-thujone (17.5-59.9%), camphor (0.10-0.16%), Essential oil of beta-thujone/epoxy ocimene mixed chemotypes: alpha-thujone (0.7-1.68%), beta-thujone (20.9-40.6%), Essential oil of cis-chrysanthenol chemotype: alpha-thujone 2.65-21.6%, beta-thujone (3.75-25.9%)		Council of Europe. 2000. Natural sources of Flavours. Report No. 1. Council of Europe Publishing. ISBN 978-92-871-4324-2
<i>Artemisia afra</i> Willd.	Compositae (Asteraceae)	Aerial part	Essential oil: bicyclic monoterpenes, e.g. alpha-thujone (62.9%), beta-thujone (15.07%), camphor (5.72%) and monoterpene etheroxide: 1,8-cineole (10.65%)		Council of Europe. 2005. Active principles (constituents of chemical concern) combined in natural sources of flavours. Ed. Council of Europe Publishing. http://www.coe.int/t/eseccol_convention/sc-sipublic_Health/Flavouring_substances/Active%20principles.pdf
<i>Artemisia annua</i> L.	Compositae (Asteraceae)	Leaf	Essential oil: bicyclic monoterpenes, e.g. camphor (2.58%-37.50%).	Contains sesquiterpene lactones, e.g. artemisinin (cardiarrane-type sesquiterpene lactone endoperoxide) and derivatives. Recommendation of WHO not to use artemisinin-containing herbs to avoid possible resistance of <i>Plasmodium</i> sp. (causing malaria)	Duke's Phytochemical and Ethnobotanical Databases. www.ars-grm.gov/dikey/ . Walt, J.M., & Breyer-Grandwilk, M.G. 1992. Medicinal and poisonous plants of Southern and Eastern Africa. E. & S. Livingston Ltd., Edinburgh and London OCLC NUMBER 1279138 van Wyk, B.-E., van Outshoorn, B. and Gerickse, N. 1997. Medicinal plants of South Africa. Biza, Pretoria, ISBN 1875093095
<i>Artemisia cina</i> Berg	Compositae (Asteraceae)	Flower bud	Essential oil: bicyclic monoterpenes (2.3%); e.g. santoni and eudesmanolide derivatives, etheroxide monoterpene: 1,8-cineole		Zhengwen Y. et al. 2010. Preliminary study of quality standards of essential oil in cultured <i>Artemisia annua</i> . <i>Zhongguo Yaoxue Zazhi</i> (Beijing, China), 45(2), 98-101.
<i>Artemisia eriantha</i> Ten.	Compositae (Asteraceae)	Aerial part	Essential oil: bicyclic monoterpenes, e.g. thujones (up to 90%).		Burtonel J. 2009. Pharmacognose, (phytochimie, Plantes médicinales), Ed. Tec & Doc. Lavoisier, Paris. 4ème édition. ISBN: 978-2-7430-1188-8
<i>Artemisia frigida</i> Willd.	Compositae (Asteraceae)	Aerial part	Essential oil: bicyclic monoterpenes, e.g. beta-thujone (5%).		Medical and aromatic plants - Industrial profiles. 2002. Edited by Dr. Roland Hartman. Volume 18. Edited by Collin W. Wright. Taylor and Francis ISBN: 0-415-27212-2
<i>Artemisia genipi</i> Stechm.	Compositae (Asteraceae)	Aerial part	Essential oil: monoterpene etheroxide: 1,8-cineole; bicyclic monoterpenes, e.g. alpha (26%), and beta (6.8%) thujones.		Council of Europe. 2005. Active principles (constituents of chemical concern) contained in natural sources of flavours. Ed. Council of Europe Publishing. http://www.coe.int/t/eseccol_convention/sc-sipublic_Health/Flavouring_substances/Active%20principles.pdf
					Lopes-Luz D. et al. 2008. Screening of chemical composition, antimicrobial and antioxidant activities of <i>Artemisia</i> essential oils. <i>Phytochemistry</i> 69(8), 1732-1738
					Council of Europe. 2000. Natural sources of Flavours. Report No. 1. Council of Europe Publishing. ISBN 978-92-871-4324-2.
					Bicht C. et al. 1982. On the composition of the essential oils of <i>Artemisia genipi</i> Weber and <i>Artemisia umbelliformis</i> Lam. <i>Zeitschrift für Lebensmittel-Untersuchung und -Forschung</i> , 175(3), 182-185.

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Botanical name	Family	parts of plants of possible concern	Chemical of concern	Remarks on toxic/adverse effects(s) not known to be related to the identified chemical(s) of concern	Specific References
<i>Artemisia maritima</i> L. (<i>Scorpioidium maritimum</i> (L.) Pojarkov.)	Compositae (Asteraceae)	Flower bud	Essential oil: monoterpene etheroxide: 1,8-cineole (41.1%), bicyclic monoterpenes e.g. L-(+)-camphor (20.3%), beta-thujone (1.1%), sesquiterpene lactones: e.g. santolin and eudesmanolide derivatives		Shah A et al. 2011. Studies on the chemical composition and possible mechanisms underlying the antispasmodic and bronchodilatory activities of the essential oil of <i>Artemisia maritima</i> L. Arch. Pharm. Res. 34: 1227-1238.
<i>Artemisia mutellina</i> Vill. (<i>A. umbelliformis</i> Lam.)	Compositae (Asteraceae)	Aerial part	Essential oil: bicyclic monoterpenes: e.g. alpha-thujone (57.7%), beta-thujone (6.6%)		Council of Europe. 2000. Natural sources of Flavours. Report No. 1. Council of Europe Publishing. ISBN 978-92-871-4324-2 Council of Europe. 2005. Active principles (constituents of chemical concern) contained in natural sources of Flavours. EC Council of Europe Publishing. http://www.coe.int/t/tacon/active/active20principles.pdf
<i>Artemisia pallens</i> DC.	Compositae (Asteraceae)	Aerial part	Essential oil: monoterpene etheroxide: 1,8-cineole; phenylpropanoids: e.g. methyl Eugenol.		Gadali and Khan. 1980. Essential oil of <i>Artemisia pallens</i> . Indian Perfum. 24: 101-109 Council of Europe. 2000. Natural sources of Flavours. Report No. 1. Council of Europe Publishing. ISBN 978-92-871-4324-2 Isidorov VA. 2001. Gas chromatographic analysis of essential oils with preliminary partition of components. Phytochem. Analysis 12: 87-90.
<i>Artemisia portica</i> L.	Compositae (Asteraceae)	Aerial part	Essential oil: bicyclic monoterpenes: e.g. alpha-thujone (13.5-30%), beta-thujone (3.3-4.2%), monoterpene etheroxide: 1,8-cineole (12-23%)		Council of Europe. 2000. Natural sources of Flavours. Report No. 1. Council of Europe Publishing. ISBN 978-92-871-4324-2
<i>Artemisia umbelliformis</i> L. See A. mutellina Vill.					
<i>Artemisia valisneria</i> Mill. (<i>A. valisneria</i> Lam., <i>Scorpioidium valisnerium</i> (Mill.) Sojak, <i>S. valisnerium</i> (Lam.) Y.R.Lindl)	Compositae (Asteraceae)	Aerial part	Essential oil: bicyclic monoterpene: e.g. camphor (33.3%); monoterpene etheroxide: 1,8-cineole (17%), phenylpropanoids: e.g. methylchavicol		Council of Europe. 2000. Natural sources of Flavours. Report No. 1. Council of Europe Publishing. ISBN 978-92-871-4324-2 Council of Europe. 2005. Active principles (constituents of chemical concern) contained in natural sources of Flavours. EC Council of Europe Publishing. http://www.coe.int/t/tacon/active/active20principles.pdf
<i>Artemisia vulgaris</i> L.	Compositae (Asteraceae)	Aerial part	Essential oil: bicyclic monoterpenes: e.g. alpha-thujone (56.3%), beta-thujone (7.5%), camphor (20%); monoterpene etheroxide: 1,8-cineole (26.8%)		Frome D., Parfiter H. J. and Anton R. 2005. <i>Plantes à risques</i> . Ed: Ter et Doc-Lavoisier. ISBN 978-2-7430-0907-1 Gib AL et al. 2008. A new iridoid alkaloid from <i>Azurdo donax</i> L. J. Asian Nat. Prod. Res. 10: 105-109.
<i>Arundo donax</i> L.	Poaceae (Gramineae)	Whole plant Rhizome	Genus in which species may contain oxalate raphides, glycosidic saponins (e.g. aronin), lignans, nec-lignans. Iridoid alkaloids: e.g. doxanin		Scharnberg BT et al. 2002. Determination of antispasmodic acid I and II in North American species of <i>Asarum</i> and <i>Aristolochia</i> . Pharmazie. 57(10): 686-689. PMID 12429949 EMEA Committee on Herbal Medicinal Products. 2005. Public statement on the use of herbal medicinal products containing asarone. EMEA/HMP/C/39219/2005 Dan Y et al. 2010. Activities of essential oils from <i>Asarum heterotropoides</i> var. <i>mandshuricum</i> against the phytopathogens. Crop Protection. 29(3): 295-299
<i>Asarum</i> spp.	Aristolochiaceae	Whole plant	Genus in which species may contain nitric phenanthrenic derivatives: e.g. aristolochic acids, aristolactams and phenylpropanoids: e.g. asarones, methyl Eugenol		Selye G et al. 1987. A. syriaca poisoning of cattle. Magyar. Allatorv. Lapja. 42(1): 56-58 Abe F and Yamachi T. 2000. An aristropane bi-oxide and 3-thiazolidinone derivatives of double-linked cardenolide glycosides from the roots of <i>Asclepias tuberosa</i> . Chem. Pharm. Bull. 48(7): 991-993.
<i>Asclepias syriaca</i> L. (<i>Asclepiadaceae</i>)	Asclepiadaceae	Rhizome	Cardenolide glycosides from latex: e.g. asclepin		Zhao GX et al. 1993. Biologically active acetogenins from stem bark of <i>Asimina triloba</i> . Phytochemistry. 33(9): 1065-1073. Geng-Xian Z et al. 1994. Asarin, asarinach, and asarinic: novel highly cytotoxic bisindole alkaloids from <i>Asimina triloba</i> . J. Med. Chem. 37(13): 1971-1976
<i>Asclepias tuberosa</i> L. (<i>Asclepiadaceae</i>)	Asclepiadaceae	Rhizome	Cardenolide glycosides from latex: e.g. asclepin		Council of Europe. 2000. Natural sources of Flavours. Report No. 1. Council of Europe Publishing. ISBN 978-92-871-4324-2.
<i>Asclepias vinetolokum</i> L. See <i>Vinetoctium nigrum</i> Moench.					
<i>Asimina triloba</i> (L.) Dur.	Annonaceae	Seed	Acetogenins: e.g. asinin, asininach, asininach		
<i>Aspidosperma quebracho-blanco</i> Schidl. (<i>Aspidiadaceae</i>)	Aspidiadaceae	Bark and wood	Iridoid alkaloids from bark (0.3-1.5%); e.g. aspidospermin (30%), quebrachine (yohimbine) (70%), decahydroaspidospermin (5%), aspidospermin (3%), aspidospermatidine (3%), 1-methylaspidospermatidine (0.5%), quebrachinine, quebrachidine		

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<i>Aspidosperma tomentosum</i> Mart.	Apocynaceae (Asclepiadaceae)	Bark and wood	Indole alkaloids: e.g. aspidospermine, quebrachine (yohimbine).	Swainsonine has been found in e.g. <i>A. leptoglossus</i> and <i>A. usitanicus</i> , but discussion is ongoing as to whether swainsonine is endogenously produced or by the endophyte <i>Embellisia</i> spp.	Gilberta B. et al. 1996. Alkaloid studies. The alkaloids of twelve aspidosperma species. Tetrahedron. 21(5). 1141-1166
<i>Astragalus</i> spp.	Leguminosae (Fabaceae)	Whole plant	Genus in which species may contain polyhydroxyindolizidine alkaloids: e.g. swainsonine, and may concentrate selenium	<i>A. trigonus</i> DC. and <i>A. gummifera</i> Labill. have a toxic effect on the central nervous system of livestock, leading to death. <i>A. lentiginosus</i> and <i>A. melisissus</i> have a toxic effect during pregnancy, leading to abortion and abnormalities in fetus cardiac function. Selenium concentrated by e.g. <i>A. hispidulus</i> .	Molyneux RJ, and James LF. 1992. Loco intoxication. Indolizidine Alkaloids of Spotted Locoweed (<i>Astragalus lentiginosus</i>). Science. 216(4542). 190-191 Baker DC et al. 1987. Selenium in developing pigs fed selenium from various sources. J Anim Sci. 65(suppl 1). 351 Bram K et al. 2003. Production of swainsonine by fungal endophytes of locoweed. Mycol Res. 107. 980-988. Ralpils H. et al. 2008. Relationships between the endophyte <i>Embellisia</i> spp. and the toxic alkaloid swainsonine in major locoweed species (<i>Astragalus</i> and <i>Oxytropis</i>). J. Chem. Ecol. 34:32-38
<i>Allyrium filix-femina</i> (L.) Roth	Woodsiaceae	Root and shoot		Thiaminase from fresh shoots	Schofield JJ. 2000. Discovering wild plants - Alaska, W. Canada and the Northwest Alaska Northwest books. ISBN 089240-365-9
<i>Attractylis gummifera</i> L.	Compositae (Asteraceae)	Root	Di terpene glycosides derived from kaurene: e.g. atractyliside, carboxyatractyliside, wedekeside		Georgiou ML. 1988. Hepatotoxicity due to <i>A. gummifera</i> L. Clin. Toxicol. 26(7). 487-493
<i>Atropa</i> spp.	Solanaceae	Whole plant	Genus in which species may contain tropane alkaloids: e.g. hyoscyamine, atropine, scopolamine.	The fresh plant contains L-hyoscyamine, the dried plant atropine (racemic mixture).	Frome D., Pfander H.J. and Anton R. 2009. Plantes à risques. Ed. Tec et Doc-Lavoisier, ISBN. 978-2-7430-0907-1
<i>Aucuba japonica</i> Thunb.	Garryaceae	Fruit		Causes fever and vomiting	Frome D., Pfander H.J. and Anton R. 2009. Plantes à risques. Ed. Tec et Doc-Lavoisier, ISBN. 978-2-7430-0907-1
<i>Azadirachta indica</i> A.Juss. (<i>Melia azadirachta</i> L.)	Melastaceae	Leaf and seed		The aqueous extract from the leaf, neem oil from the kernel, neem cake (the solid residue following the expulsion of the kernel oil) have all caused reduced fertility or caused infertility (e.g. by retarding spermiogenesis) in studies with male rats, mice, rabbits and guinea pigs. Oral administration of neem oil to female rats caused infertility or had abortive effect. Female contraceptive tablets from neem extracts are extensively used in India.	Kurose K and Yabagai M. 2005. Components of the essential oils of <i>Azadirachta indica</i> A. Juss. <i>Azadirachtin</i> <i>salutaris</i> , <i>Yellon</i> , and <i>Azadirachtin</i> <i>excelsa</i> (sadd), <i>lactos</i> and their comparison. J. Wood Sci. 51(2). 185-188. Moravil M et al. 2008. Sterility and abortive effects of the commercial neem (<i>Azadirachta indica</i> A. Juss.) extract Neem-kazal-TIS on female rat (<i>Rattus norvegicus</i>). Turk. J. Zool. 32. 155-162.
<i>Bambusa bambos</i> (L.) Voss (<i>Bambusa auriculata</i> (Retz.) Willd.)	Poaceae (Gramineae)	Shoot	Cyanogenic glycosides and derivatives: e.g. taxiphyllin		Hegnauer R. 1963. Chemotaxonomie der Pflanzen. Birkhauser Verlag Berlin. Vol. 2
<i>Bambusa vulgaris</i> Wendl.	Poaceae (Gramineae)	Shoot	Cyanogenic glycosides and derivatives: e.g. taxiphyllin (immature shoot tips: 8000 mg HC(NK/g)		Narley F. 1980. Toxicological aspects of cyanogenesis in tropical foodstuffs in Toxicology in the Tropics. Editors R.L. Smith and E.A. Bababurni, Taylor & Francis Ltd. London. 53-73.
<i>Banisteriopsis campi</i> (Spruce ex Graeb.) Morton	Malpighiaceae	Whole plant	Indole alkaloids (0.11-0.33%): e.g. harmine, harmaline.		Freeland CS et al. 1999. Behavioral profile of constituents in ayahuasca, an Amazonian psychoactive plant mixture. Drug Alcohol Depend. 54. 183-184 Calleway JC et al. 2005. Phytochemical analyses of <i>Banisteriopsis campi</i> and <i>Psychotria viridis</i> . J. Psychoactive Drugs 37(2). 145-150
<i>Baptisia</i> spp.	Leguminosae (Fabaceae)	Aerial part	Genus in which species may contain quinolizidine alkaloids: e.g. cystisine, N-methylcystisine and anagyriine		Cramer Wf and Turner Bl. 1967. Systematic significance of lupane alkaloids with particular reference to <i>Baptisia</i> (Leguminosae). Evolution. 21. 508-517
<i>Boerhaavia betulina</i> (Bergius) Bartl & H.L.Wendl. (<i>Agathosma betulina</i> (Bergius) Pilans)	Rutaceae	Leaf	Essential oil: monoterpene ketone: e.g. (S)-(-)-pulegone (3% - some chemotypes up to 70%).		Le-Baldin M et al. 2001. Buchu (<i>Agathosma betulina</i> and <i>A. crenulata</i> , Rutaceae) essential oils: their pharmacological action on guinea pig leuam and antimicrobial activity on microorganisms. J. Pharm. Pharmacol. 53(4). 579-582 Gordon, W. P. et al. 1982. Hepatotoxicity and pulmonary toxicity of pennyroyal oil and its constituent terpenes in the mouse. Toxicol. Appl. Pharmacol. 65. 415-424 EC Scientific Committee on Food. 2002. Opinion of the Scientific Committee on Food on pulegone and menthofuran. SCH/CS/FAN/LAV/VOU/03/AD02
<i>Belamcanda punctata</i> Moench (<i>B. chinensis</i> (L.) DC.)	Indicaceae	Root	1,4-benzoxquinone derivatives: e.g. belamcandaquinones A and B Methylated isoflavones: e.g. lectodrigemlin, ingenin, belamcandain		Yanaki, M et al. 1990. Isoflavones of <i>Belamcanda chinensis</i> . Phana Medica 56(3): 335. Fukuyma, Y et al. 1993. Belamcandaquinones A and B. novel dimeric 1,4-benzoxquinone derivatives possessing cyclooxygenase inhibitory activity. Tetrahedron Letters 34(47). 7653-7656
<i>Berberis vulgaris</i> L.	Berberidaceae	Root	Isoquinoline alkaloids: e.g. berberine (0.5 - 6%), palmatine, jatrorrhizine, and bisbenzyltetrahydroisoquinoline alkaloids: e.g. berbamine, oxycanthine, isoberberine.		Szau R et al. 1988. Isoquinoline alkaloids from <i>Berberis vulgaris</i> subsp. australis. Phytochemistry. 49(8). 2545-2549 Frome D., Pfander H.J. and Anton R. 2009. Plantes à risques. Ed. Tec et Doc-Lavoisier, ISBN. 978-2-7430-0907-1
<i>Boerhaavia diffusa</i> L.	Nyctaginaceae	Whole plant	Alkaloids: pumamine Rolenoids: e.g. boerhavinones		Wright M. and Anton R. 2003. Plantes thérapeutiques. Ed. Tec et Doc-Lavoisier. ISBN. 2-7430-0831-5 (2ème édition)
<i>Borago</i> spp.	Borraginaceae	Aerial part	Genus in which species may contain unsaturated pyrrolizidine alkaloids: e.g. lycopsamine, 7-acetyl-lycopsamine, amarillin, supinine.		Manu KA et al. 2009. Anti-neoplastic potential of Punarnawa, an alkaloid from <i>Boerhaavia diffusa</i> Linn. Immunobiology. 2009;214(4):24-255. Ahmed Bekacem A et al. Non-pyruvated rotenoids, a new class of potent breast cancer resistance protein inhibitors. J Med Chem. 2007. 50(8):1933-1938.
<i>Boswellia terrestris</i> Birdw.	Burseraceae	Bark	Essential oil from the gum resin: bicyclic monoterpenes: e.g. beta-thujone and phenylpropanoids: e.g. methyl Eugenol		Chroliker M. 2003. Hepatic sinusoidal obstruction syndrome: toxicity of pyrrolizidine alkaloids. Hepatol. 39. 437-446 Bruneton J. 2009. Pharmacognosie, (phytochimie, Plantes médicinales). Ed. Tec et Doc-Lavoisier, Paris. 4ème édition. ISBN. 978-2-7430-1188-8 Hamm S et al. 2005. A chemical investigation by headspace-SPME and GC-MS of volatile and semi-volatile terpenes in various oilseam samples. Phytochemistry. 66(12). 1499-1514.

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<i>Boswellia serrata</i> Hook.	Burseraceae	Bark	Essential oil from the gum resin; phenylpropanoids (up to 11%); e.g. methylchavicol.		Burton J. 2009. Pharmacognosy. (Phytochemistry, Plantae mediferae). Ed. Tec & Doc Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1188-8. Went M. and Anton R. 2003. Plantae heteroquidales. Ed. Tec & Doc Lavoisier, ISBN: 2-7430-0631-5 (2ème édition). Hamm S. et al. 2005. A chemical investigation by headspace SPME and GC-MS of volatile and semi-volatile terpenes in various olibanum samples. Phytochemistry, 66(12):1499-1514.
<i>Brachyglottis</i> spp.	Compositae (Asteraceae)	Leaf	Genus in which species may contain unsaturated pyrrolizidine alkaloids; e.g. senecionine.		Mortimer PH and White EP. 1967. Hepatotoxic substance in <i>Brachyglottis repanda</i> . Nature, 214, 1255-1256.
<i>Bregantia</i> spp.	Aristolochiaceae	Root	Genus in which species may contain isocoumarin alkaloids; e.g. chankarine, and irifric phenanthrene derivatives; e.g. aristolochic acids.	<i>Bregantia</i> sometimes falsified with <i>Aristolochia</i>	Karnal VN et al. 1956. Studies on Indian medicinal plants I. Characterization of chankarine, an alkaloid isolated from <i>Bregantia wallichii</i> R. Br. (n. o. Aristolochiaceae). Indian J Med Res. 1956. May;46(9):1418-25.
<i>Brassicica nigra</i> (L.) W.D.J.Koch	Brassicaceae (Cucitaceae)	Aerial part	Glucosinolates (especially in the seed); e.g. sinigrasin (= allylglucosinolate) (1-2%), allylthiocyanate and derivatives; e.g. gluconaprine, glucosulfurine, gluco-sulpherine.	Health Canada advises consumers not to use the products containing aristolochic acid. Ottawa, Health Canada, 2004 July 26. www.hc-sc.gc.ca/english/protectio/protectio/warnings/2004/2004_433.htm	Burton J. 2009. Pharmacognosy. (Phytochemistry, Plantae mediferae). Ed. Tec & Doc Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1188-8. Halkier BA and Gershenzon J. 2006. Biology and Biochemistry of Glucosinolates. Annual Review of Plant Biology, 57, 303-333.
<i>Bryera anthelmintica</i> Kunth. (<i>Higenia abyssinica</i> J.F. Gmel.)	Rosaceae	Flower	Phenolglycosid derivatives: koslochin, protokoshin, koshin (α-β-)	Visual deficits and retinotoxicity observed in humans	Low et al. 1985. Visual deficits and retinotoxicity caused by the naturally occurring anthelmintics, Embelia ribs and <i>Higenia abyssinica</i> . Toxicol. Appl. Pharm. 81(2): 220-230. Singh IP and Bhatnagar SB. 2006. Phenolglycosid compounds of natural origin. Nat. Prod. Rep. 23, 558-591.
<i>Buceca javanica</i> (L.) Merr.	Simarubaceae	Bark	Quassinoids (nortriterpenoids); e.g. buceanine.		Burton J. 2009. Pharmacognosy. (Phytochemistry, Plantae mediferae). Ed. Tec & Doc Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1188-8.
<i>Bugmansia</i> spp.	Solanaceae	Aerial part	Genus in which species may contain tropane alkaloids; e.g. scopolamine.		Frome D, Pfander HJ and Anton R. 2009. Plantae à risques. Ed. Tec & Doc Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-0907-1.
<i>Buruleia</i> spp.	Scitaceae	Root	Genus in which species may contain indole alkaloids (beta-carboline derivatives); e.g. harmine, tetrahydroharmine, harmaline, manachine, manacine, dimethyltryptamine derivatives and amides; e.g. pyrrole-3-acetoxamide.		Frome D, Pfander HJ and Anton R. 2009. Plantae à risques. Ed. Tec & Doc Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-0907-1.
<i>Bryonia</i> spp.	Cucurbitaceae	Whole plant	Genus in which species may contain oxygenated tetracyclic, triterpene derivatives; e.g. cucurbitacins		Frome D, Pfander HJ and Anton R. 2009. Plantae à risques. Ed. Tec & Doc Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-0907-1.
<i>Butea superba</i> Hook.	Leguminosae (Fabaceae)	Root	Steroidal alkaloids with amine groups; e.g. butine, cyclobutene, butamine, and tripenoid alkaloids; e.g. diacetylbuxadine, demethylcycloalkuranine.	Oral dosing with dried tuber powder for 90 days reduced testosterone levels in male rats dosed with 10, 150 and 200 mg/kg b.w. Effects on uterizing hormone level found in male orchidectomized rats and ovariectomized female rats.	Cherdswasart W et al. 2008. Androgen disruption and toxicity tests of <i>Butea superba</i> Roxb. A traditional herb used for treatment of erectile dysfunction in male rats. Maturitas 60, 131-137. Cherdswasart W et al. 2010. Mulegic and antitumor effects of the traditional herb used for treating erectile dysfunction. <i>Butea superba</i> Roxb. Biosci Biotechnol Biochem 74, 923-927. Matsuyama S et al. 2009. Androgenic activity of the Thai traditional male potency herb, <i>Butea superba</i> Roxb., in female rats. J Ethnopharmacol 121, 123-129. Matsuyama S et al. 2010. Uterizing hormone reduction by the male potency herb, <i>Butea superba</i> Roxb. Braz. J Med Biol Res. 43, 843-852.
<i>Buxus sempervirens</i> L.	Buxaceae	Whole plant	Steroidal alkaloids with amine groups; e.g. butine, cyclobutene, butamine, and tripenoid alkaloids; e.g. diacetylbuxadine, demethylcycloalkuranine.		Alta-ur R et al. 1999. New steroidal alkaloids from the roots of <i>Buxus sempervirens</i> . J. Nat. Prod. 62(5), 665-669. Ala A et al. 2002. New triterpenoid alkaloids from <i>Buxus sempervirens</i> . Z. Naturforsch. 57c, 21-28.
<i>Caladium</i> spp.	Araceae	Whole plant	Genus in which species may contain calcium oxalates		Frome D, Pfander HJ and Anton R. 2009. Plantae à risques. Ed. Tec & Doc Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-0907-1.
<i>Calamintha asperidens</i> Jord. See <i>Chilopodium menthifolium</i> spp.					
<i>Calceolae</i> (L.) Govaerts	Compositae (Asteraceae)	Leaf	Sesquiterpene germacranolides; e.g. calcidine, lurenol esters		Mayrgruber L et al. 1986. Psychopharmacologic analysis of an alleged onerogenic plant. <i>Calceolae zaezelochii</i> . J. Ethnopharmacol. 18(3), 228-243.
<i>Calendula officinalis</i> L.	Compositae (Asteraceae)	Flower	Calcium oxalate raphides	Hydro-alcoholic extract (1g/kg during 30 days in the rat): increase of urea and transaminases. Hydro-alcoholic extract did not affect male fertility nor had toxic effects in early and middle periods of pregnancy. However, the extract caused maternal toxicity when administered during the fetal period of pregnancy.	Silva EJ and Gonçalves ES. 2007. Toxicological studies on hydroalcoholic extract of <i>Calendula officinalis</i> L. Phytother. Res. 21, 332-336. Silva EJ et al. 2008. Reproductive assessment of hydroalcoholic extract of <i>Calendula officinalis</i> L. in Wistar rats. Phytother Res. 23(10), 1352-1358
<i>Callia palustris</i> L.	Araceae	Whole plant	Calcium oxalate raphides		Almeida KF and McCam JM. 1995. AMA Handbook of poisonous and injurious plants. American Medical Assoc. Chicago, IL, USA.
<i>Calotropis</i> spp.	Apocynaceae	Whole plant	Genus in which species may contain cardenolide glycosides and steroidal components; e.g. pregnanone		Barreiro AF et al. 2005. Chemical composition of the essential oils of leaves and wood <i>Calotropis articulata</i> (Vahl) Masters. Journal of Essential Oil Research. 17(12), 3033-3039
<i>Calycanthus floridus</i> L.	Ranunculaceae	Whole plant	Lactones; e.g. protoanemonin	Protoanemonin only present in the fresh herb	Bruni A et al. 1986. Protoanemonin detection in <i>C. palustris</i> J. Nat. Prod. 49(6), 1172-1173.
<i>Calycanthus floridus</i> L.	Calycanthaceae	Bark	Bisbenzylisoquinoline alkaloids; e.g. calycanthine		Akhteghi H. 2008. Chemical composition of the essential oil from stems of <i>Calycanthus floridus</i> L. var. <i>oblongifolius</i> from Iran. Chem. Nat. Compd. 44(6), 661-662.
<i>Calystegia sepium</i> R.Br.	Convolvulaceae	Whole plant	Polyhydroxy-noropane alkaloids; e.g. calystegines (5-316 mg/kg in the dried plant); jalapine like cardiac glycosides (mainly in the root)		Shoif Y et al. 2001. Calystegines in <i>Calystegia sepium</i> derive from the tropane alkaloid pathway. Phytochemistry, 56(6), 883-889. Brunton J. 2009. Pharmacognosy. (Phytochemistry, Plantae mediferae). Ed. Tec & Doc Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1188-8

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Botanical name	Family	Parts of plants of possible concern	Chemical of concern	Remarks on toxic/adverse effect(s) not known to be related to the identified chemical(s) of concern	Specific References
<i>Cannella sasanqua</i> Thunb.	Theaceae	Seed	Sasanqua thieropoid saponins		Shen J et al. 2008. Evidence of gastro-intestinal system as an active and toxic target of Sasanqua saponins extract. Exp. Toxicol. Pathol. 60(1) 43-49
<i>Cannella sinensis</i> (L.) Kunze (<i>Thea sinensis</i> L.)	Theaceae	Leaf	Methylated xanthine derivatives: caffeine (2-4%), theophylline (traces) and catechins: e.g. epigallocatechingallate (6-12%)	Reported cases of hepatotoxicity (green tea)	Borkovec HL. 2006. Hepatotoxicity associated with supplements containing Chinese green tea (<i>Cannella sinensis</i>). Ann Intern Med. 144(1), 66-71. Eur J Clin Pharmacol. 2006; Mar 7; 144(3):380 EFSA Scientific Cooperation (ESCO) Working Group on Botanicals and Botanical Preparations 2009. Advice on the EFSA guidance document for the safety assessment of botanicals and botanical preparations intended for use as food supplements. Based on real case studies on request of EFSA. EFSA Journal 19(2):280
<i>Caragana odorata</i> (Lam.) Hook.f. & Thoms.	Annoaceae	Aerial part	Essential oil: phenylpropanoids: e.g. safrole, isosafrole		Council of Europe. 2008. Natural sources of flavonoids. Report No. 3. Council of Europe Publishing. ISBN 978-92-871-6442-3. SCF. 2001. Opinion on the safety of the presence of safrole (1-(3H)-1,3-dimethyl-4H-benzene) in flavourings and other food ingredients with flavouring properties. available at: http://ec.europa.eu/food/fs/sc/S200116_n.pdf
<i>Canarium indicum</i> L. (<i>Canarium commune</i> L.)	Bursaceae	Bark of the trunk			Canarium luzonicum (Miq.) A. Gray or Manila elemi : oleoresin gives rise to essential oil which contains elemene (0.5-8%), elemol (1-15%) (norme NF-ISO 10624-1989). Bruleton J. 2009. Pharmacoposie, (Phytochimie, Plantes médicinales). Ed. Tec & Doc Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1188-8
<i>Cannabis</i> spp.	Cannabaceae	Flowering top (female)	Genus in which species may contain cannabinoids (terpenophenols); e.g. tetrahydrocannabinol		Frome D., Pfander H.J. and Anton R. 2009. Plantes à risques. Ed. Tec et Doc Lavoisier, ISBN: 978-2-7430-0907-1 Bruleton J. 2009. Pharmacoposie, (Phytochimie, Plantes médicinales). Ed. Tec & Doc Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1188-8
<i>Capsella bursa-pastoris</i> (L.) Medik.	Brassicaceae (Cuciferae)	Aerial part	Phenylethanols: e.g. tyramine oxalates		Bruleton J. 2009. Pharmacoposie, (Phytochimie, Plantes médicinales). Ed. Tec & Doc Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1188-8
<i>Caragana arborescens</i> Lam.	Leguminosae (Fabaceae)	Seed	Lectins		Bicot R et al. 1999. Purification and characterization of two lectins from Caragana arborescens seeds. Molecules. 116 (4), 48-51
<i>Carapichea jpecacuarina</i> (Brot.) L. Andersson See <i>Cephaelis</i> spp.					
<i>Cardiospermum halicacabum</i> L.	Sapindaceae	Leaf and seed	Cyanogenic glycosides from the leaf	Traces of alkaloids in the seed, although no information is available on their nature.	EMEA Committee for Veterinary Medicinal Products. 1999. Cardiospermum halicacabum - Summary report. EMEA/MRL/664/99-Final Ragupathy S et al. 2007. Exploring ethnobiological classifications for novel alternative medicine: a case study of Cardiospermum halicacabum L. (Madakakalm, baloon vine) as a traditional herb for treating rheumatoid arthritis. Ethnobotany. 19. 1-16
<i>Carum carvi</i> L.	Apiaceae (Umbriferae)	Fruit	Essential oil: monoterpene ketone: e.g. (S)-(+)-carvone (50-65%)		Council of Europe. 2000. Natural sources of flavonoids. Report No. 1. Council of Europe Publishing. ISBN: 978-92-871-4324-2
<i>Carophyllus aromaticus</i> L. See <i>Syzgium aromaticum</i> (L.) Merr. & L.M. DC.					
<i>Caryota</i> spp.	Arecaceae (Palmae)	Whole plant	Genus in which species may contain cyanogenic glycosides in the leaf and oxalate raphides		Frome D., Pfander H.J. and Anton R. 2009. Plantes à risques. Ed. Tec et Doc Lavoisier, ISBN: 978-2-7430-0907-1
<i>Casuarina edulis</i> Llave & Lex	Fabaceae	Leaf and seed	Imidazole alkaloids: e.g. casuarinidine		Pavia GM. 2003. Biochemical targets of plant bioactive compounds. CRC Press. Demille J. and Demeyer K. 2010. Anthraquinones in plants. Source, safety and applications in gastrointestinal health. Nottingham University Press. ISBN: 978-1-897676-32-5
<i>Cassia</i> spp.	Leguminosae (Fabaceae)	Aerial part	Genus in which species may contain hydroxyanthracene glycosides and derivatives (1,8-dihydroxyanthraquinones).		Bruleton J. 2009. Pharmacoposie, (Phytochimie, Plantes médicinales). Ed. Tec & Doc Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1188-8
<i>Castanea sativa</i> Mill.	Fagaceae	Aerial part		Hydrolysable tannins: e.g. ellagitanins used at high doses and over a long period may have a negative impact on liver	Bruleton J. 2009. Pharmacoposie, (Phytochimie, Plantes médicinales). Ed. Tec & Doc Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1188-8
<i>Catha edulis</i> (Vahl) Forsk. ex Endl.	Celastraceae	Leaf	Phenethylamines: e.g. (-)-cathine (fresh and young leaf) ; norepinephrine (cathine) and norepinephrine (dried and/or old leaf)		Al-Masary M et al. 2002. Investigation into the toxicological effects of <i>Catha edulis</i> leaves: a short term study in animals. Phytother. Res. 16(2), 127-132.
<i>Catharantus</i> spp.	Apocynaceae	Whole plant	Genus in which species may contain iridoid alkaloids: e.g. vindoline, catharanthine (monochodols), vindosifine, vindosidine, leucosifine (bis-indoles), sinalbinine, akamamine (dihydro-indoles)		Bruleton J. 2009. Pharmacoposie, (Phytochimie, Plantes médicinales). Ed. Tec & Doc Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1188-8
<i>Caulophyllum thalictroides</i> (L.) Michx. (<i>Lonicera thalictroides</i> L.)	Berberidaceae	Whole plant	Quinoline alkaloids: e.g. cystine, bapifoline and N-methylcystisine in leaf and fruit		Rao RB et al. 2002. Nicotinic toxicity from fracture of blue chestnut (<i>C. thalictroides</i>) used as an abortifacient. Vet. Hum. Toxicol. 44(4), 221-222 Frome D. and Pfander H.J. 1997. Giftblanzen. Ein Handbuch für Apotheker, Ärzte, Toxikologen und Biologen. Wissenschaftliche Verlagsgesellschaft mbH. ISBN: 3-9047-1468-8
<i>Cedrus</i> spp.	Pinaceae	Aerial part	Genus in which species may contain bicyclic monoterpenes: e.g. thujones in the essential oil		Council of Europe. 2005. Active principles, constituents of medicinal compound contained in natural sources of flavonoids. Ed. Council of Europe Publishing http://www.coe.int/t/cesocai/consensus/consolidable_health/Flavonoids_substances_Aktiv%20pflanzliche.pdf
<i>Cephaelis</i> spp.	Rubiaceae	Root	Genus in which species may contain isouquinoline monoterpene alkaloids (2,0 - 3,5%), e.g. emetine, cephaeline, psychotrine, anetraniline, and dioxycorins: e.g. lobosidine		Bruleton J. 2009. Pharmacoposie, (Phytochimie, Plantes médicinales). Ed. Tec & Doc Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1188-8
<i>Cestrum</i> spp.	Solanaceae	Whole plant	Genus in which species may contain diterpene glycosides: e.g. parquine, carboxyparquine, and steroid glycosides: e.g. 1,25-dihydroxycycloalcaloiden, salsosidine.		Dunand R et al. 1999. Intoxication in cattle from <i>C. diurnum</i> . Vet. Hum. Toxicol. 41(1), 26-27
<i>Ceraria islandica</i> (L.) Ach.	Parmeliaceae	Lichen	Dibenzofuran derivatives: e.g. usnic acid	<i>C. islandica</i> reported to concentrate heavy metals	Bruleton J. 2009. Pharmacoposie, (Phytochimie, Plantes médicinales). Ed. Tec & Doc Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1188-8 Went M. and Anton R. 2003. Plantes Hépatoprotectrices. Ed. Tec et Doc Lavoisier, ISBN: 2-7430-0631-5 (2ème édition) Araškisen MM et al. 1986. Toxicity of Ireland lichen and reindeer lichen. Arch. Toxicol. Suppl. 9:406-408
<i>Chaenomeles speciosa</i> Nakai	Rosaceae	Seed	Cyanogenic glycosides		Frome D., Pfander H.J. and Anton R. 2009. Plantes à risques. Ed. Tec et Doc Lavoisier, ISBN: 978-2-7430-0907-1

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<i>Chamaefilum ulinum</i> (L.) A. Gray	Meliastaceae	Whole plant	Steroidal saponins: e.g. chamaefilin (glucoside of diosgenin), heliosides A and B, calcium oxalate		Chandler VL et al. 2011. Structure and absolute configuration of heliosides A and B, new saponins from <i>Chamaefilum ulinum</i> . J. Nat. Prod. 74(7):1557-1560. Molovic NJ et al. 2011. The truth about false unicorn (<i>Chamaefilum ulinum</i>): total synthesis of 23R,24S-chrysoarigenol B defines the structure and stereochemistry of the major saponin from this medicinal herb. Chemistry, 17(27):7578-7591. Penyol P et al. 2011. Appalachian plant monographs: <i>Chamaefilum ulinum</i> (L.) Gray. False Unicorn root. Retrieved from http://www.frostburg.edu/arc/appalachian-plants/
<i>Cheiranthus cheiri</i> L.	Brassicaceae (Cruciferae)	Aerial part	Cardenolides: e.g. cheiranthin (strophanthidin derivative)		Frome D., Pfander H.J. and Anton R. 2009. <i>Plantes à risques</i> . Ed. Tec et Doc-Lavoisier, ISBN: 978-2-7430-0907-1 Lai ZH et al. 2002. Cardiac glycosides from <i>Erysimum cheiranthoides</i> . Chem Pharm Bull (Tokyo) 50(6):981-982
<i>Chelidonium majus</i> L. (<i>Chelidonium umbelliferum</i> Stokes)	Papaveraceae	Whole plant	Benzophenanthridine alkaloids (2% in root), e.g. chelidonic, chelerythrine, sanguinarine, protopine, and protoberberine derivatives: e.g. berberine, stylopine, coptisine		Gu Y et al. 2010. Simultaneous determination of seven main alkaloids of <i>Chelidonium majus</i> L. by ultra-performance LC with photodiode-array detection. J. Sep. Sci. 33(8), 1004-1009. Morio PA et al. 2009. Hepatitis from Greater celandine (<i>Chelidonium majus</i> L.): review of literature and report of a new case. J. Ethnopharmacol. 124(2):328-332
<i>Chenopodium album</i> L.	Amaranthaceae (Chenopodiaceae)	Leaf	Essential oil: peroxylated monoterpane: ascaridole (45%)		Brurleton J. 2009. Pharmacognosy, (Phytochimie, Plantes médicinales), Ed. Tec & Doc-Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1188-8 Frome D, Pfander HJ and Anton R. 2009. <i>Plantes à risques</i> , Ed. Tec et Doc-Lavoisier, ISBN: 978-2-7430-0907-1
<i>Chenopodium ambrosioides</i> L. var. <i>anthemifolium</i> (L.) A. Gray (<i>Chenopodium ambrosoides</i> L.)	Amaranthaceae (Chenopodiaceae)	Aerial part	Essential oil: peroxylated monoterpane: ascaridole		Brurleton J. 2009. Pharmacognosy, (Phytochimie, Plantes médicinales), Ed. Tec & Doc-Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1188-8 Frome D, Pfander HJ and Anton R. 2009. <i>Plantes à risques</i> , Ed. Tec et Doc-Lavoisier, ISBN: 978-2-7430-0907-1
<i>Chondrodendron</i> spp.	Menispermaceae	Whole plant	Genus in which species may contain quaternary bis-aminonium isocoumarin alkaloids: e.g. (+)-tubocurarine, and tertiary alkaloids: e.g. (-)-curine, (+)-isochondrodendrine, (+)-chondrocucurine, and tertiary bisbenzylisocoumarin: e.g. lincaline, linacouline		Brurleton J. 2009. Pharmacognosy, (Phytochimie, Plantes médicinales), Ed. Tec & Doc-Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1188-8
<i>Chrysanthemum cheiranthifolium</i> (Trew.) Vs. <i>See Tanacetum cheiranthifolium</i> (Trew.) S.M.B.P.	Compositae (Asteraceae)	Flower	Essential oil: monoterpane etheroxide: 1,8-cineole and bicyclic monoterpane: camphor.		Shunqing Z et al. 2005. Chemical composition and antimicrobial activity of the essential oils of <i>Chrysanthemum indicum</i> . J. Ethnopharmacol. 96(1-2): 151-158
<i>Chrysanthemum indicum</i> L.	Compositae (Asteraceae)	Flower	Essential oil: monoterpane etheroxide: 1,8-cineole and bicyclic monoterpane: camphor.		Brurleton J. 2005. <i>Plantes toxiques (Végétaux dangereux pour l'homme et les animaux)</i> , Ed. Tec & Doc-Lavoisier, Paris, 3ème édition, ISBN: 2-7430-0806-7
<i>Chrysanthemum leucanthemum</i> L. <i>See Leucanthemum vulgare</i> Lam. <i>Chrysanthemum vulgare</i> (L.) Bernh. <i>See Tanacetum vulgare</i> L.					Barnes J, Anderson L. A., Phillips J, David 2007. <i>Herbal Medicines Third edition</i> ISBN 978 0 85369 623 0. EMA HMP-C-2010. Assessment report on <i>Chimifuga racemosa</i> (L.) Nutt., nitrome. EMA/HMP/C/396/8/2008
<i>Cnicus</i> spp.	Apiaceae (Umbelliferae)	Whole plant	Genus in which species may contain polyynes: e.g. (-)-cnicolxin		Brurleton J. 2009. Pharmacognosy, (Phytochimie, Plantes médicinales), Ed. Tec & Doc-Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1188-8
<i>Cnicifluga racemosa</i> (L.) Nutt. (<i>Cnicifluga serpentina</i> Pursh, <i>Adiantum racemosa</i> L.)	Ranunculaceae	Whole plant	Genus in which species may contain quinine alkaloids: e.g. quinine, quinidine, cinchonine, cinchonidine	Herb under scrutiny for hepatotoxicity	Brurleton J. 2009. Pharmacognosy, (Phytochimie, Plantes médicinales), Ed. Tec & Doc-Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1188-8 Council of Europe. 2008. <i>Natural sources of flavonoids</i> . Report No. 3. Council of Europe Publishing, ISBN 978-92-871-8422-3
<i>Cniciona</i> spp.	Rubiaceae	Bark	Genus in which species may contain quinine alkaloids: e.g. quinine, quinidine, cinchonine, cinchonidine		Teuscher E., Anton R. and Lobstein A. 2005. <i>Plantes aromatiques</i> , Ed. Tec et Doc-Lavoisier, ISBN: 2-7430-0720-6 Abraham K. et al. 2011. Relative bioavailability of coumarin from chironom and chironom-containing foods compared to isolated coumarin: a four-way crossover study in human volunteers. Mol. Nutr. Food Res. 55(4):644-653. Woytinn F. et al. 2010. Quantification of flavonoid constituents in chironom: high variation of coumarin in cassa bark from the German retail market and in authentic samples from Indonesia. J. Agric. Food Chem. 58(19):10588-10575
<i>Chenamonium camphora</i> (L.) J.Presl.	Lauraceae	Wood	Bicyclic monoterpane: camphor; monoterpane etheroxide: 1,8-cineole; phenylpropanoid: safrole		EMA Committee on Herbal Medicinal Products (CHMP). Public Statement on the use of herbal medicinal products containing methyl Eugenol. Doc.ref. EMA/HMP/MC/138363/2005
<i>Chenamonium cassia</i> (Nees) Blume (<i>Chenamonium aromaticum</i> Nees)	Lauraceae	Aerial part	Essential oil from the bark (20ml/kg): coumarin (1.5-4.0 g/kg) Essential oil from the leaf and young stem: coumarin (1.5-4%)		EMA Committee on Herbal Medicinal Products (CHMP). Public Statement on the use of herbal medicinal products containing methyl Eugenol. Doc.ref. EMA/HMP/MC/138363/2005
<i>Chenamonium platyphyllum</i> (Diels) C.K. Allen	Lauraceae	Aerial part	Reported to contain the phenylpropanoid methyl Eugenol in unspecified quantities		EMA Committee on Herbal Medicinal Products (CHMP). Public Statement on the use of herbal medicinal products containing methyl Eugenol. Doc.ref. EMA/HMP/MC/138363/2005
<i>Chenamonium rigidissimum</i> H.T.Chiang	Lauraceae	Wood	Essential oil: phenylpropanoids: e.g. safrole (61.72%), methyl Eugenol (28.62%)		EMA Committee on Herbal Medicinal Products (CHMP). Public Statement on the use of herbal medicinal products containing methyl Eugenol. Doc.ref. EMA/HMP/MC/138363/2005
<i>Chenamonium septentrionale</i> Hand-Mazz.	Lauraceae	Unspecified	Phenylpropanoids: e.g. methyl Eugenol		EMA Committee on Herbal Medicinal Products (CHMP). Public Statement on the use of herbal medicinal products containing methyl Eugenol. Doc.ref. EMA/HMP/MC/138363/2005
<i>Chenamonium verum</i> J.Presl. (<i>Chenamonium zeylanicum</i> Blume, C zeylanicum Nees)	Lauraceae	Aerial part	Essential oil from the bark (0.5-1.3%): monoterpane etheroxide: 1,8-cineole (<3%), bicyclic monoterpenes: e.g. camphor (traces), phenylpropanoids: e.g. chamanaldehyde (62%) and safrole (<0.5%), methyl Eugenol (traces), coumarin (<0.5%) Essential oil from the leaf: 1,8-cineole (<1%), safrole (<3%), coumarin (<1%), methyl Eugenol (0.01%)		EMA Committee on Herbal Medicinal Products (CHMP). Public Statement on the use of herbal medicinal products containing methyl Eugenol. Doc.ref. EMA/HMP/MC/138363/2005
<i>Cissampelos pareira</i> L. (<i>Cocculus orbiculatus</i> DC.)	Menispermaceae	Root and stem	Isocoumarin alkaloids: e.g. havoline, havylidine, tropilobisquinoline alkaloids: e.g. pareinoline A and B Bisbenzylisocoumarin alkaloids in the stem: e.g. coccolobicalanine A, 10-hydroxyisochlidiolone		Bařna, A. et al. 2009. Antibiotic and immunomodulatory activity of the alkaloidal fraction of <i>Cissampelos pareira</i> Linn. Sci Pharm. 78(1):21-31 Amresh, G. et al. 2008. Toxicological screening of traditional medicine <i>Laghubatha</i> (<i>Cissampelos pareira</i>) in experimental animals. J. Ethnopharmacol. 116(3):454-460. Ganguly, M. et al. 2007. Antifeedant activity of the methanolic leaf extract of <i>Cissampelos pareira</i> in female albino mice. J. Ethnopharmacol. 111(3):688-691

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<i>Citrus adami</i> L. (<i>C. viscosus</i> Stokes, <i>C. grandiflorus</i> Pour., <i>C. limonaria</i> Hoffmanns. & Adamicus L., <i>Ladurium officinarum</i> Sieber)	Citracaceae	Leaf and twig	Essential oil: bicyclic monoterpenes e.g. alpha-thujone (0.8%), monoterpene ether oxide: 1,8-sinene (0.2%)	Inflammation of gastrointestinal tract with bloody diarrhoea described, toxic compounds not known. Cucurbitacins in plant material: low content in young leaves, 1-3 g/kg in old leaves and stems.	Barceloux DG. 2008. Medical botany of natural substances: foods, fungi, medicinal herbs, plants and venomous animals. John Wiley & sons, Hoboken, New Jersey. ISBN-10: 0-471-72761-X Gryll J., Søborg I. and Andersson H.C. 2006. Cucurbitacins in plant food. <i>Tema Nord</i> , 556. Nordic Council of Ministers. ISBN: 92-893-1381-1
<i>Citrus colocythis</i> (L.) Schrad. (<i>Cucumis colocythis</i> L.)	Cucurbitaceae	Fruit	Oxygenated tetracyclic terpenes; e.g. cucurbitacins.		Council of Europe. 2000. Natural sources of Flavonoids. Report No. 1. Council of Europe Publishing. ISBN 978-92-871-4324-7
<i>Citrus aurantium</i> L. (<i>C. aurantium</i> L. ssp. <i>amara</i> Engelm., <i>C. aurantium</i> L. ssp. <i>sinensis</i> L., <i>C. aurantium</i> L. ssp. <i>aurantium</i> L., <i>C. aurantium</i> var. <i>diclos</i> <i>Citrus aurantium</i> var. <i>Bergamota</i>)	Rutaceae	Aerial part	Essential oil: furanocoumarins; e.g. 5-methoxypsoralen (0.15-0.81%), Urtripe whole fruit: hydroxyphenylethylamine: synephrine (2.28 mg/g) Ptercap: synephrine (3.27 mg/g).		Council of Europe. 2000. Natural sources of Flavonoids. Report No. 1. Council of Europe Publishing. ISBN 978-92-871-4324-7 EFSA ESCO Working Group on Botanical Preparations. 2009. Advice on the EFSA guidance document for the safety assessment of botanicals and botanical preparations intended for use as food supplements, based on real case studies. The EFSA Journal. 7(9): 280. Teuscher E., Anton R. and Lobstein A. 2005. <i>Plantes aromatiques</i> . Ed. Tec et Doc-Lavoisier. ISBN: 2-7430-0720-6
<i>Citrus limon</i> (L.) Burm.f. (<i>Citrus medica</i> var. <i>limon</i> L., <i>Citrus limonum</i> Risso)	Rutaceae	Fruit, leaf, peel and pulp	Peel: Phenylethan, 5- and 8-geranoxypsoralen, Essential oil from the peel: furanocoumarins (psoralen, 5-methoxypsoralen (bergapten) 4-87 mg/kg, 8-methoxypsoralen (xanthoxon), 5,8-dimethoxypsoralen (isopimpinellin), imperatorin, oxypseudocoumarin 26-728 mg/kg		WICHTI M. and Anton R. 2003. <i>Plantes thérapeutiques</i> (Tradition, pratique, officinale, science et thérapeutique). Ed. Tec & Doc-Lavoisier, Paris, 26ème édition, 692 pages, ISBN: 2-7430-0631-5 Council of Europe. 2000. Natural sources of Flavonoids. Report No. 1. Council of Europe Publishing. ISBN 978-92-871-4324-2 Benincasa M. et al. 1990. Analysis of lemon and bergamot essential oils by HPLC with microbore columns. <i>Chromatographia</i> 30(6):271-6 Wagstaff D. J. 1991. Dietary exposure to furanocoumarins. <i>Regul. Toxic Pharmacol.</i> 14:261-272 Kulkarni TR. et al. 2005. Study of anti-fertility effect of lemon seeds (<i>Citrus limonum</i>) in female albino mice. <i>Indian J. Physiol. Pharmacol.</i> 49(3): 305-312 IOFI (International Organization of Flavor Industries) cited in Temahoud 1996:600 Schulz H. et al. 1992. Charakterisierung von Grapefruitöl und Saft durch HPLC. <i>Z. Lebensmittel-Forsch.</i> 198:254-258 Stanley W. L. et al. 1971. Citrus coumarins. <i>J. Agric. Food Chem.</i> 19(6):1106-1110. Wagstaff D. J. 1991. Dietary exposure to furanocoumarins. <i>Regul. Toxic Pharmacol.</i> 14:261-272
<i>Citrus paradisi</i> Macfad. (<i>Citrus paradisi</i> Macfad., <i>Citrus grandis</i> (L.) Osbeck var. <i>aracemosa</i> (Roem.) B.C. Stone, <i>Citrus decurmana</i> (L.))	Rutaceae	Fruit, leaf, peel and pulp	Essential oil from the peel: furanocoumarins; e.g. psoralen, 5-hydroxypsoralen (bergapten), 5-methoxypsoralen (0.0005-0.013%), 5-geranylpsoralen (bergamottin)	so called "grape-fruit seed extract" has been known to contain e.g. quaternary ammonium compounds (e.g. benzethonium chloride)	
<i>Citrus reticulata</i> Blanco (<i>Citrus nobilis</i> Andr. non Louf)	Rutaceae	Bark and fruit	Essential oil: furanocoumarins; e.g. 8-methoxypsoralen		Benincasa M. et al. 1990. Analysis of lemon and bergamot essential oils by HPLC with microbore columns. <i>Chromatographia</i> 30(6):61-71 Zhou XM et al. 2012. Preventive effects of Citrus reticulata essential oil on theomycin-induced pulmonary fibrosis in rats and the mechanism. <i>Zhong Xi Yi Jie He Xue Bao</i> 10(2):200-208 Xue F. et al. 2012. Subacute toxicity assessment of carotenoids extracted from citrus peel (Nantingjinhui, Citrus reticulata Blanco) in rats. <i>Regul. Toxicol. Pharmacol.</i> 62(1):16-22
<i>Claviceps</i> spp.	Clavicipitaceae	Sclerotium	Genus in which species may contain ergot alkaloids derived from lysergic acid: e.g. ergometrine, ergotamine and ergoxines		Fantegrossi W.E. et al. 2008. The behavioral pharmacology of hallucinogens. <i>Biochem. Pharmacol.</i> 75: 17-33 Anton R. et al. 2000. Du Claviceps purpurea à l'ergot: l'ergot de seigle, son application et sa toxicité". <i>Industries des Céréales</i> , 119: 28-30 Eadie M. J. 2003. Convulsive ergotism: epidemics of the serotonin syndrome? <i>Lancet Neurol.</i> 2: 429-434.
<i>Clematis</i> spp.	Ranunculaceae	Whole plant	Genus in which species may contain lactones: e.g. protoanemonins and ranunculin (precursor) in the fresh herb	Adulteration with aristolochic acids containing species reported in the literature Clematis species are not known to contain aristolochic acid, however species <i>C. armandi</i> and <i>C. moniana</i> may be described by the same Chinese Pinyin name as an Aristolochia species. Protoanemonin only present in fresh herb.	Frome D., Pfander HJ and Anton R. 2009. <i>Plantes à risques</i> . Ed. Tec et Doc-Lavoisier, ISBN: 978-2-7430-0907-1
<i>Crocodendrum ibicouratum</i> L.	Lamiaceae (Labiatae)	Root	Macrocytic spermidinic alkaloids and deltanone-type diterpenes	Triterpene saponins	Frome D., Pfander HJ and Anton R. 2009. <i>Plantes à risques</i> . Ed. Tec et Doc-Lavoisier, ISBN: 978-2-7430-0907-1
<i>Crotopodium mentifolium</i> ssp. <i>ascendens</i> (Jordan) Govaerts (<i>Calceolaria ascendens</i> Jord.)	Lamiaceae (Labiatae)	Aerial part	Essential oil: monoterpene ketones: pulegone and derivatives: cis-isopulegone (75.2%), pulegone (6.9%), neo-isopulegone (6%), trans-isopulegone (4.5%)		Castillo P. et al. 2007. Composition and antimicrobial activity of the essential oil of <i>Crotopodium ascendens</i> (Jordan) Sampayo from Medellin. <i>Flavour and Fragrance Journal</i> 22(2):139-144
<i>Civula minima</i> Regel	Amaryllidaceae	Unspecified	Isoquinoline alkaloids: e.g. lycorine		Iken M. A. et al. 1992. Isolation of alkaloids from <i>C. minima</i> Regel. <i>J. Nat. Prod.</i> 45 (5): 564-573 Sewarn V. et al. 2001. Supercritical fluid extraction and analysis of compounds from <i>Civula minima</i> for diuretic activity. <i>Pharm. Med.</i> 67(5): 451-455
<i>Civula rosea</i> Jacq.	Clusiaceae (Guttiferae)	Unspecified	Polysoprenylated benzophenones; e.g. nannosone		Costa-Rubio O. et al. 2002. Polysoprenylated benzophenones in Cuban propolis: biological activity of nannosone. <i>Z. Naturforsch. C</i> 57(3-4): 372-378. Diaz-Carballeo D. et al. 2003. Novel antitumor compound isolated from <i>Civula rosea</i> . <i>Int J Clin. Pharmacol. Ther.</i> 41(12): 622-623
<i>Chidosecolus</i> spp.	Euphorbiaceae	Whole plant	Genus in which species may contain cyanogenic glycosides (limarin) ranging from 0.8 to 15 µg HCN equivalent/gram fresh weight.		Kul J.O. et al. 2004. Antioxidant capacity and phenolic content in leaf extracts of tree sorrel (<i>Chidosecolus</i> spp.). <i>J. Agric. Food Chem.</i> 52(1):17-21 Cordeiro RS. et al. 1983. The presence of limarin in <i>Chidosecolus oligandrus</i> (Euphorbiaceae). <i>Am. Acad. Bras. Cienc.</i> 55(1): 125-128 Loarca-Piña G. et al. 2010. Antioxidant, antimutagenic, and antidiabetic activities of edible leaves from <i>Chidosecolus chrysanthus</i> Mx. <i>Veg. J. Food Sci.</i> 75(2): 68-72
<i>Coccolus</i> spp.	Menispermaceae	Fruit	Genus in which species (e.g. <i>C. orbiculatus</i> , <i>C. trilobus</i>) may contain different alkaloids among which bisbenzyltetrahydroisoquinoline alkaloids: e.g. tetrandrine		Burleton J. 2009. <i>Pharmacognosie, Phytochimie, Plantes médicinales</i> . Ed. Tec & Doc-Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1188-8

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Botanical name	Family	parts of plants of possible concern	Chemical of concern	Remarks on toxic/adverse effects(s) not known to be related to the identified chemical(s) of concern	Specific References
<i>Codonopsis pilosula</i> (Franch.) Nannf.	Campanulaceae	Root		Saponin triterpenyl esters	Wakana D et al. 2011. The new triterpenyl esters, codonoplates A-C, isolated from <i>Codonopsis pilosula</i> . J Nat Med. 65(1):18-23
<i>Coffea arabica</i> L. (<i>Coffea vulgaria</i> Moench.)	Rubiaceae	Seed (bean)	Methylated xanthine derivative: caffeine Green coffee bean: 0.8 - 1.4% caffeine on dry basis		Andersson H. Cel. et al. 2004. Intake of caffeine and other methylxanthines during pregnancy and risk for adverse effects in pregnant women and their foetuses. TemaNord 565. IARC. 1991. Monograph No 51. Coffee, tea, mate, methylxanthines and methylglyoxal Mazzafari P. et al. 1992. Breeding for low seed caffeine content of coffee (<i>Coffea L.</i>) by interspecific hybridization. Euphytica 59:55-60. Bruneton J. 2009. Pharmacognosie, (PHY)ochimie, Plantes médicinales). Ed. Tec & Doc-Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1188-8 Clifford MN and Wilson KC. 1985. Coffee botany, biochemistry and production of beans and beverage. Croom Helm Ed. London.
<i>Coffea canephora</i> Pierre ex Froehner (<i>Coffea robusta</i> Lind. ex De Wild)	Rubiaceae	Seed (bean)	Methylated xanthine derivative: caffeine Green coffee bean: 1.7-4.0% caffeine on dry basis	In general somewhat higher caffeine content (up to 50%) in Robusta coffee compared to Arabica.	Andersson H. Cel. et al. 2004. Intake of caffeine and other methylxanthines during pregnancy and risk for adverse effects in pregnant women and their foetuses. TemaNord 565. IARC. 1991. Monograph No 51. Coffee, tea, mate, methylxanthines and methylglyoxal Mazzafari P. et al. 1992. Breeding for low seed caffeine content of coffee (<i>Coffea L.</i>) by interspecific hybridization. Euphytica 59:55-60. Council of Europe. 2007. Natural sources of Flavonurings. Report No. 2. Council of Europe Publishing. ISBN 978-92-871-6156-7 Clifford MN and Wilson KC. 1985. Coffee botany, biochemistry and production of beans and beverage. Croom Helm Ed. London.
<i>Cola acuminata</i> (P. Beauv.) Schott & Endl. (<i>Cola pseudo-acuminata</i> Engl., <i>Sterealia acuminata</i> P. Beauv.)	Malvaceae	Seed	Methylated xanthine derivatives: caffeine (2.4-2.6%), theobromine <0.1%.		Council of Europe. 2000. Natural sources of Flavonurings. Report No. 1. Council of Europe Publishing. ISBN 978-92-871-4324-7
<i>Cola milda</i> (Vent.) Schott & Endl. (<i>Cola acuminata</i> (P. Beauv.) Schott&Endl. var. <i>laticola</i> K.Schum., <i>Cola verta</i> K. Schum.)	Malvaceae	Seed	Methylated xanthine derivatives: caffeine (1.5-3.5%), theobromine 1%, theophylline		Council of Europe. 2000. Natural sources of Flavonurings. Report No. 1. Council of Europe Publishing. ISBN 978-92-871-4324-7
<i>Colchicum</i> spp.	Colchicaceae	Whole plant	Genus in which species may contain phenethylisoquinoline alkaloids: e.g. colchicine		Forne D, Pfander HJ and Anton R. 2009. Plantes à usages, Ed. Tec et Doc-Lavoisier, ISBN: 978-2-7430-0907-1
<i>Coelus forskohlii</i> (Willd.) Briq. (<i>Plectranthus barbatus</i> Andr.)	Lamiaceae (Labiatae)	Whole plant	Bicyclic diterpene with cyclic ether and lactone: forskohline	Novel Food catalogue: food use other than food supplement use would fall under the NF-Regulation	Forne D, Pfander HJ and Anton R. 2009. Plantes à usages, Ed. Tec et Doc-Lavoisier, ISBN: 978-2-7430-0907-1
<i>Colutea arborescens</i> L.	Leguminosae (Fabaceae)	Leaf and seed	Quinolizidine alkaloids: e.g. cystine Non-protein amino acid: L-canavanine (5%)		Rohm L., Daurerier M. and Korman K. 1994. Giftpilzen - Pflanzengifte, Vorkommen WIKUNG Therapie, ecomed ISBN 3-609-44810-4 Forne D., Pfander H. J. et Anton R., « Plantes à usages », Ed. Tec et Doc-Lavoisier (2009), ISBN: 978-2-7430-0907-1 Grosvenor PW et al. 1996. Colubiquone and coulehydrone, antifungal isoflavonoids from <i>Colutea arborescens</i> . Phytocochemistry. 43(2):377-380 Naimonik GN et al. 1994. Study of tannins in <i>Comarum palustre</i> L., a bound form of tannins. Angew Chem. 13, ZT-28.
<i>Comarum palustre</i> L.	Rosaceae	Root		High content of tannins. Intake of high doses of tannins may cause hepatotoxicity	Bassene E et al. 1986. African medicinal plants. Alkaloids of <i>Comarum micranthum</i> G. Don (Kirkella). Ann Pharm Fr. 44(3), 191-196 Ogan AU. 1972. The alkaloids in the leaves of <i>Comarum micranthum</i> . Studies on West African medicinal plants. VII. Planta Med. 21(2):210-7 Wesch CR. 2010. Chemistry and Pharmacology of Kinelba (<i>Comarum micranthum</i>), a West-African medicinal plant. PhD Thesis. New Brunswick - University of New Jersey, available at: www.nrcs3.libraries.udel.edu/outprints.php?printid=9535
<i>Combebum micranthum</i> G.Don. (<i>Callium</i> , <i>C. floribundum</i> , <i>C. parviflorum</i> , <i>C. rainboallii</i>)	Combrataceae	Leaf		Presence of flavan-ppiferoline alkaloids. Novel Food catalogue: food use other than food supplement use would fall under the NF-Regulation	Forne D, Pfander HJ and Anton R. 2009. Plantes à usages, Ed. Tec et Doc-Lavoisier, ISBN: 978-2-7430-0907-1
<i>Commiphora mikulii</i> Engl.	Burseraceae	Old-gum-resin from the trunk	Essential oil (0.4%) with phenylpropanoids: e.g. methylchavicol. In unspecified quantities. Terpenoids: e.g. myrcene, dmyrcene, polymyrcene		Delgado F. et al. 1993. Study on embryo-toxicity of beta-myrcene in the rat. Food Chem Toxicol 31(1), 31-35. Bruneton J. 2009. Pharmacognosie, (PHY)ochimie, Plantes médicinales). Ed. Tec & Doc-Lavoisier, Paris, 4ème édition, 1269 pages, ISBN: 978-2-7430-1188-8 Hagers Handbuch der Pharmazwissenschaften Praxis 1996, Springer Verlag, ISBN 3-540-52686-9
<i>Commiphora myrrha</i> (Nees) Engl.	Burseraceae	Old-gum-resin from the trunk	Volatile fraction: furanosesquiterpenes: e.g. curzerenone, methoxy-furanolene, furanoclemenes, furano-germanaranes	Volatile fraction present only in the freshly collected oleogum resin Hepato-nephrotoxicity described	Ormer SA et al. 1999. Effects on rats of <i>Commiphora myrrha</i> extract given by different routes of administration. Vet. Hum., Toxicol. 41(6), 193-6 Wicthl M. and Anton R. 2003. Plantes thérapeutiques (Tradition, pratique officinale, science et thérapeutique), Ed. Tec & Doc-Lavoisier, Paris, 2ème édition, 692 pages, ISBN: 2-7430-0931-3
<i>Conium maculatum</i> L.	Apiaceae (Umbelliferae)	Whole plant	Piperidine alkaloids: conine (3% in immature fruit, 1% in mature fruit). In the rest of the plant: γ-coniine (more active than conine).		Forne D, Pfander HJ and Anton R. 2009. Plantes à usages, Ed. Tec et Doc-Lavoisier, ISBN: 978-2-7430-0907-1 Council of Europe. 2008. Natural sources of Flavonurings. Report No. 3. Council of Europe Publishing. ISBN 978-92-871-6422-3
<i>Convolvularia majalis</i> L.	Asplragaceae	Whole plant	Cardenolide glycosides (0.2-0.4% in dried leaf and 0.5% in flower and seed); e.g. convallatoxin and glicoxalactone and convallactone in the seed.		Forne D, Pfander HJ and Anton R. 2009. Plantes à usages, Ed. Tec et Doc-Lavoisier, ISBN: 978-2-7430-0907-1
<i>Convolvulus</i> spp.	Convolvulaceae	Whole plant	Genus in which species may contain tropane alkaloids (tryptamine derivatives): e.g. ergine, ysergol, clavines Genus in which species may contain tropane alkaloids: e.g. tropanol, pseudotropanol. Genus in which some species contain a resin (roots) with strong purgative effect: e.g. Jalapine		Forne D, Pfander HJ and Anton R. 2009. Plantes à usages, Ed. Tec et Doc-Lavoisier, ISBN: 978-2-7430-0907-1 Bruneton J. 2005. Plantes toxiques (Végétaux dangereux pour l'homme et les animaux). Ed. Tec & Doc-Lavoisier, Paris, 2ème édition, ISBN: 2-7430-0866-7

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<i>Copallina officinalis</i> (Jacq.) L.	Leguminosae (Fabaceae)	Bark		Presence of chemically not defined diterpenes in the bark oleoresin	Chen F. et al. 2008. Within-pit distribution and emission of sesquiterpenes from <i>Copallina officinalis</i> . Plant Physiol Biochem. 47(11-12):1017-1023. Brito NM, et al. 2010. The effect of copaliba balsam on Walker 256 carcinoma inoculated into the vagina and uterine cervix of female rats. Acta Ch. Bras.25(2): 176-180
<i>Copis</i> spp.	Ranunculaceae	Whole plant	Genus in which species may contain isoquinoline alkaloids: e.g. berberine, stylopine, copisine		Frome D, Pfander HJ and Anton R. 2009. Plantes à risques. Ed. Tec & Doc-Lavoisier, ISBN: 978-2-7430-0907-1 Bruneton J. 2005. Plantes toxiques (Végétaux dangereux pour l'homme et les animaux). Ed. Tec & Doc-Lavoisier, Paris, 3ème édition, ISBN : 2-7430-0906-7
<i>Corchorus olitorius</i> L.	Malvaceae	Seed	Cardenolide glycosides: erythronoside, olitorosides A and B, corolioside, hevelicoside, camogrol, perflorigen, diglotoxigenin, glucovalatomonoside, deglucovalatomonoside, exaltatomonoside.		Bruneton J. 2009. Pharmacognosie, (phytochimie, Plantes médicinales), Ed. Tec & Doc-Lavoisier, Paris, 4ème édition, ISBN : 978-2-7430-1188-8 Hagers Handbuch der Pharmazeutischen Praxis, Springer Verlag, 1999, ISBN: 3-540-52688-9 Nakamura T et al. 1998. Cardenolide glycosides from seeds of <i>Corchorus olitorius</i> . Phytochemistry, 48(7):2097-2101.
<i>Coriandrum sativum</i> L.	Apiaceae (Umbelliferae)	Aerial part	Essential oil from the fruit: bicyclic monoterpene: camphor (3-9%)		Wicini M, and Anton R. 2003. Plantes thérapeutiques (Tradition, pratique officielle, science et thérapeutique). Ed. Tec & Doc-Lavoisier, Paris, 2ème édition, 692 pages, ISBN : 2-7430-0631-5 Bruneton J. 2009. Pharmacognosie, (phytochimie, Plantes médicinales), Ed. Tec & Doc-Lavoisier, Paris, 4ème édition, ISBN : 978-2-7430-1188-8 Hagers Handbuch der Pharmazeutischen Praxis, Springer Verlag, 1999, ISBN: 3-540-52688-9 Council of Europe. 2005. Active principles (constituents of chemical concern) contained in natural sources of flavonoids. Ed: Council of Europe Publishing http://www.coe.int/t/tae/social_consensus/oc-sop/dluc_neutral/flavonoid_substances/Active%20principles.pdf
<i>Coriaria myrsinifolia</i> L.	Coriariaceae	Aerial part	Sesquiterpene lactones: e.g. coriamyrtin, conarin, coriamyrtin.	High concentration of coriamyrtin in berries	Frome D, Pfander HJ and Anton R. 2009. Plantes à risques. Ed. Tec & Doc-Lavoisier, ISBN: 978-2-7430-0907-1 Hagers Handbuch der Pharmazeutischen Praxis, Springer Verlag, 1999, ISBN: 3-540-52688-9 de Haro L, et al. 2005. Poisoning by <i>Coriaria myrsinifolia</i> Linaeus: a new case report and review of the literature. Toxicon, 46(6):600-603
<i>Coriaria thymifolia</i> Humb. & Bonpl.	Coriariaceae	Aerial part	Sesquiterpene lactones: e.g. coriamyrtin, conarin, pseudolulone, lulone.		Duke J.A. 1985. Handbook of medicinal herbs. CRC Press, Inc. ISBN 0-8493-3630-9
<i>Coronilla scorpioides</i> Koch.	Leguminosae (Fabaceae)	Whole plant	Cardenolides: e.g. lycorinose and the aglycone lycorinogenine.		Hagers Handbuch der Pharmazeutischen Praxis, Springer Verlag, 1999, ISBN: 3-540-52688-9 Srinova-Sawani. 1997. Pharmacology of <i>Coronilla scorpioides</i> , a new cardiac drug. Farmacol Toksikol. (3), 59-63 Kornissenko Nf et al. 1989. On the chemotaxonomic characterization of <i>Coronilla scorpioides</i> and <i>C. repanda</i> . Planta Med. 17(2): 170-177
<i>Coronilla varia</i> L.	Leguminosae (Fabaceae)	Whole plant	Seed: cardenolide, e.g. lycorinose and deglucohyrcanose Other plant parts (except seeds): 3- nitropanoic acid derivatives		Frome D, Pfander HJ and Anton R. 2009. Plantes à risques. Ed. Tec & Doc-Lavoisier, ISBN: 978-2-7430-0907-1 Gold K et al. 1991. Studies on the distribution of a naturally occurring nitrophenolic acid in crownvetch (<i>Coronilla varia</i> , Fabaceae). Econ. Botany 45(3): 324-338
<i>Corydalis</i> spp.	Papaveraceae	Whole plant	Genus in which species may contain isoquinoline alkaloids: 6% of dry weight in tuber: e.g. bulbocapnine, corydaline, corydine, copisine, palmatine, N-methylbulandamine, allocorydopine, protopine, corycavidine, glaucine, corydine, bulbocapnine, corydaline, corypalmine, tetrahydropalmatine, canadine, thalictrate.		Zhong-Ze Ma et al. 2008. Isoquinoline alkaloids isolated from <i>Corydalis yanhusuo</i> and their binding affinities at the dopamine D1 receptor. Molecules 13(9): 2303-2312 Frome D, Pfander HJ and Anton R. 2009. Plantes à risques. Ed. Tec & Doc-Lavoisier, ISBN: 978-2-7430-0907-1
<i>Corynanthe</i> spp.	Rubiaceae	Bark	Genus in which species may contain yohimbane alkaloids: e.g. corynanthine, yohimbine (= quabacinine)		Bruneton J. 2009. Pharmacognosie, (phytochimie, Plantes médicinales), Ed. Tec & Doc-Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1188-8 Frome D, Pfander HJ and Anton R. 2009. Plantes à risques. Ed. Tec & Doc-Lavoisier, ISBN: 978-2-7430-0907-1
<i>Coarctaster</i> spp.	Rosaceae	Whole plant	Genus in which species may contain cyanogenic glycosides: prunasin from the bark; amygdalin and prunasin from the fruit (amygdalin, prunasin)		Hagers Handbuch der Pharmazeutischen Praxis, Springer Verlag, 1999, ISBN: 3-540-52688-9
<i>Coarctaster oppositifolia</i> Taub. (<i>Latalia oppositifolia</i> Aubl.)	Leguminosae (Fabaceae)	Seed	Coumarin		Braskar VH and al. 2009. Evaluation of the anti-fertility activity of stem bark of <i>Cataeva nurvala</i> buch. Afr J Biotech 8, 6453-6456 Sharma B.B. et al. 1983. Antifertility Screening of Plants. Part I. Effect of Ten Indigenous Plants on Early Pregnancy in Albino Rats. Pharm Biol 21 (4), 183-187 Fennell CW and van Steeden J. 2001. Citrus species in traditional and modern medicine. J. Ethnopharmacol. 72(1), 15-26
<i>Crateva nurvala</i> Buch.-Ham. (<i>Cratevalopsperma</i> Kuz.)	Caprifoliaceae	Bark	Thienopyrans (lupane type): e.g. lupcol	Antifertility activity (decrease of the implantation) when administered orally to rats.	Gastaldo P. 1987. Compendio della Flora Officinale Italiana Padova. Ed. Piccin, ISBN: 8629305982, 9788629305987 Ozcan M, et al. 2006. Constituents of the Essential Oil of Sea Fennel (<i>Citharum maritimum</i> L.). Growing Wild in Turkey. J. Medicinal Food. 9(1), 128-130 Frome D., Pfander H. J. and Anton R. 2009. Plantes à risques. Ed. Tec & Doc-Lavoisier, ISBN: 978-2-7430-0907-1 Asteris K, et al. 2004. Patterns of Pyridizoline alkaloids in 12 Ethiopian <i>Crotalaria</i> species. Biochemical Systematics and Ecology, 32(10), 915-930. Hagers Handbuch der Pharmazeutischen Praxis, Springer Verlag, 1999, ISBN: 3-540-52688-9
<i>Critinum asiaticum</i> L.	Amaryllidaceae	Bulb	Isoquinoline alkaloids (Amaryllidaceae alkaloids): e.g. protoirine, lycorine, citrifine, cinnamine		Frome D, Pfander HJ and Anton R. 2009. Plantes à risques. Ed. Tec & Doc-Lavoisier, ISBN: 978-2-7430-0907-1
<i>Critinum maritimum</i> L.	Apiaceae (Umbelliferae)	Leaf	Essential oil: phenylpropanoids: e.g. methylchavicol (3-4%)		Frome D, Pfander H. J. and Anton R. 2009. Plantes à risques. Ed. Tec & Doc-Lavoisier, ISBN: 978-2-7430-0907-1
<i>Crotalaria</i> spp.	Leguminosae (Fabaceae)	Aerial part	Genus in which species may contain unsaturated pyridizoline alkaloids		Frome D, Pfander H. J. and Anton R. 2009. Plantes à risques. Ed. Tec & Doc-Lavoisier, ISBN: 978-2-7430-0907-1
<i>Crotone</i> spp.	Euphorbiaceae	Whole plant	Genus in which species may contain diterpene esters (phorbol-esters), Isoquinoline alkaloids (gopifoline, morpholine, podocarpiline type alkaloids) and lectins, e.g. coum		Duke J.A. 1985. Handbook of medicinal herbs. CRC Press, Inc. ISBN 0-8493-3630-9 Cook DR et al. 1990. Suspected <i>Cryptospegia grandiflora</i> (luber wine) poisoning in horses. Austr. Vet. J. 67(9) 344
<i>Cryptospegia</i> spp.	Apocynaceae	Whole plant	Genus in which species may contain cardenolide glycosides e.g.: oleandrigenin 3-hammnoside and aglycones: e.g. oleandrigenin, glicosigenin, 16-amhydroglicosigenin, 16-propionylglicosigenin.		

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Botanical name	Family	Parts of plants of possible concern	Chemical of concern	Remarks on toxic/adverse effect(s) not known to be related to the identified chemical(s) of concern	Specific References
<i>Cucumis sativus</i> L.	Cucurbitaceae	Whole plant	Possible occurrence of the oxygenated tetracyclic terpenes: cucurbitacin C in leaf and fruit and of cucurbitacins C and B in root.		Van Keulen JA, 1981. Fluorodensitometric estimation of cucurbitacin-C in leaves of <i>Cucumis sativus</i> L. Plant Foods for Human Nutrition (Formerly Qualitas Panoram), 31(2), 129-137.
<i>Cucurbita maxima</i> Duch.	Cucurbitaceae	Whole plant	Possible occurrence of oxygenated tetracyclic terpenes: cucurbitacins B and C.		Rehm S et al. 1957. Bitter principles of the cucurbitaceae. VIII.—cucurbitacins in seedlings—occurrence, biochemistry and general aspects. J. Sci. Food. Agr. 8(12), 687 - 691.
<i>Cucurbita pepo</i> L.	Cucurbitaceae	Fruit	Possible occurrence of oxygenated tetracyclic terpenes: cucurbitacins.	Fruits of cultivated squash and other pumpkins have been cultured to be "free of cucurbitacins", and are assumed to contain a suppressor gene of a mutation responsible for absence of cucurbitacins. However, back-mutations occur randomly which may lead to plants with toxic and bitter fruits.	Terna Nord 2006:556. Cucurbitacins in plant food. Nordic Council of Ministers. ISBN 92-893-1381-1
<i>Cuminum cyminum</i> L.	Apiaceae (Lumbelliferae)	Fruit	Essential oil from fruit: phenylpropanoids: e.g. methylchavicol (30ppm) and monoterpenes: monoterpene etheroxide: 1,8-cineole (0.2-0.4%).		Council of Europe. 2007. Natural sources of flavonoids. Report No. 2. Council of Europe Publishing. ISBN 978-92-871-6156-7.
<i>Curcuma kwangsiensis</i> S. Lee & C. R. Liang	Zingiberaceae	Rhizome	Essential oil: monoterpene etheroxide: 1,8-cineole.		Zhu You-Ping. 1998. Chinese Materia medica. Chemistry, pharmacology and applications. CRC Press. ISBN 1-3: 978-9057022822.
<i>Curcuma longa</i> L.	Zingiberaceae	Rhizome	Essential oil: monoterpene etheroxide: 1,8-cineole and bicyclic monoterpenes: e.g. camphor		Council of Europe. 2000. Natural sources of flavonoids. Report No. 1. Council of Europe Publishing. ISBN 978-92-871-4324-2.
(<i>Curcuma domestica</i> Val., <i>Curcuma domestica</i> L'f., <i>Amomum curcuma</i> Jacq.)	Zingiberaceae	Rhizome	Essential oil: bicyclic monoterpenes: e.g. camphor (10-16%)		Zhu You-Ping. 1998. Chinese Materia medica. Chemistry, pharmacology and applications. CRC Press. ISBN 1-3: 978-9057022822.
<i>Curcuma phaeocaulis</i> Valeton	Zingiberaceae	Rhizome	Essential oil: bicyclic monoterpenes: e.g. camphor (10-16%)		Zhu You-Ping. 1998. Chinese Materia medica. Chemistry, pharmacology and applications. CRC Press. ISBN 1-3: 978-9057022822.
<i>Curcuma wenyujin</i> Y.H.Chen & C.Ling	Zingiberaceae	Rhizome	Essential oil (3-12%): monoterpenes: monoterpene etheroxide: 1,8-cineole (up to 40%), bicyclic monoterpenes: camphor (1%);		Zhu You-Ping. 1998. Chinese Materia medica. Chemistry, pharmacology and applications. CRC Press. ISBN 1-3: 978-9057022822.
<i>Curcuma xanthorrhiza</i> Roxb.	Zingiberaceae	Rhizome	Essential oil (3-12%): monoterpenes: monoterpene etheroxide: 1,8-cineole (up to 40%), bicyclic monoterpenes: camphor (1%);		U.S. et al. 2011. Chemical Composition and Product Quality Control of Turmeric (<i>Curcuma longa</i> L.). Pharmaceutical Cosm. 2, 28-54.
<i>Cyathula officinalis</i> Kuan	Amaranthaceae (Chenopodiaceae)	Root	Coumarins: e.g. scoparone (6, 7-dimethoxycoumarin)	The saponins (federgerin- and gypsoginim-type saponins) are thought to stimulate uterus contraction and can lead to abortion, however scoparone is probably the causative agent.	Bruleton J. 2009. Pharmacognosie, (Phytochimie, Plantes médicinales), Ed. Tec & Doc. Lavoisier. Paris, 4ème édition, 1269 pages, ISBN : 978-2-7430-1188-8; Hagers Handbuch der Pharmazeutischen Praxis 1998; Springer Verlag. ISBN 3-540-52688-9
<i>Cycas</i> spp.	Cycadaceae	Leaf, pollen, seed	Genus in which species may contain the amine oxide: cyanasin		Chandhoke N. 1979. Scoparone: effect on reproductive processes in rats. Indian J. Exp. Biol. 17, 740-742.
<i>Cyclopentherium europaeum</i> L.	Primulaceae	Tuber	Terpene saponins: e.g. cycloperin		Reid MT et al. 2006. Rapid analysis of constituents of Radix <i>Cyclopentherium</i> using hydrophilic interaction-reverse phase LC/MS. Journal of Separation Science. 29(22), 3988 - 3995
(<i>C. andrasicum</i> Mill.)	Primulaceae	Tuber	Terpene saponins: e.g. cycloperin		Zhu You-Ping. 1998. Chinese Materia medica. Chemistry, pharmacology and applications. CRC Press. ISBN 1-3: 978-9057022822.
<i>Cymbopogon citratus</i> (DC.) Stapf	Poaceae (Gramineae)	Aerial part	Essential oil (0.2%-0.4%); bicyclic monoterpenes: alpha-thujone (up to 0.1%) and monoterpene etheroxide: 1,8-cineole (traces)		Palmsont S. 1998. The medicinal plants of India. Part 1. The Council of Europe Publishing. ISBN 978-92-871-4324-2.
<i>Cymbopogon martini</i> (Roxb.) Wilt. Watson	Poaceae (Gramineae)	Aerial part	Essential oil: phenylpropanoids: e.g. methylchavicol (traces)		Council of Europe. 2000. Natural sources of flavonoids. Report No. 1. Council of Europe Publishing. ISBN 978-92-871-4324-2.
<i>Cymbopogon nardus</i> (L.) Hook. f.	Poaceae (Gramineae)	Aerial part	Essential oil: phenylpropanoids: e.g. methylchavicol (51-204 ppm).		Council of Europe. 2000. Natural sources of flavonoids. Report No. 1. Council of Europe Publishing. ISBN 978-92-871-4324-2.
<i>Cynanchum vineosotum</i> (L.) Pers. See <i>Vincetoxicum thurandiana</i> Wedd.					
<i>Cynodon dactylon</i> (L.) Pers.	Poaceae (Gramineae)	Aerial part	Cyanogenic glycosides		Ortega P.J. 1993. Plant structure and the acceptability of different grasses to sheep. J Range Manage 46: 232-236.
<i>Cynoglossum</i> spp.	Borraginaceae	Whole plant	Genus in which species may contain unsaturated pyridizidine alkaloids		Van Dam N et al. 1995. Distribution, biosynthesis and turnover of pyridizidine alkaloids in <i>Cynodossium officinale</i> . Phytochemistry. 39(2), 287-292.
<i>Cyperus rotundus</i> L.	Cyperaceae	Rhizome	Sesquiterpene pyridine alkaloids: rotundines A-C (0.21%-0.24%); butadienolide glycosides (0.62%-0.74%)		Rehman AB. 2007. Pharmacological studies on traditional medicine (<i>Cyperus rotundus</i>) used in Pakistan. Thesis. Department of pharmacology, University of Karachi.
<i>Cyrtopodium calceolus</i> L.	Orchidaceae	Root	Quinones e.g. cyrtopedin a non-terpeneoid phenanthraquinone		Jeong S. Y et al. 2000. Rotundines A-C, three novel sesquiterpene alkaloids from <i>Cyperus rotundus</i> . J. Nat. Prod. 63: 675-678.
<i>Cytisus</i> spp.	Leguminosae (Fabaceae)	Whole plant	Genus in which species may contain quinazoline alkaloids: e.g. cytisine		Schmalte H and Hansen BM. 1979. A new sensitizing quinine from Italy. slipper (<i>Cyrtopodium calceolus</i>). Naturwissenschaften. 66(10), 527-528.
<i>Dalechampia scandens</i> L.	Euphorbiaceae	Leaf and stem	Diterpenes: cyanogenic glycosides: lectins	Presence of histamine	Barnes J, Anderson LA, Phillips JD. 2007. Herbal medicines (3rd ed). London: Pharmaceutical Press. ISBN 978-0-85369-642-1
<i>Daphne</i> spp.	Thymelaeaceae	Whole plant	Genus in which species may contain diterpene esters: e.g. daphnane derivatives		Hagers Handbuch der Pharmazeutischen Praxis 1998; Springer Verlag. ISBN 3-540-52688-9
<i>Datura</i> spp.	Solanaceae	Whole plant	Genus in which species may contain tropane alkaloids: e.g. atropine, scopolamine	The fresh plant contains hyoscyamine, twice more active than atropine (racemic mixture)	Jose S. Flores JS et al. 2001. Plantas de la flora Yucatanense que provocan alguna toxicidad en el humano. Rev Biomed 12: 66-96.

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<i>Daucus carota</i> L.	Apiaceae (Umbelliferae)	Fruit	Essential oil: phenylpropanoids e.g. methylisoeugenol, methylgeranyl, elemicin, beta asarone		Saad HEA et al. 1995. Essential oils of <i>Daucus carota</i> ssp. <i>Maximus</i> . <i>Pharm Acta Helv</i> 70: 79-84.
<i>Delphinium</i> spp.	Ranunculaceae	Whole plant	Genus in which species may contain diterpene alkaloids, e.g. alicine, alicone, delicosine, methylxacaconine		Frome D., Pfander H.J. and Anton R. 2009. <i>Plantes à risques</i> . Ed. Tec et Doc-Lavoisier, ISBN: 978-2-7430-0907-1
<i>Delris</i> spp.	Leguminosae (Fabaceae)	Root	Genus in which species may contain rotenoids: e.g. rotenone		Burtonel J. 2009. <i>Pharmacopositive (P)lycopolime, Plantes médicinales</i> . Ed. Tec & Doc-Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1188-8
<i>Desmodium</i> spp.	Leguminosae (Fabaceae)	Whole plant	Genus in which species may contain tryptamine derivatives: e.g. 5-methoxy-dimethyltryptamine and 5-hydroxy-dimethyltryptamine (bufotenine)		Troul K. 1997. 'Toul's notes on the genus <i>Desmodium</i> . (Chemistry, Ethnomedicine, Pharmacology, Synonyms and Miscellany). Copyright ©1997 by Troul & Friends & 2002 by Troul's Notes Mydtronic Productions. Adapted for webviewing March 2004
<i>Dianthus caryophyllus</i> L.	Caryophyllaceae	Aerial part	Terpene saponins		Burtonel J. 2009. <i>Pharmacopositive (P)lycopolime, Plantes médicinales</i> . Ed. Tec & Doc-Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1188-8
<i>Dicentra spectabilis</i> (L.) Lem.	Papaveraceae	Whole plant	Isoquinoline alkaloids from epigeal parts (0.17%), and from the root (0.25%) e.g.: dihydroscopamine, sanguinarine, scoulerine, chelanthidine, conidine, and protopine		Isralov JA et al. 1984. Alkaloids of <i>Dicentra</i> . <i>Chem Nat Prod</i> 20 (1): 74-76
<i>Dichondra repens</i> J.H. Forst. & G. Forst.	Convolvulaceae	Whole plant	Coumarins: e.g. scopolein		Obidia O and Chasi SC. 1991. Coumarin compounds in cassava diets: 2 health implications of scopolein in garli. <i>Plant Foods for Human Nutrition</i> 41: 283-289
<i>Dicliptera alba</i> L.	Rubiaceae	Whole plant	Furanocoumarins (psoralens) from leaf: e.g. bergapten, xanthoxon, autaprene; Furoquinoline alkaloids from leaf: e.g. haplopine, robustine, dictamine, and gamma-agarine; Essential oil from leaf: phenylpropanoids: e.g. methylchavicol and trans-anethole; Furanocoumarin alkaloids from root (0.04%-0.09%); e.g. dictamine (0.003%), gamma agarine (0.002%),...; Essential oil from root: sesquiterpene lactones: e.g. transionin (1.2%) ; Furoquinoline alkaloids from root bark: e.g. dictamine (0.29%), gamma agarine (0.014%)		Muller H. 1978. Phototoxicity of <i>Dicliptera alba</i> . <i>Compt Rendus Acad Sci Paris</i> 287: 289-291 Besser KHC et al. 1994. The essential oil composition of <i>Dicliptera alba</i> from Turkey. <i>Phytother Res</i> 8: 481-482 Mizuta M, Kawanishi H. 1985. Mutagenic activities of dictamine and gamma-agarine from dictamnolide cortex (Rubiaceae). <i>Mutat. Res</i> 144: 221-225 Prakash A. O. et al. 1986. Evaluation of some indigenous plants for anti-implantation activity in rats. <i>Proste</i> 25: 151-155.
<i>Dicliptera dasycaulis</i> Turcz.	Rubiaceae	Whole plant	Furoquinoline alkaloids from root: e.g. dictamine, trigonelline, skrimianine (β-agarine), γ-agarine, dasycaepamine, polydesmine; Furanocoumarins from aerial part: e.g. psoralen, bergapten, xanthoxon		But PPH. (Ed.) 1997. Northeast Asia. International collation of traditional and folk medicine. A project of Unesco. World Sc. Pub. ISBN: 981023130X.
<i>Difenbachia</i> spp.	Araceae	Whole plant	Genus in which species may contain oxalate raphides, proteolytic enzymes and cyanogenic glycosides		Kubajala B. et al. 1981. Study of <i>Difenbachia</i> induced oedema in mice and rats hindpaw: respective role of oxalate needles and tyrosin-like protease. <i>Toxicol. Appl. Pharmacol.</i> 58: 444-451
<i>Digitalis</i> spp.	Plantaginaceae	Whole plant	Genus in which species may contain cardenolides (digitalis glycosides): e.g. digoxin		Frome D., Pfander H.J. and Anton R. 2009. <i>Plantes à risques</i> . Ed. Tec et Doc-Lavoisier, ISBN: 978-2-7430-0907-1
<i>Dioscorea</i> spp.	Dioscoreaceae	Tuber	Genus in which species may contain pyridinal alkaloids: e.g. Dioscoreine		Frome D., Pfander H.J. and Anton R. 2009. <i>Plantes à risques</i> . Ed. Tec et Doc-Lavoisier, ISBN: 978-2-7430-0907-1
<i>Diplocisia</i> spp.	Menispermaceae	Whole plant	Genus in which species may contain isoquinoline alkaloids (aporphine-type alkaloids): e.g. metidine, asinoline, acutidine		Corneil GA (Ed.). 2000. <i>Chemistry and Biology</i> . Volume 54 (Alkaloids). Academic Press. ISBN: 0-12-469954
<i>Diplopelys cabrerana</i> (Cultrac.) B.Gates	Malpighiaceae	Whole plant	Tryptamine alkaloids: e.g. dimethyltryptamine, harmaline derivatives		Books LC. 2010. <i>Psychedelic Tryptamine Carriers: Palicope Cubensis, Ayahausca, Psilocybin Mushrooms, Mimosa Tenillora, Harmal. List of Psilocybin Mushrooms</i> . General Books LLC. ISBN: 1155980922, 9781155980928.
<i>Diphenyl odorata</i> (Alba) Willd.	Leguminosae (Fabaceae)	Seed	Pentane/dichloromethane extract: coumarin 3,6 g/kg; camphor >1 mg/kg; Methanol extract: coumarin 23.25 g/kg; Tonka bean absolute: coumarin 390.510 g/kg.		Council of Europe. 2007. <i>Natural sources of flavonoids</i> . Report No. 2. Council of Europe Publishing. ISBN 978-92-87-6156-7.
<i>Dracoulum</i> spp.	Araceae	Whole plant	Genus in which species may contain calcium oxalate raphides		Campos R. and Balick ML. 2009. <i>Plants of Semillas Sagradas: An Ethnomedicinal Garden in Costa Rica</i> . Edited by Ruth Costales and Katherine Herrera. Frica Luna Nueva Editores de Costa Rica, S.A. Rafael. ISBN: 978-0-615-27415-7
<i>Drosera anglica</i> Huds.	Droseraceae	Aerial part	Naphthoquinone derivatives: e.g. pumbinagin		Balaji YPS (Ed.). 1993. <i>Biotechnology in agriculture and forestry</i> 24. <i>Medicinal and aromatic plants</i> . V. Springer, Verlag. ISBN: 3-540-56009-4
<i>Drosera intermedia</i> Hayne	Droseraceae	Aerial part	Naphthoquinone derivatives: e.g. pumbinagin		Balaji YPS (Ed.). 1993. <i>Biotechnology in agriculture and forestry</i> 24. <i>Medicinal and aromatic plants</i> . V. Springer, Verlag. ISBN: 3-540-56009-4
<i>Drosera rotundifolia</i> L.	Droseraceae	Aerial part	Naphthoquinone derivatives: e.g. pumbinagin		Balaji YPS (Ed.). 1993. <i>Biotechnology in agriculture and forestry</i> 24. <i>Medicinal and aromatic plants</i> . V. Springer, Verlag. ISBN: 3-540-56009-4
<i>Dryobalanops aromatica</i> C.F. Gaertn.	Dipterocarpaceae	Stem	Bicyclic monoterpene alcohol: e.g. borned and bicyclic monoterpene ketone: e.g. camphor		Segel E and Watson S. 1986. <i>Camphor toxicity</i> . <i>Pediatr. Clin N Am</i> . 33: 375-379
<i>Dryopteris</i> spp.	Dryopteridaceae	Whole plant	Genus in which species may contain flicine which is a mixture of different acylphloguonoids (e.g. aspidin, albispidin). Some species may also contain the norsesquiterpene dialutiside		Klaus Wienther (Ed.) 2010. <i>Fern ecology</i> . Cambridge University Press. ISBN: 9780521728201.
<i>Duboisia</i> spp.	Solanaceae	Whole plant	Genus in which species may contain tropane alkaloids: e.g. atropine, scopolamine...	The fresh plant contains hyoscyamine, twice more active than atropine (racemic mixture)	Pearm J. 1981. <i>Clinical hyoscyne poisoning with alkaloids of the native corkwood</i> . <i>Duboisia. Med. J. Aust</i> 2 (8): 422-423
<i>Dysphania ambrosioides</i> (L.) Mesyakin & Clemans (Chenopodium ambrosioides L.)	Amaranthaceae (Chenopodiaceae)	Leaf and seed	Essential oil from leaf (0.7%) and from urticae seed (2.5%); peroxygenated monoterpenes: e.g. ascaridole (from 10% to 70% depending on the origin) and phenylpropanoids: e.g. safrole		Kills MM. 1985. <i>Studies on the traditional herbal antiepileptic chenopodium ambrosioides L.: Ethnopharmacological evaluation and clinical field trials</i> . Soc. Sci. Med. 21: 879-886 http://www.food-info.net/uk/index.htm Federation Proceedings 1948. <i>Federation of American Societies for Experimental Biology</i> . 7: 252
<i>Dysphania anthriscinica</i> (L.) Mesyakin & Clemans (Chenopodium anthriscinicum L.; C. anthriscinicum L. var. anthriscinicum L.)	Amaranthaceae (Chenopodiaceae)	Leaf and seed	Essential oil: peroxygenated monoterpene: ascaridole and phenylpropanoids: e.g. safrole		Federation Proceedings 1948. <i>Federation of American Societies for Experimental Biology</i> . 7: 252
<i>Ecalium siderium</i> (L.) A. Rich.	Cucurbitaceae	Aerial part	Oxygenated tetracyclic terpenes: cucurbitachins (from fruit: 3.84%, from stem: 1.34%, from leaf: 0.34%)		Cuauhtli Genetics Cooperative Report 26:66-69 (2003). Maryland USA. ISSN: 1064-5594
<i>Echinops ritro</i> L.	Compositae (Asteraceae)	Seed	Quinolone alkaloids (0.5%); e.g. echinopsine		Tuova AD et al. 1957. <i>Pharmacology of the new alkaloid Echinopsine</i> . <i>Pharmacol. Toxicol</i> 20: 236-240 Di Cosmo F. et al. 1982. <i>Photo-induced fungicidal activity elicited by naturally occurring thiophene derivatives</i> . <i>J. Pestic. Sci.</i> 13: 589-594.

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<i>Echinops sphaerocephalus</i> L.	Compositae (Asteraceae)	Seed	Quinoline alkaloids: e.g. echinopsine		Tuova AD, et al. 1957. Pharmacology of the new alkaloid Echinopsine. <i>Pharmacol Toxicol.</i> 20: 236-240. Di Cosimo F, et al. 1992. Photo-induced fungicidal activity elicited by naturally occurring thienopyrene derivatives. <i>J. Pestic. Sci.</i> 13: 589-594.
<i>Echium</i> spp.	Borraginaceae	Whole plant	Genus in which species may contain unsaturated pyridizidine alkaloids		Frome D., Pfander H.J. and Anton R. 2009. <i>Plantes à risques</i> . Ed. Tec et Doc. Lavoisier. ISBN: 978-2-7430-9907-1. Eyad Sarf Ibrahim. 2007. Thesis. Isolation and characterization of pyridizidine alkaloids from <i>Echium geraniatum</i> For. (boraginaceae). The Faculty of Graduate Studies, Jordan University of Science and Technology.
<i>Elettaria cardamomum</i> (L.) Maton.	Zingiberaceae	Fruit	Essential oil: phenylpropenoids: e.g. methyl Eugenol (0.1%), and monoterpene etheroxide: 1,8-cineole (up to 51.3%)		Council of Europe. 2005. Active principles (constituents of chemical concern) contained in natural sources of flavonoids. Ed. Council of Europe Publishing. http://www.coe.int/t/e/social_consensus/doc_solubible_healthyflavonoidsubstancesActive%20principles.pdf
<i>Embellia</i> spp.	Primulaceae	Fruit	Genus in which species may contain the benzquinone embelin		Prakash AO et al. 1992. Antiplasmodial mechanism of action of Embelin in rats. <i>Phytother Res</i> 6:28-33. Gupta RS et al. 1989. Antispermatic effect of embelin, a plant benzquinone on male albino rats in rats in vivo and in vitro. <i>Contraception</i> 39: 307-320
<i>Ephedra</i> spp.	Ephedraceae	Aerial part	Genus in which species may contain phenylethylamine alkaloids: e.g. ephedrine, pseudoephedrine.		Burleton J. 2009. Pharmacognosy. (Phytochimie, Plantes médicinales). Ed. Tec & Doc. Lavoisier. Paris. 4ème édition. ISBN: 978-2-7430-1188-9
<i>Epidolium</i> spp.	Onagraceae	Aerial part	Genus in which some species may contain macrocyclic elaglantrins: oenotherin A and oenotherin B, considered responsible for the acetylcholinesterase or bromelase and 5-alpha-reductase present in prostate cells. Other compounds like flavonoids and steroids may contribute to the effect.		Ducuy B et al. inhibition of 5 alpha-reductase and aromatase by the elaglantrins oenotherin A and oenotherin B from <i>Epidolium</i> species. <i>Planta Med.</i> 63(2):111-4. Vitaleo A et al. 2003. Extracts of various species of <i>Epidolium</i> inhibit proliferation of human prostate cells. <i>Pharmacology</i> 69(2):79-87 Hernandez A, Bucari F. 1997. Studies of <i>Epidolium angustifolium</i> extracts on growth of accessory sexual organs in rats. <i>J. Ethnopharmacol</i> 55: 179-183.
<i>Epidium grandiflorum</i> C.Morren.	Berberidaceae	Whole plant	Flavonol glycoside: icaritin	Listed as Novel Food in the NF Catalogue	Wazuno M et al. 1989. Seasonal fluctuation of flavonol glycosides in <i>Epidium</i> species. <i>Yakugaku Zasshi</i> 109(4):271-272. Koga S et al. 1991. Studies on <i>Epidium</i> species: Flavonol glycosides and isozymes. <i>Biochemical Systematics and Ecology</i> 19(4): 315-318 Wazuno M et al. 1989. Seasonal fluctuation of flavonol glycosides in <i>Epidium</i> species. <i>Yakugaku Zasshi</i> 109(4):271-272. Koga S et al. 1991. Studies on <i>Epidium</i> species: Flavonol glycosides and isozymes. <i>Biochemical Systematics and Ecology</i> 19(4): 315-318
<i>Epidium sagittatum</i> (Siebold & Zucc.) Maxim.	Berberidaceae	Whole plant	Flavonol glycoside: icaritin (in fresh leaf: 0.013%)		Roth L., Daurerier M. and Korman K. 1994. <i>Complizen - Pflanzenzettel - Vorkommen Wirkstoffe</i> . Therapie. ecomed. ISBN: 3-509-64810-4.
<i>Equisetum palustre</i> L.	Equisetaceae	Aerial part	Piperidine alkaloids: e.g. palustrine (0.01-0.3%)	Reported toxicly on livestock	Kopp B, et al. 2004. 4H-Chromenone Glycosides from <i>Eranthis hyemalis</i> (L.) Salisbury. <i>Helv Chim Acta</i> 74: 611-616 Gupta RC (ed). 2007. <i>Veterinary toxicology: Basic and Clinical principles</i> . Elsevier Inc. ISBN: 978-0-12-517467-2
<i>Eranthis hyemalis</i> (L.) Salisb.	Ranunculaceae	Root	Chromenone glycosides: e.g. erantnin		Hasagawa T. 2005. Estimation of Cyanogenic Glycosides and Their Degradation Products in <i>Eranthis japonica</i> Steud. under Various Storage and Processing Methods. <i>Bulletin of the Public Health Laboratory of Chiba Prefecture</i> 26: 51-60.
<i>Erechtites</i> spp.	Compositae (Asteraceae)	Whole plant	Genus in which species may contain unsaturated pyridizidine alkaloids: e.g. senecionine, seneciphylline		Zhuang YF. 2002. Determine the quantity of amygdalasin in <i>Erechtites japonica</i> leaf by HPLC. <i>Shant Pharmaceutical Journal</i> 14(5): 64-65
<i>Eriobotrya japonica</i> (Thunb.) Lindl.	Rosaceae	Leaf and seed	Cyanogenic glycoside: amygdalin (0.06%)		Garrera P.M. 2003. Folk medicine and minor nutriment in the folk traditions of Central Italy (Marche, Anruzzo and Latium). <i>Filomat</i> 74: 516-544. Kartal M, et al. 2006. Terpene Saponins from <i>Eryngium campense</i> . <i>J. Natl. Prod.</i> 69: 1105-1108.
<i>Eryngium campense</i> L.	Apiaceae (Umbelliferae)	Aerial part	Essential oil from fresh herb (0.09%); Turanocoumarins: e.g. bergapten (0.014% in fruit); Polyenes from falcarinone type: e.g. falcarinon, falcaranol		Burleton J. 2005. <i>Plantes toxiques (Végétaux dangereux pour l'homme et les animaux)</i> . Ed. Tec & Doc. Lavoisier. Paris. 3ème édition. 618 pages. ISBN: 2-7430-0806-7
<i>Erythrina</i> spp.	Leguminosae	Aerial part	Genus in which species may contain benzyltetrahydroisquinoline alkaloids: e.g. erythrine, erysodine		Frome D., Pfander H.J. and Anton R. 2009. <i>Plantes à risques</i> . Ed. Tec et Doc. Lavoisier. ISBN: 978-2-7430-9907-1
<i>Erythrophloeum suaveolens</i> (Guill. & Perr.) Benth	Leguminosae (Fabaceae)	Bark and seed	Diterpenoid amides: e.g. cassaine		Burleton J. 2009. Pharmacognosy. (Phytochimie, Plantes médicinales). Ed. Tec & Doc. Lavoisier. Paris. 4ème édition. ISBN: 978-2-7430-1188-9
<i>Erythroxylum</i> spp.	Erythroxylaceae	Whole plant	Genus in which species may contain tropane alkaloids: e.g. cocaine	Occure found in 14 species (there are about 50-60 species)	Gartner S et al. 2005. Alkaloids from <i>Erythroxylum californica</i> and their capacity to inhibit binding of [³ H]-Hydroxy-2-(di-N-propylamino)seratin to 5-HT _{1A} receptors in vitro. <i>J. Nat. Prod.</i> 69: 432-435. Proença da Cunha A, et al. 2003. <i>Plantas e Produtos Vegetais em Filoterapia</i> . Fundação Calouste Gulbenkian, Lisboa. ISBN: 978-972-31-1010-4.
<i>Eschscholzia californica</i> Cham.	Papaveraceae	Aerial part			Burleton J. 2009. Pharmacognosy. (Phytochimie, Plantes médicinales). Ed. Tec & Doc. Lavoisier. Paris. 4ème édition. ISBN: 978-2-7430-1188-9
<i>Eucalyptus</i> spp.	Myrtaceae	Leaf	Genus in which species may contain monoterpene etheroxide: 1,8-cineole		Burleton J. 2009. Pharmacognosy. (Phytochimie, Plantes médicinales). Ed. Tec & Doc. Lavoisier. Paris. 4ème édition. ISBN: 978-2-7430-1188-9
<i>Eunymus atropurpureus</i> Jacq.	Celastraceae	Whole plant	Cardiotoxic glycosides (digitals-like) in fruit (seed): e.g. eunomside, eunobeside, eunomnoside, sesquiterpene alkaloids (0.1%), e.g. evonine, eozonine, evonine		Meleiro P, et al. 2000. A Short Review on Cardiac Steroids and their Antigonandrine Analogs. <i>Molecules</i> 5: 51-81
<i>Eunymus europaeus</i> L.	Celastraceae	Whole plant	Cardiotoxic glycosides (digitals-like) in fruit (seed): e.g. eunomside, eunobeside, eunomnoside, sesquiterpene alkaloids (0.1%), e.g. evonine, eozonine, evonine		Meleiro P, et al. 2000. A Short Review on Cardiac Steroids and their Antigonandrine Analogs. <i>Molecules</i> 5: 51-81
<i>Eupatorium</i> spp.	Compositae (Asteraceae)	Whole plant	Genus in which species may contain unsaturated pyridizidine alkaloids: e.g. supinine, rindene		Malerio P et al. 2000. A Short Review on Cardiac Steroids and Their Antigonandrine Analogs. <i>Molecules</i> 5: 51-82
<i>Euphorbia</i> spp.	Euphorbiaceae	Whole plant	Genus in which species may contain diterpene-esters (phorbol esters) in the latex: e.g. tigliane, ingenane and daphnane types	These compounds are found in two families: Euphorbiaceae & Thymelaeaceae. Some of these esters are cocarcinogenic agents.	Frome D., Pfander H.J. and Anton R. 2009. <i>Plantes à risques</i> . Ed. Tec et Doc. Lavoisier. ISBN: 978-2-7430-9907-1

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This compendium lists in alphabetical order botanicals without any judgment on whether these are suitable or not suitable for food applications in Europe. This compendium is part of a preliminary work undertaken by EFSA to harmonise the methodology across its panels for assessing the safety of botanicals and botanical preparations used in food and food supplements. Without prejudice to the existing legal framework, such compendium has no legal status and may not be used as support or evidence in any disagreement or dispute pertaining to the legal classification of products or substances. This compendium is a living document and is therefore open for additional contributions and comments.

Botanical name	Family	Parts of plants of possible concern	Chemical of concern	Remarks on toxic/adverse effect(s) not known to be related to the identified chemical(s) of concern	Specific References
<i>Eurycoma longifolia</i> Jack	Simarubaceae	Root	Nonterpenoid quassinoids, e.g. eurycomanone, eurycomalacton, eurycomanol, iridic alkaloids, beta catholic alkaloids and indolomonoterpenic alkaloids, e.g. canthin-9-one		Warab NA et al. 2010. The effect of <i>Eurycoma longifolia</i> Jack on spermatogenesis in estrogen-treated rats. <i>Chines Med J</i> 35(1): 93-8 Shird AN et al. 2010. The anti-osteoporotic effect of <i>Eurycoma longifolia</i> in aged ovariectomized rat model. <i>The journal of Traditional Chinese Medicine</i> , 15-5 Council of Europe. 2000. Natural sources of flavonoids. Report No. 1. Council of Europe Publishing. ISBN 978-92-871-4324-2.
<i>Eremia purpuria</i> (L.) Ach. <i>Erodia rufescens</i> (A.Juss.) Hook. f. ex Benth.)	Rubiaceae	Lichen Fruit	Essential oil: bicyclic monoterpene, e.g. alpha and beta thuyones (about 10%), camphor Indo-pyridic-quinazoline alkaloids, e.g. evodiamine, rutecarpine		Shoji N, et al. 1986. Isolation of evodiamine, a powerful cardiotonic principle, from <i>Erodia rufescens</i> Benth. (<i>Rubiaceae</i>). <i>J Pharm Sci</i> 75(6): 612-3 Konomi T, et al. 2003. Saponin-biotin type diterpenes from <i>Erodia rufescens</i> Benth. <i>PHYTOCHEMISTRY</i> 64(4): 835-840
<i>Excoecaria agallocha</i> L.	Euphorbiaceae	Whole plant	Diterpene esters (abudane type), e.g. excoecarins	Contact with plant can cause temporarily blindness. Used in fertility regulations and having uterotropic activity	Nelson E, et al. 1989. Genotoxic effects of substrate treatments with wood dust extracts on the nasal epithelium of rats: assessment by the micronucleus and 32P-postlabelling. <i>Arch Toxicol</i> 62(8): 586-589
<i>Fagus sylvatica</i> L.	Fagaceae	Fruit and wood	Furic oxalates (2.95%)	Wood dust possibly mutagenic activity.	Kajimoto T et al. 1989. Sesquiterpene and diene derivatives from <i>Fagus sylvatica</i> . <i>Phytochemistry</i> 28: 1761-1763 Tisserand, R. and Balacs, T., 1995. <i>Essential Oil Safety: A Guide for Health Care Professionals</i> . Churchill Livingstone, Edinburgh. ISBN: 0443052603.
<i>Ferula assu-foetida</i> L.	Apiaceae (Umbelliferae)	Root		Oleo gum resin: sesquiterpene coumarins, e.g. asacoumarin A, B. One case of methaemoglobinemia described in an infant.	Kelly KJ et al. 1994. Methemoglobinemia in an infant treated with the folk remedy glicened astofoetida. <i>Pediatrics</i> 73: 717-719
<i>Ferula hermonis</i> Boiss.	Apiaceae (Umbelliferae)	Root and seed	Daucane sesquiterpene esters from root resin, ferulidin (ferulidol-P-hydroxybenzoate), ferulidin (ferulidol benzoate) and the sesquiterpene alcohol ferulidol (jascokeanadiol)	Reproductive toxicity and infertility effect of <i>Ferula hermonis</i> in mice Oil extract from the seeds administered acutely in rats enhances erectile function; chronic administration induces toxic effects (decreased body weight, hepatomegaly and atrophic testis)	Merza H Homayri et al. 2002. Reproductive toxicity and infertility effect of <i>Ferula hermonis</i> extracts in mice. <i>Therapeutics</i> 57 (9): 2247-2256. El-Thaber TS et al. 2001. <i>Ferula hermonis</i> "Zalhuhr" and enhancing erectile function in rats: efficacy and toxicity study. <i>Int J Impot Res</i> 13: 247-251. Hilmi C, et al. 2007. Evaluation of the antibacterial activities of <i>Ferula hermonis</i> (Boiss.). <i>Lebanese Science Journal</i> 8 (2): 135-150
<i>Ficaria verna</i> L.	Moraceae	Whole plant	Furanocoumarins from latex, e.g. psoralen and bergapten		Tisserand, R. and Balacs, T., 1995. <i>Essential Oil Safety: A Guide for Health Care Professionals</i> . Churchill Livingstone, Edinburgh. ISBN: 0443052603. Hagers Handbuch der Pharmazwissenschaften Paris 1998. Springer Verlag. ISBN 3-540-52688-9
<i>Foeniculum vulgare</i> Mill. <i>Foeniculum vulgare</i> Mill. ssp. <i>piperitum</i> (Utrac Coutinho)	Apiaceae (Umbelliferae)	Aerial part	Essential oil from the aerial part: phenylpropanoids, e.g. trans-anethole, methylchavicol (2.3-4.9%) Essential oil from the urine seed: methylchavicol (11.9-56.1%) Essential oil from ripe seed: methylchavicol (61.8%)		Council of Europe. 2008. Natural sources of flavonoids. Report No. 3. Council of Europe Publishing. ISBN 978-92-871-6422-3. García-Alvarez N et al. 2000. Chemical composition of Ferula oil. <i>Foeniculum vulgare</i> Miller, from Spain. <i>Journal of Essential Oil Research</i> , 12(2): 159-162.
<i>Foeniculum vulgare</i> Mill. ssp. <i>vulgare</i> var. <i>vulgare</i>	Apiaceae (Umbelliferae)	Fruit	Essential oil from the aerial part: phenylpropanoids, e.g. trans-anethole, methylchavicol Essential oil from the seed: methylchavicol (3.5-12%)		Council of Europe. 2005. Active principles (constituents of chemical concern) combined in natural sources of flavonoids. Ed. Council of Europe Publishing http://www.coe.int/cesocoll_consensusdoc_sopublic_health/Flavonoids_substancesActive%20in%20natural%20sources.pdf
<i>Foeniculum vulgare</i> Mill. ssp. <i>vulgare</i> var. <i>duice</i> (Mill.) Batt. & Trab.	Apiaceae (Umbelliferae)	Fruit	Essential oil from the seed: phenylpropane derivative, e.g. trans-anethole, methylchavicol (1.5-3.1%)		Council of Europe. 2008. Natural sources of flavonoids. Report No. 3. Council of Europe Publishing. ISBN 978-92-871-6422-3. Council of Europe. 2005. Active principles (constituents of chemical concern) combined in natural sources of flavonoids
<i>Fritillaria</i> spp.	Liliaceae	Bulb	Genus in which species may contain steroidal and isosteroidal alkaloids, e.g. peritrisine, verticine, verticrone, imperatrine, isovericine, elatrine Benzylisoquinoline alkaloids (protoberberines), e.g. protopine (38%), sinactine, cryptopine, fumariline and sanguinarine Genus in which species may contain isoquinoline alkaloids (Amaryllidaceae alkaloids), e.g. galantamine, lycorine		HL P, et al. 2001. Determination of isosteroidal alkaloids and glucosides in <i>Fritillaria</i> by HPLC-ELSD. <i>Journal of Chromatography</i> , 909 (2): 207-214 Gorbunov N.P. et al. 1977. Preparation and the antipruritic activity of the total alkaloids of <i>Fumaria officinalis</i> L. <i>Pharm. Chem.</i> 11(5): 640-642 Sidiqimov B, et al. 2003. Galantamine distribution in Bulgarian <i>Galanthus</i> spp. <i>Pharmazie</i> 58(12): 935-936
<i>Fumaria officinalis</i> L.	Papaveraceae	Aerial part			Burleton J 2005. <i>Plantes toxiques (Végétaux dangereux pour l'homme et les animaux)</i> . Ed. Tec & Doc. Lavoisier, Paris, 3ème édition, 618 pages, ISBN : 2-7430-0806-7. Raschka, H.R. et al. 2008. Acute and subchronic oral toxicity of <i>Fumaria officinalis</i> in rats. <i>J Ethnopharmacol</i> 116 (1): 21-26.
<i>Galanthus</i> spp.	Amaryllidaceae	Aerial part			Jacquemond-Coleil I, et al. 2001. Identification of the alkaloids of <i>Galipaea officinalis</i> by gas chromatography-mass spectrometry. <i>Phytochem Anal</i> 12: 312-319. Rakotonjan JH, et al. 1998. Alkaloids from <i>Galipaea officinalis</i> . <i>Pharma medica</i> 64(6): 762-763.
<i>Gallega officinalis</i> L.	Leguminosae (Fabaceae)	Aerial part	Quandine derivatives, e.g. gallegine (in herb 0.1%-0.3%, in seed up to 0.5%), peganine		Council of Europe. 2000. Natural sources of flavonoids. Report No. 1. Council of Europe Publishing. ISBN 978-92-871-4324-2. Tisserand, R. and Balacs, T., 1995. <i>Essential Oil Safety: A Guide for Health Care Professionals</i> . Churchill Livingstone, Edinburgh. ISBN: 0443052603
<i>Galipaea officinalis</i> Hancock (<i>Cuscuta officinalis</i> (Harcock) Engl.)	Rubiaceae	Bark and wood	Tetrahydroquinoline alkaloids (40%), e.g. angusturene, galpepine, cusparepine, galiphrine and quinine alkaloids, e.g. galipine and turoquinoline alkaloids, e.g. maculosidine		Jayaprakash G.K. and Sivarajah K.K. 2002. Determination of organic acids in leaves and fruits of <i>Galipaea officinalis</i> (Desr.) by LC. <i>J Pharmaceut Biomed</i> 26 (2): 379-384
<i>Galium odoratum</i> (L.) Scop. (<i>Asperula odorata</i> L.)	Rubiaceae	Aerial part	Coumarin (0.7-1.7% in dried herb)	Coumarin in dried herb: 1.06% in April/May, 0.44-0.93% in August	Saito M et al. 2005. High dose of <i>Galipaea officinalis</i> is effective in suppressing fat accumulation in developing male Zucker obese rat, but highly toxic to the testis. <i>Food Chem Toxic</i> 43: 411-419.
<i>Galipaea camptogla</i> Desr. See <i>Galipaea gummi-gutta</i> (L.) Roxb.	Rubiaceae				Brann H, Frohe D. 1987. Heilpflanzen-Lexikon für Ärzte und Apotheker: 5. Auflage. Stuttgart: Galesy Fischer Verlag. ISBN 3437110713, 9783437110719 Asano J et al. 1996. Cytotoxic xanthones from <i>Galipaea officinalis</i> . <i>Phytochemistry</i> 41(3): 815-820.
<i>Galipaea gummi-gutta</i> (L.) Roxb. (<i>Galipaea camptogla</i> Desr.)	Rubiaceae (Guttiferae)	Whole plant	(-) Hydroxydic acid (HCA) from fruit. Gum resin from bark		Council of Europe. 2000. Natural sources of flavonoids. Report No. 1. Council of Europe Publishing. ISBN 978-92-871-4324-2. Tisserand, R. and Balacs, T., 1995. <i>Essential Oil Safety: A Guide for Health Care Professionals</i> . Churchill Livingstone, Edinburgh. ISBN: 0443052603
<i>Galipaea hainburyi</i> Hook.f.	Clusiaceae (Guttiferae)	Whole plant	Bark gum resin, polyphenylated xanthones, e.g. gambogic acid, isogambogenin, isomerenillicol, (-) Hydroxydic acid (HCA) from fruit		Jayaprakash G.K. and Sivarajah K.K. 2002. Determination of organic acids in leaves and fruits of <i>Galipaea officinalis</i> (Desr.) by LC. <i>J Pharmaceut Biomed</i> 26 (2): 379-384
<i>Galipaea indica</i> (Thou.) Choisy	Clusiaceae (Guttiferae)	Fruit and leaf	(-) Hydroxydic acid (leaf: 4.1-4.6%, fruit: 10.3-12.7%)		Jayaprakash G.K. and Sivarajah K.K. 2002. Determination of organic acids in leaves and fruits of <i>Galipaea officinalis</i> (Desr.) by LC. <i>J Pharmaceut Biomed</i> 26 (2): 379-384

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<i>Garcinia morella</i> Desr.	Clusiaceae (Guttiferae)	Whole plant	Bark gum resin (gamboge), tetrahydropyrid xanthones: morolic acid, isomorolic acid, alpha- and beta-guttiferins and derivatives... Seed coat: morillin, beta- and alpha-guttiferin		Sarithanam K. and Rao P.L. 1968. Antibiotic principles of <i>Garcinia morella</i> . Part XII - Characterization of beta- and alpha-guttiferins as cardiac glycosides of gamboge and seed coat of <i>G. morella</i> . Indian J. Exp. Bot. 6, 158-169. Khare C.P. 2007. Indian Medicinal Plants, Springer Verlag, ISBN: 978-0-387-70637-5. Tomlinson B. et al. 2000. Toxicity of complementary therapies: an eastern perspective. J. Clin. Pharmacol. 40, 451-456.
<i>Gaultheria fragrantissima</i> Vahl.	Ericaceae	Leaf	Essential oil from leaf (0.7-0.8%), salicylic acid derivatives: methylsalicylate (99.6%)		Baruah A.K.S. and Bhargat S.D. 1976. Oil of Indian wintergreen. Indian J. Pharmacy. 38: 56-57.
<i>Gaultheria procumbens</i> L.	Ericaceae	Whole plant	Essential oil from leaf: methyl salicylate (98%)		Bruneton J. 2009. Pharmacognosie, (Phytochimie, Plantes médicinales), Ed. Tec & Doc. Lavoisier, Paris, 4ème édition, 1269 pages, ISBN: 978-2-7430-1188-8. Tisserand R. and Balacs T. 1995. Essential oil safety, A Guide for Health Care Professionals, Churchill Livingstone, Edinburgh, ISBN: 0432052803
<i>Geissospermum vellosii</i> Allem.	Apocynaceae	Bark	Indole and beta-carboline alkaloids: e.g. geissospermine, flavopipterine, vellosine, geissoschizoline (and derivatives), geissoschizoline N4-oxide and 1,2-dihydrogeissoschizoline), pauserperidine		Mbenanku F. et al. 2011 Isolation and structural elucidation of indole alkaloids from <i>Geissospermum vellosii</i> by mass spectrometry. J. Chromatogr. B. Analyt. Technol. Biomed. Life Sci. 2011 Dec 26; [Epub ahead of print] Bemis D.L. et al. 2009 beta-carboline alkaloid-enriched extract from the amazonian rain forest tree <i>paua</i> <i>peruvia</i> suppresses prostate cancer cells. J. Soc. Integr. Oncol. 7(2), 59-65. Aravujo JQ et al. 2011 Docking of the alkaloid geissospermine into acetylcholinesterase: a natural scaffold targeting the treatment of Alzheimer's disease. J. Mol. Model. 17(6), 1401-1412
<i>Geissemium</i> spp.	Gesneriaceae	Whole plant	Genus in which species may contain indol- and oxindolalkaloids: e.g. geissemine, sempervivine		Frome D., Pfander H.J. and Anton R. 2009. Plantae 4 inquest. Ed. Tec et Doc Lavoisier, ISBN: 978-2-7430-0907-1.
<i>Genista tinctoria</i> L.	Leguminosae (Fabaceae)	Aerial part	Quinoline alkaloids: e.g. anagyrine, oxisine (0.7-0.8%), sparteine, lupanine, lupinine		Gazalier A.M. et al. 1991. Isolation, analysis, biosynthesis, and modification of the alkaloid oxisine. Chem. Nat. Compd. 27(3), 259-269. Wink M. 1986. Acquired toxicity - the advantages of specializing on alkaloid-rich lupins to <i>Macrospilton albivittae</i> (Aphididae). Naturwissenschaften 73, 210-212. EMEA Committee for Veterinary Medicinal Products. 1999. <i>Ginkgo biloba</i> . Lesinier E. and Drevwe C. 2010. <i>Ginkgo biloba</i> and ginkgol. J. Nat. Prod. 73, 86-92.
<i>Ginkgo biloba</i> L.	Ginkgoaceae	Leaf and seed (ovule)	Alkylphenols from leaf: ginkgolide acids: e.g. bilobol, cardanol, cardols and ginkgol. ginkgolatoxin.	Leaf: Sesquiterpene lactones: bilobalide... Diiterpene lactones: 0.06-0.23% ginkgolides.	Ovule: human intoxication reported in Japan with some lethal outcomes. Carried and boiled seeds contain only 1% of the content in fresh seeds. Also roasted seeds contain ginkgol. Symptoms of intoxication with the seeds: vomiting, seizures and unconsciousness
<i>Glaucium corniculatum</i> (L.) Rudolph <i>spp. refractum</i> (Nab) Cullen	Papaveraceae	Aerial part	Isosquinoline alkaloids: e.g. pectinoline, glutifidine, dehydrocorydine, (+)-norbracteoline, thalictrodine, bulbocapnine (1.2%), diacetyline (0.7%), protopine (0.42%), corydine (<0.1%), glaucine (<0.1%), o-allylcorydopine (<0.1%)		WHO. 1999. WHO monographs on selected medicinal plants. Volume 1. World Health Organization, ISBN 92.4.154517.8
<i>Glaucium flavum</i> Crantz	Papaveraceae	Whole plant	Isosquinoline alkaloids (aporphine alkaloids): e.g. glaucine (from under detection level to over 3.6%)		Khrushchinskii L.G. and Vachnashvitz V.Y. 2000. Alkaloids of <i>Glaucium corniculatum</i> and <i>G. flavum</i> growing in Georgia. Chem. Nat. Compd. 36(2), 225-226. Sharief A. et al. 1985. Alkaloids of Papaveraceae. XII. Alkaloids of <i>Glaucium corniculatum</i> subspecies <i>refractum</i> . J. Nat. Prod. 48(5), 855-856. Shamma M. and Guiraudou H. 1985. Aporphinoid alkaloids. Nat. Prod. Rep. 2, 227-233.
<i>Glechoma hederacea</i> L.	Lamiaceae (Labiatae)	Aerial part	Pyridoline alkaloids linked with a tropane-like skeleton: e.g. hederacines A and B. Essential oil from flowering aerial part: monoterpenic etheroxide: 1,8-cineole (1.9 - 4.6%)	The plant has been reported to cause illness and death in horses and cattle. The potential toxic constituents have not been identified.	Dargatzis P.I. et al. 2008. Detection of the pharmaceutical agent glaucine as a recreational drug. Eur. J. Clin. Pharmacol. 64, 553-554. Peleli B. et al. 1986. Alkaloid content in various chemotypes of <i>Glaucium flavum</i> from Israel. Phytocchemistry 27(4), 1021-1024. Pellino V. et al. 2010. Alkaloids from <i>Glaucium flavum</i> from Sardinia. Nat. Prod. Research. 24(11), 1033-1035.
<i>Globularia alypum</i> L.	Plantaginaceae	Leaf and root		Increased embryo resorption observed in pregnant rats after intragastric administration of 800 mg/kg of an ethanolic extract of the dried leaves from dry 1-30 pregnant rats.	Cooper M.R. and Johnson A.W. 1998. Poisonous plants and fungi in Britain, Africa and human poisoning. The Stationery Office, ISBN 0-11-242381-5. Kumarasamy Y. et al. 2003. Isolation, structure elucidation and biological activity of hederacine A and B. two unique alkaloids from <i>Glechoma hederacea</i> . Tetrahedron 59, 6403-6407. Mackie D. et al. 2005. Chemical composition of essential oils of <i>Glechoma hederacea</i> L. growing wild in Virginia district. Chemia. 16(3-4), 47-50. Reddovic N. et al. 2010. Volatile constituents of <i>Glechoma hirsuta</i> Waldst. & Kl. and <i>Glechoma hederacea</i> L. (Lamiaceae) Bal. Chem. Soc. Ethnop. 24(1), 67-76.
<i>Gloriosa spp.</i>	Colechicaceae	Whole plant	Genus in which species may contain topolone alkaloids: e.g. colchicine		Elbehika A. et al. 2000. Fetotoxic potentials of <i>Globularia arabica</i> and <i>Globularia alypum</i> (Dobuniaceae) in rats. J. Ethnopharmacol. 72, 215-219. Natarumukwe D. et al. 1994. Quantitative determination of colchicine in <i>Gloriosa simplex</i> Liliaceae of Rwanda Central Africa. Pharm. Medicines et Phytoterapie. 18(1), 24-27.

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<i>Glycine max</i> (L.) Merr.	Leguminosae (Fabaceae)	Seed	Soybean agglutinin; (N-acetylglucosamine-specific lectin), protease inhibitors and other toxic proteins. Total isoflavones 945-4208 µg/g a.s. 67-516 µg/g daidzin, 91-1079 µg/g genistin, 12-177 µg/g glycin, 217-788 µg/g malyngaldin, 43-158 µg/g malyngelychin, 64-2446 malyngenisin, 4-3-266 µg/g genistein.		Becker-Ritt A. B. et al. 2004. Antinutritional and/or toxic factors in soybean (<i>Glycine max</i> (L.) Merr.) seeds: comparison of different cultivars selected in the southern region of Brazil. J. Sci. Food Agric. 84, 253-270. BfR (Federal Institute for Risk Assessment). 2007. Isolated isoflavones are not without risk. http://www.bfr.bund.de/cm/349/isolated_isoflavones_are_not_without_risk.pdf . Mendonça N. et al. 2010. Fast analysis of isoflavones by HPLC-performance liquid chromatography using a column packed with fused-core particles. Talanta 82, 1986-1994. Palisau H.B. and Jefferson W. 2010. The pros and cons of phytoestrogens. Front. Neuroendocrin. 31, 400-419. Sun J.M. et al. 2011. Rapid HPLC method for determination of 12 isoflavone components in soybean seeds. Agric. Sci. China 10(1), 70-77.
<i>Glycyrrhiza glabra</i> L.	Leguminosae (Fabaceae)	Root	Phenylpropanoids: e.g. methylchavicol in unspecified quantities Terpene saponins with glycyrrhizin (potassium and calcium salts of glycyrrhizic acids) as major components.		EMA/HMPC. 2005. Public statement on the use of herbal medicinal products containing estragole. EMA/HMPC/1372/22005 Scientific Committee on Food (SCF). 2003. Opinion of the Scientific Committee on Food on Glycyrrhizic acid and its ammonium salt SC/FCS/ADD/EDU/225 Final 10 April 2003. Wichtl M. 2002. Teedrogen und Phytopharmaka. Ein Handbuch für die Praxis auf wissenschaftlicher Grundlage. Wissenschaftliche Verlagsgesellschaft mbH. ISBN 3-8047-1694-X. WHO. 1999. WHO monographs on selected medicinal plants. Volume 1. World Health Organization. ISBN 92.4.154517.8. Scientific Committee on Food (SCF). 2003. Opinion of the Scientific Committee on Food on Glycyrrhizic acid and its ammonium salt SC/FCS/ADD/EDU/225 Final 10 April 2003. Kulkarni V. and Vatsrajanyulu A. 2010. Toxicological studies on aqueous extract of <i>Gmelina arborea</i> in rodents. Pharm. Biol. 48(12), 1413-1420. Sahu R. et al. (2010) In vivo micronucleus assay of <i>Gmelina arborea</i> Roxb (Gambhari) extract. J. Adv. Pharm. Tech. Res. 1, 22-29. Percy R.G. et al. 1996. Seed gossypol variation within <i>Gossypium hirsutum</i> L. cotton. Crop. Sci. 36, 193-197. Wales G.M.H. et al. 1998. Gossypol: reasons for its failure to be accepted as a safe, reversible male antifeedant. <i>Insect. J. Anim. Ecol.</i> 21, 8-12. Kaya G. I. and Mezig M.F. 2008. Quantitative determination of cucurbitacin E and cucurbitacin I in homoeopathic mother tincture of <i>Gnathia officinalis</i> by HPLC. Pharmazie 63(12), 651-653. Stum S. and Stuppler H. 2000. Analysis of cucurbitacins in medicinal plants by high-pressure liquid chromatography-mass spectrometry. Phytochem. Anal. 11, 121-127.
<i>Glycyrrhiza uralensis</i> Fisch. ex DC.	Leguminosae (Fabaceae)	Root	Terpene saponins with glycyrrhizin (potassium and calcium salts of glycyrrhizic acids) as major components.		
<i>Gmelina arborea</i> Roxb.	Lamiaceae (Labiatae)	Leaf		An aqueous extract of the leaves was tested for genotoxicity in the micronucleus test. P.o. doses of 0, 286 and 667 mg/kg b.w. of the extract was given to mice. A significant (P < 0.01) increase in % micronuclei was seen with both doses indicating a mutagenic potential of the leaves.	
<i>Gossypium</i> spp.	Malvaceae	Root bark and seed	Genus in which species may contain gossypol, a terpenoid aldehyde		
<i>Gnathia officinalis</i> L.	Plantaginaceae	Whole plant	Oxygenated tetracyclic terpenes: e.g. cucurbitacin I-glucofide, cucurbitacin E-glucofide, glicofide.		
<i>Grewia</i> spp.	Malvaceae	Bark, flower and shoot	Genus in which species may contain harmaline alkaloids (beta-carbolines)		
<i>Gnifonia simplicifolia</i> (M. Vahl x DC.) Baill.	Leguminosae (Fabaceae)	Seed	5-Hydroxyxylopholan derivatives: 20.83% on a fresh weight basis		Alkaloids present in <i>Grewia tilaefolia</i> , <i>Grewia hirsuta</i> , <i>Grewia asiatica</i> , <i>Grewia laxa</i> . In <i>G. bicolor</i> : harmalin, 6-methoxyharmalin, 6-hydroxyharmalin.
<i>Gnindelia squarrosa</i> (Pursh) Dunal	Compositae (Asteraceae)	Aerial part		One study reported sheep mortality after grazing <i>G. squarrosa</i> . Further study showed that the plant concentrates selenium from the soil, up to toxic levels.	
<i>Guaiacum officinale</i> L.	Zygophyllaceae	Bark	Resin (gum) from the bark: 15% petroleum ether soluble compounds: lignans (+)-guaiaric acid, meso-dihydroguaiaric acid and meso-nordihydroguaiaric acid; 70% ether soluble compounds: other lignans such as dehydroguaiaric acid; Juroquialcin/alpha-guaiacolic acid and its 4-methyl ether, various tetrahydrofurans		
<i>Guarea</i> spp.	Melastomaceae	Seed		Genus in which seed and fruit of some species may contain undefined hallucinogenic alkaloids The bark decoctions reported to have an abortifacient and emetic effect.	
<i>Guaiaria gaumeri</i> Greenm.	Annonaceae	Bark	Phenylpropanoids: e.g. alpha-santonine		
<i>Gymnosporia senegalensis</i> (Lam.) Loes.	Celastraceae	Root bark		Genotoxic effect seen with a dichloromethane extract (lipid fraction) of the root bark that induced micronuclei in human white blood cells. Toxic effects with the extract in Comet assay.	
<i>Madecarus senegalensis</i> (Lam.) Exell					
<i>Gynocardin odorata</i> R.Br.	Achariaceae	Leaf and seed	Cyanogenic glycosides: e.g. gynocardin		
<i>Hagezia abyssinica</i> J.F. Gmel.					
<i>See Braveia antioquiensis</i> Kunth.					
<i>Hammamelis virginiana</i> L.	Hammamelidaceae	Bark and leaf	Essential oil from the fresh leaf: (0.01 - 0.05%); phenylpropanoid: e.g. safrole (content max 0.2% of the volatile oil).	Leaf and bark up to 10% tannins. Hydrolysable tannins used at high doses and over a long period may have a negative impact on liver	

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Handroctonus heptaphyllus (Vahl) Matis (<i>Tabebuia heptaphylla</i> (Vahl.) Toledo, (<i>Tabebuia</i> sp. (K. Schum.) Standl.), <i>Tecoma</i> sp. (K. Schum.)	Bignoniaceae	Wood	Naphthoquinones, e.g. Iapachanol (Iapachtonone), Iapachol, Ulymans	Iapachol induced dose-dependently a clastogenic effect in vivo in rats in the micronucleus and chromosome aberration assays.	Felicio A.C. et al. 2002. Fetal growth in rats treated with Iapachol. <i>Contraception</i> 66, 289-293. Guerra M.O. et al. 1999. Intereceptive effect of Iapachol in rats. <i>Contraception</i> , 60, 305-307. Guerra M.O. et al. 2001. Toxicology of Iapachol in rats, embryofetally. <i>Rev. Bras. Biol.</i> 61(1), 171-174. Maistro E.L. et al. 2010. Iapachol induces clastogenic effects in rats. <i>Planta Medica</i> 76(9), 858-862. Schmieda-Hitschmann G. and Papastegou F. 2003. Naphthoquinone derivatives and Iignans from the Paraguayan crude drug "Iay" py'ã (<i>Tabebuia heptaphylla</i> , Bignoniaceae). <i>Z. Naturforsch.</i> 58c, 495-501.
Harungana madagascariensis Lam. ex Pol. (<i>Harungia madagascariensis</i> (Lam. ex Pol.) Chrys.)	Hypericaceae	Root	Prenylated polyphenolic anthranoids, e.g. harungadagascarin A and B	Increased levels of progesterone and estradiol-17β were found in female mice and increased testosterone level was found in male mice receiving drinking water with 5 g powdered root/100 ml drinking water.	Nishimoto N. et al. 1984. Pteridoses and nonterpenoid saponins from <i>Puffia paniculata</i> . <i>Phytochemistry</i> 23(1), 139-142. Oshima M. and Gu Y. 2003. <i>Puffia paniculata</i> -induced changes in plasma estradiol-17β, progesterone and testosterone levels in mice. <i>J. Reprod. Dev.</i> 49(2), 175-180.
Hebanthe eriantha (Polk.) Pedersen (<i>Puffia paniculata</i> (Mart.) Kuntze, <i>Gomphena paniculata</i> (Mart.) Moq., H. <i>paniculata</i> Mart.)	Amaranthaceae (Chenopodiaceae)	Root			Council of Europe. 2007. Natural sources of Flavonurins. Report No. 2. Council of Europe Publishing. ISBN 978-92-871-6156-7. Tisserand, R. and Balacs, T., 1995. Essential oil safety. A Guide for Health Care Professionals. Churchill Livingstone, Edinburgh. ISBN 043205263
Hedera pulegioides (L.) Pers.	Lamiaceae (Labiatae)	Aerial part	Essential oil: monocyclic monoterpene ketone, e.g. pulegone 30-80%, bicyclic monoterpenes: e.g. menthofuran and monoterpene etheroxide, 1,8-cineole		Cooper M.R. and Johnson A.W. 1998. Poisonous plants and fungi in Britain. Animal and human poisoning. The Stationery Office, ISBN 0-11-242981-5. Förme D., Pfander HJ and Anton R. 2009. <i>Plantes à risques</i> , Ed. "Ter et Doc-Lavoisier", ISBN: 978-2-7430-0907-1
Hedera helix L.	Araliaceae	Aerial part	Leaf: terpenoid saponins (2.5%-5.7%), e.g. alpha-hederin.	Inotoxication caused by the fruits ('berries').	Council of Europe. 2005. Active principles (constituents of chemical concern) contained in natural sources of flavonurins. Ed. Council of Europe Publishing. http://www.coe.int/t/tacon/conv/conv2005-06-20public_health/flavonurins_substancesActive20principles.pdf
Hedyotium flavum Roxb.	Zingiberaceae	Rhizome	Essential oil from rhizomes: monoterpenes: e.g. 1,8-cineole (up to 42%)		Rothier A. 1990. Alkaloids of <i>Henna montana</i> . <i>Phytochemistry</i> 29(5), 1683-1686. Rumala C.S. et al. 2008. Alkaloids from <i>Henna salicifolia</i> . <i>Phytochemistry</i> , 69, 1756-1762.
Hemina spp.	Lythraceae	Leaf	Genus in which species may contain biphenylquinolizidine lactone alkaloids, e.g. dehydrocodeine, hemidine, and phenylquinolizidine ester alkaloids, e.g. apresoline.	Alkaloids present in: <i>Hemina salicifolia</i> , <i>H. myrtifolia</i> , and <i>H. montana</i>	Cavalli J.F. et al. 2001. Constituents of the essential oil of six <i>Helichrysum</i> species from Madagascar. <i>Flavour Fragr. J.</i> 16, 253-256. Molterbeck S. et al. 1997. Chemical composition and analyses of enantiomers of essential oils from Madagascar. <i>Flavour Fragr. J.</i> 12, 63-69.
Helichrysum gymnoccephalum Humbert (<i>Asieraceae</i>)	Compositae (Asteraceae)	Aerial part	Essential oil: monoterpene etheroxide: 1,8-cineole (60 to 68%)		Idaomar M. et al. 2002. Cytotoxicity and antigenotoxicity of some essential oils evaluated by wing spot test of <i>Drosophila melanogaster</i> . <i>Mol. Res.</i> 5(15), 61-68. Blanchin A. et al. 2003. A comparative study of volatile constituents of two <i>Helichrysum italicum</i> (Rohb.) Guss. Don fil subspecies growing in Corsica (France), Turkey and Sardinia (Italy). <i>Environ. Fragr. J.</i> 18, 487-491
Helichrysum italicum (Rohb.) Guss.	Compositae (Asteraceae)	Aerial part	Essential oil from flower: monoterpene etheroxide: 1,8-cineole (0.3 to 1%)		Förme D., Pfander H.J. and Anton R. 2005. <i>Plantes à risques</i> , Ed. "Ter et Doc-Lavoisier", ISBN: 978-2-7430-0907-1
Heliolepis spp.	Borraginaceae	Whole plant	Genus in which species may contain unsaturated pyrrolizidine alkaloids, e.g. heliotrine, cynoglossine		Dalabeez Plants poisonous to livestock, Cornell University (http://www.Ansi.com/edu/PlantsPoisonousToLivestock) Cornell.edu/PlantsPoisonousToLivestock
Helioscopus spp.	Ranunculaceae	Aerial part	Genus in which species may contain cardiac glycosides: butadienolides: e.g. helioscophin		Byrleton J. 2005. <i>Plantes toxiques (Végétaux dangereux pour l'homme et les animaux)</i> , Ed. "Ter & Doc-Lavoisier, Paris, 5ème édition, ISBN: 2-7430-0905-7
Heptatica nobilis Schreb. (<i>Aionome heptatica</i> L.)	Ranunculaceae	Aerial part	Unsaturated lactone: protocarnomin	Protocarnomin only present in fresh herb	Cooper M.R. and Johnson A.W. 1998. Poisonous plants and fungi in Britain. Animal and human poisoning. The Stationery Office, ISBN 0-11-242981-5. Förme D. and Pfander H.J. 1997. Giftpflanzen. Ein Handbuch für Apotheker, Ärzte, Toxikologen und Biologen. Wissenschaftliche Verlagsgesellschaft mbH, ISBN 3-8047-1485-8
Heracleum mantegazzianum Somlier & Lavier	Apiaceae (Umbelliferae)	Whole plant	Furocoumarins (1.3%), e.g. bergapten, isopimpinellin, imperatorin		Pira E. et al. 1989. <i>Heracleum mantegazzianum</i> growth phases and furocoumarin content. <i>Contact Dermatitis</i> , 21, 300-303.
Heracleum sphondylium L.	Apiaceae (Umbelliferae)	Whole plant	Furocoumarins: e.g. bergapten, isopimpinellin, imperatorin		Bicchi C. et al. Chemical diversity of the constituents from the secondary structures of <i>Heracleum sphondylium</i> ssp. <i>sphondylium</i> . <i>Phytochemistry</i> 29(6), 1863-1867. Cooper M.R. and Johnson A.W. 1998. Poisonous plants and fungi in Britain. Animal and human poisoning. The Stationery Office, ISBN 0-11-242981-5.
Hernaria glabra L.	Caryophyllaceae	Whole plant			Rohkum H. 2008. Acute and sub-chronic toxicity of an aqueous extract of the leaves of <i>Hernaria glabra</i> in rodents. <i>J. Ethnopharmacol.</i> 116, 378-386, WITM 11, 2002.
Hibiscus sabdariffa L.	Maliaceae	Calyx	Oxalic acid (0.55%)	Compounds responsible for the adverse effects are not identified.	Teerogun and Piyokhontana. Ein Handbuch für die Praxis auf wissenschaftlicher Grundlage. Wissenschaftliche Verlagsgesellschaft mbH, ISBN 3-8047-1854-X.
Hibiscus sabdariffa L.	Maliaceae	Calyx	Oxalic acid (0.55%)	Compounds responsible for the adverse effects are not identified.	Pra E. et al. 1989. <i>Heracleum mantegazzianum</i> growth phases and furocoumarin content. <i>Contact Dermatitis</i> , 21, 300-303. Bicchi C. et al. Chemical diversity of the constituents from the secondary structures of <i>Heracleum sphondylium</i> ssp. <i>sphondylium</i> . <i>Phytochemistry</i> 29(6), 1863-1867. Cooper M.R. and Johnson A.W. 1998. Poisonous plants and fungi in Britain. Animal and human poisoning. The Stationery Office, ISBN 0-11-242981-5.

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<i>Hippomane manchella</i> L.	Euphorbiaceae	Aerial part	Phorbolsters from leaf and sap; Irindole alkaloid in fruit, possibly physostigmine.		Bandaranayake W. M. 2002. Bioactivities, bioactive compounds and chemical constituents of mangrove plants. Wetlands: Ecology and Management. 10: 421-452.
<i>Houttuynia andygosanthera</i> Wall. ex A.B.C.	Apocynaceae	Bark, root and seed	Steroid alkaloids: e.g. conessine, isocoumestranine, kudusessine,....		Burtonet J. 1999. Pharmacognosie, 3ème édition. Ed. Tec et Doc-Lavoisier. ISBN-2-7430-0315-4
<i>Hoodia gordonii</i> (Masson) Decne	Apocynaceae	Whole plant	Oxyprogesterone steroidal glycosides: e.g. hoodigosides and hoodistanosides A-B.	Listed as Novel Food in the NF Catalogue	Shukla V.J. et al. 2009. Pregnane glycosides from <i>Hoodia gordonii</i> . Phytochem. 70(5), 675-683. Janssen H.G. et al. 2008. Quantification of appetite suppressing steroid glycosides from <i>Hoodia gordonii</i> in dried plant material, purified extracts and food products using HPLC-UV and HPLC-MS methods. Anal. Chim. Acta. 617: 200-207. Pawar R.S. et al. 2007. New calogegenin glycosides from <i>Hoodia gordonii</i> . Steroids. 72, 881-891. Vermaek I. et al. 2011. <i>Hoodia gordonii</i> : an up-to-date review of a commercially important anti-obesity plant. Planta Medica. 77: 1149-1160
<i>Hosulinda opposita</i> Vahl	Lamiaceae (Labiatae)	Leaf	Essential oil from leaf; monoterpene etheroxide: 1,8-cineole (72%). Essential oil from fruit: bicyclic monoterpene: camphor (65%) Flavonone: 8-prenylaringenin		Chagnonda L.S. and Charachia J.C. 2005. The essential oil of wild and cultivated <i>Hosulinda opposita</i> Vahl. from Zimbabwe. Flavou. Fragr. J. 20(2). 193-195
<i>Humulus lupulus</i> L.	Cannabaceae	Inflorescence			Burtonet J. 1999. Pharmacognosie, 3ème édition. Ed. Tec et Doc-Lavoisier. ISBN-2-7430-0315-4
<i>Huperzia selago</i> (L.) Schrank & Mart. (<i>Lycopodium selago</i> L.)	Lycopodiaceae	Aerial part	Lycopodium alkaloids (lycodine class): e.g. huperzine A, huperzine B, N-methyl-huperzine B, huperzine		Boloz A. et al. 2008. The determination of huperzine A in European Lycopodiaceae species by HPLC-UV-MS. Phytochem. Analysis. 17(5): 332-336. Fidgenhauer N. et al. 2000. Identification with huperzine A, a potent anticholinesterase found in the <i>He</i> tribe moss. Clin. Toxicol. 38(7): 803-808.
<i>Huperzia serrata</i> (Thunb.) Trevis (<i>Lycopodium serratum</i> Thunb.)	Lycopodiaceae	Aerial part	Sesquiterpene alkaloids: e.g. huperzine A (approximately 0.007%) and huperzine B		Ma X. Gang D.R. 2004. The Lycopodium alkaloids. Nat. Prod. Rep. 21: 762-772.
<i>Hura crepitans</i> L.	Euphorbiaceae	Whole plant	Latex contains diterpenes (daphnane type): e.g. hippomane A (huraloxin) and hippomane B		Nelson L.S., Shim R.D., Balick M.J. 2007. Handbook of Poisonous and Injurious Plants. Second Edition. Stagger USA. ISBN-10: 0-597-31288-4.
<i>Hycihrilis orientalis</i> L.	Asparagaceae	Flower	Reported to contain the phenylpropanoid methylchavicol in unspecified quantities		EMEA-HMPC. 2005. Public statement on the use of herbal medicinal products containing estragole. EMEA/HMPC/137212/2005 Tisserand R. and Balazs T. 1995. Essential oil safety. A Guide for Health Care Professionals. Churchill Livingstone. ISBN: 0443052603.
<i>Hydnocarpus</i> spp.	Actinaceae	Seed	Genus in which species may contain in their seed oil unsaturated cycloperenyl acids mainly chauliognic acid, hydrocarnic acid, gonic acid.	The seed oil is referred to as non-edible. Oral intake of the seed oil may cause e.g. nausea, diarrhoea, hyperventilation	Krist S et al. 2008. Volatile compounds and thacylglycerol composition of original fatty plant oils. Eur. J. Lipid Sci. Technol. 110: 1271-140 Perrins GA. et al. 1937. Studies of chauliognic group oils. Industrial and Engineering Chemistry August: 939-942. Roth L., Dauberer M. and Kormanik K. 1994. Giftpflanzen - Pflanzenzöologie. Vorkommen WFRNun. Therapie. second. ISBN: 3-809-54810-4.
<i>Hydrastis canadensis</i> L.	Ranunculaceae	Whole plant	Isoquinoline alkaloids: e.g. hydrastine, berberine		Burtonet J. 1999. Pharmacognosy. Phytochemistry. Medicinal Plants. 2nd ed. Ed. Interedit Ltd. ISBN: 15829266-63-7
<i>Hyoscyamus</i> spp.	Solanaceae	Whole plant	Genus in which species may contain tropane alkaloids: e.g. atropine, hyoscyamine,....	The fresh plant contains hyoscyamine, twice more active than atropine (racemic mixture)	Burtonet J. 2005. Paras exiguus (Gleditsia) darigereux pour Thomme et les animaux. Ed. Tec & Doc. Lavoisier. Paris, 3ème édition. ISBN: 2-7430-0808-7
<i>Hypericum maculatum</i> Crantz	Hypericaceae	Aerial part	Dianthones and derivatives: e.g. hypericin (0.06-0.34%), pseudohypericin (0.25-1.45%)		Martoni P et al. 2006. Secondary metabolites variation in <i>Hypericum maculatum</i> and its relatives. Biochemical Systematics and Ecology 34: 565-59.
<i>Hypericum perforatum</i> L.	Hypericaceae	Aerial part	Dianthones and derivatives: e.g. hypericin, pseudo-hypericin; prenylated phenogluconol derivative: e.g. hypericon; xanthone derivatives.		Burtonet J. 2009. Pharmacognosie. (Phytochimie, Plantes médicinales). Ed. Tec & Doc. Lavoisier. Paris. 4ème édition. ISBN: 978-2-7430-1188-8
<i>Hyptis suaveolens</i> (L.) Poit.	Lamiaceae (Labiatae)	Aerial part	Essential oil: monoterpene etheroxide: 1,8-cineole (up to 44%). Also reported to contain methyl Eugenol in unspecified quantities	An aqueous crude extract of the leaves administered to rats for 28 days caused changes in histopathology and clinical chemistry suggestive of hepatotoxic and nephrotoxic effects.	EMEA-HMPC. 2004. Final position paper on the use of herbal medicinal products containing methyl Eugenol. EMEA/HMP/373/03 Cavalcanti ESB et al. 2004. Larvicidal Activity of Essential Oils from Brazilian Plants against <i>Aedes aegypti</i> L., Mem Inst Oswaldo Cruz. Rio de Janeiro, 99(5): 541-544. Odele G.M. and Abatan M.O. 2003. Histopathological and serum biochemical changes following oral administration of aqueous crude extracts of <i>Hyptis suaveolens</i> . Urena kobaia and <i>Cleome viscosa</i> in rats. Tropical Veterinary 22(1): 9-15.
<i>Hyssopus officinalis</i> L.	Lamiaceae (Labiatae)	Aerial part	Essential oil from aerial part: phenylpropanoids: e.g. methyl Eugenol (0.09-3.8%), methylchavicol (4.8%), monoterpenes: e.g. monoterpene etheroxide: 1,8-cineole; bicyclic monoterpene: e.g. pinocamphe (40%), isopino-camphe (30%), thujones (traces)		Council of Europe. 2007. Natural sources of Flavonoids. Report No. 2. Council of Europe Publishing. ISBN 978-92-871-6156-7.
<i>Isatis amara</i> L.	Brassicaceae (Cruciferae)	Whole plant	Curcubitrinins: e.g. curcubitrin E and I (0.2-0.4% in the seeds, 0.06% in flowers, 0.02% in the shoots and 0.006% in roots)		Tisserand R. and Balazs T. 1995. Essential oil safety. A Guide for Health Care Professionals. Churchill Livingstone. Edinburgh. ISBN: 0443052603
<i>Ilex aquifolium</i> L.	Aquifoliaceae	Aerial part	Cyanogenic glucoside: e.g. menisdaurin in ripe fruits		Reichling J. & Salek R. (2002) <i>Ilex aquifolium</i> (holly) Prof. ether. Helgoländer. Forsh. Komplementärmed. Arznei. 8: 21-33.
<i>Ilex paraguariensis</i> A. St.-Hil.	Aquifoliaceae	Leaf	Methylated xanthine derivatives: e.g. caffeine (0.2-2.0%), theobromine (1.0-1.2%), theophylline (0.05%)		Nairnsied A. and Wray V. 1990. Structural revision of a putative cyanogenic glucoside from <i>Ilex aquifolium</i> . Phytochemistry. 29(12). 3934-3936.
<i>Ilex vomitoria</i> Ait.	Aquifoliaceae	Fruit and leaf	Methylated xanthine derivatives: e.g. caffeine (0.3-0.9%), theobromine (0.03-0.31%)		Vaschocha B., Canguelara S. 2003. Fíolórula. Vademecum de Prescripcíon, Masson, 4. Ed. ISBN: 84-4581720-3, 978944581720-4
<i>Illicium anisatum</i> L. (<i>L. villosissimum</i> Seibold & Zucc.)	Schisandraceae	Bark and fruit	Essential oil: sesquiterpene lactones: e.g. anisatin (1205 mg/kg mean content in fruit), neoisatin, pseudoanisatin and phenylpropanoids: e.g. methyl Eugenol (9.8%)		Lung A.Y. and Foster S. 1996. Encyclopaedia of common natural ingredients used in food, drugs and cosmetics. John Wiley & Sons. Inc. ISBN: 0-471-50529-8.
<i>Illicium verum</i> Hook.	Schisandraceae	Fruit	Essential oil (0.5-6%), phenylpropanoids: e.g. transanethole (75-90%), methylchavicol (0.34-5.04%), safrole (0.14%)		Edwards M.L., Bennett B.C. 2005. Diversity of methylxanthine content in <i>Ilex</i> species L. and <i>Ilex vomitoria</i> Ait.: assessing sources of the North American stimulant Cassina. Economic Botany 59(3): 275-285. Johnans E.S. et al. 2002. An epidemic of epileptic seizures after consumption of herbal tea. Med. Tijdschr. Geneesk. 146(17): 813-816. Ledler I. 2006. Combination of TLC and HPLC-MS/MS methods. Approach to a rational quality control of Chinese Star Anise. J. Agric. Food Chem. 54: 1970-1974.
					Council of Europe. 2000. Natural sources of Flavonoids. Report No. 1. Council of Europe Publishing. ISBN 978-92-871-4324-2. Tisserand R. and Balazs T. 1995. Essential oil safety. A Guide for Health Care Professionals. Churchill Livingstone. Edinburgh. ISBN: 0443052603.

COMPENDIUM OF BOTANICALS REPORTED TO CONTAIN NATURALLY OCCURRING SUBSTANCES OF POSSIBLE CONCERN FOR HUMAN HEALTH WHEN USED IN FOOD AND FOOD SUPPLEMENTS

This compendium lists in alphabetical order botanicals without any judgment on whether these are suitable or not suitable for food applications in Europe. This compendium is part of a preliminary work undertaken by EFSA to harmonise the methodology across its panels for assessing the safety of botanicals and botanical preparations used in food and food supplements. Without prejudice to the existing legal framework, such compendium has no legal status and may not be used as support or evidence in any disagreement or dispute pertaining to the legal classification of products or substances. This compendium is a living document and is therefore open for additional contributions and comments.

Botanical name	Family	parts of plants of possible concern	Chemical of concern	Remarks on toxic/adverse effects) not known to be related to the identified chemical(s) of concern	Specific References
<i>Ipomoea</i> spp.	Convolvulaceae	Whole plant	Genus in which species may contain resiniferolins for the gastrointestinal system. Genus in which species may contain indolizidine alkaloids and serotonin-dihydroxyamino acid conjugates. Genus in which species may contain in the aerial parts pyridizidine alkaloids, e.g. jaspagulines (jaspaguline). Genus in which species may contain in the seeds alkaloids derived from lysergic acid.		
<i>Iris foetidissima</i> L.	Iridaceae	Leaf and rhizome		Toxic principles not defined but probably present in the irritant resin. Symptoms after ingestion: vomiting, diarrhoea (sometimes with bleeding).	Barveter J. 2009. Pharmacognosie, (Phytochimie, Plantes médicinales). Ed. Tec & Doc. Lavoisier, Paris, 4ème édition, 1269 pages, ISBN: 978-2-7430-1188-8.
<i>Iris pseudacorus</i> L.	Iridaceae	Leaf and rhizome		Toxic principles not defined but probably present in the irritant resin. Symptoms after ingestion: vomiting, diarrhoea (sometimes with bleeding).	Frome D., Pfander H.J. and Anton R. 2009. Plantes à risques. Ed. Tec et Doc Lavoisier, ISBN: 978-2-7430-0907-1
<i>Isatis tinctoria</i> L.	Brassicaceae	Leaf	Quinoline alkaloids, e.g. Iryplatinin		Ikeda, K. et al. 2003. Alkaloids from the poisonous plant <i>Ipomoea carnea</i> effects on intracellular lysosomal glycosidase activities in human lymphoblastoid cultures. J. Agric. Food Chem., 51, 7642-7646.
<i>Jatropha curcas</i> L.	Euphorbiaceae	Seed	Proteins, e.g. curcin		Schunhauer-Henrique, B. et al. 2003. The Clinical, Biochemical, Haematological and Pathological Effects of Long-term Administration of <i>Ipomoea carnea</i> to Growing Goats, Vet. Res. Commun., 27: 311-319
<i>Juglans regia</i> L.	Juglandaceae	Fruit, husk and leaf	Essential oil from leaf: bicyclic monoterpenes: e.g. beta-thujone (0.29%) Core: methyl ester derivatives of oxygenated diterpenic acids	Juglone is found in 29.8% of the surface waxes of the fruit (pericarp) and 28.6% of the surface waxes of the leaves.	Janet-Stens et al., 1998. Pyridizidine alkaloids of <i>Ipomoea hederifolia</i> and related species. Phytochemistry, 47(8): 1551-1560. Göhner, A. et al. 2002. Accumulation of heavy metals in water spinach (<i>Ipomoea aquatica</i>) cultivated in the Bangkok region, Thailand. Environmental Toxicology and Chemistry, 21: 1934–1939
<i>Juniperus communis</i> L.	Cupressaceae	Cone and leaf	Essential oil from leaf: bicyclic monoterpenes: e.g. beta-thujone (0.29%)	<i>J. communis</i> var. <i>communis</i> leaf essential oil contained from 0.0-4% α -thujone and from 0.0-4% β -thujone.	Cooper M.R. and Johnson A.W. 1998. Poisonous plants and fungi in Britain. Animal and Human Poisoning. The Stationery Office, ISBN 0-11-224281-5. Cooper M.R. and Johnson A.W. 1998. Poisonous plants and fungi in Britain. Animal and Human Poisoning. The Stationery Office, ISBN 0-11-224281-5. Michael, J.P. 2005. Quinine, quinine, quinine and acetone alkaloids. Nat. Prod. Rep. 22, 627-646
<i>Juniperus oxycedrus</i> L.	Cupressaceae	Branch and wood	Wood oil (Cade oil): phenolic compounds: e.g. cresol, para-cresol in the non-volatile oil fraction	Unidentified oil is carcinogenic. Reported case of poisoning with fever, severe hypotension, renal failure and hepatotoxicity.	Stum S. and Suppiger H. 1998. Analysis of isquinoline alkaloids in medicinal plants by capillary electrophoresis – mass spectrometry. Electrophoresis, 19, 3026-3032.
<i>Juniperus phoenicea</i> L.	Cupressaceae	Cone and leaf	Leaf: lignans, e.g. deoxypodophylotoxin, beta-peltatin A Core: methyl ester derivatives of oxygenated diterpenic acids	Contraindications with severe renal disease. Based on the results of preclinical investigations it can be concluded that <i>Juniperus communis</i> extract has anti-fertility and abortifacient effects in rats	Abou-Arguive I. et al. 1986. Acute toxicity studies with <i>J. curcas</i> L. Human. Toxicol. 5, 269-274.
<i>Juniperus sibirica</i> L.	Cupressaceae	Whole plant	Essential oil: bicyclic monoterpenes: e.g. sabinyl acetate(20-53%), sabinene (20-42%)	Pregnant cows fed from gestational day 250 with leaves (4.5-5.5 kg leaves/day equivalent to 190-245 mg isocaproic acid) aborted 3-4 days after feeding.	Achenbach H. and Benirschke G. 1997. Joanesalactone and other compounds from <i>J. lycocarpus</i> . Phytochemistry, 45(1), 149-157.
<i>Juniperus thurifera</i> L.	Cupressaceae	Cone and leaf	Leaf: lignans, e.g. deoxypodophylotoxin, beta-peltatin A Core: methyl ester derivatives of oxygenated diterpenic acids	Unidentified oil is carcinogenic. Reported case of poisoning with fever, severe hypotension, renal failure and hepatotoxicity.	Colacic M. et al. 2005. Phenolic acids, syringadienylglycosides, and juglone in fruits of different cultivars of <i>Juglans regia</i> L. J. Agric. Food Chem. 53, 6390-6396.
<i>Juniperus virginiana</i> L.	Cupressaceae	Cone and leaf	Leaf: lignans, e.g. deoxypodophylotoxin, beta-peltatin A Essential oil from the leaf: phenylpropanoids, e.g. methylchavicol, methylchavicol		Frome D. and Pfander H.J. 1997. Giftparzen. Ein Handbuch für Apotheker, Ärzte, Toxikologen und Biologen. Wissenschaftliche Verlagsgesellschaft mbH. ISBN: 3-8047-1466-8.
<i>Kalanchoe pinnata</i> Lam.) Pers. (Byophyllum calycinum Salib.)	Crassulaceae	Leaf	Bulbiferolides: e.g. thyophyllin A and C		Prasad R.B.N. and Guiz P. G. 1990. Surface waxes from leaves and fruits of walnut. Phytochemistry 29(7), 2097-2099. Paulsen M.T. and Ljunggren M. 2005. The natural toxin juglone causes degradation of p53 and induces rapid H2AX phosphorylation and cell death in human fibroblasts. Toxicol. Appl. Pharmacol. 209(1), 1-9.
<i>Kalanchoe pinnata</i> Lam.) Pers. (Byophyllum calycinum Salib.)	Crassulaceae	Leaf	Bulbiferolides: e.g. thyophyllin A and C		Angiot A. et al. 2003. Chemical composition of the essential oils of <i>Juniperus</i> from ripe and unripe berries and leaves and their antimicrobial activity. J. Agric. Food Chem. 51, 3073-3078.
<i>Kalanchoe pinnata</i> Lam.) Pers. (Byophyllum calycinum Salib.)	Crassulaceae	Leaf	Bulbiferolides: e.g. thyophyllin A and C		Bukhtere R. et al. 2009. Two chemotypes of essential oils produced by the same <i>Juniperus communis</i> L. growing wild in Lithuania. Chemia, 20(3), 195-201.
<i>Kalanchoe pinnata</i> Lam.) Pers. (Byophyllum calycinum Salib.)	Crassulaceae	Leaf	Bulbiferolides: e.g. thyophyllin A and C		Gaidner E. (1998) Abortifacient effect of doxepine pine (<i>Pinus contorta</i>) and common juniper (<i>Juniperus communis</i>) on cattle. Vet. Hum. Toxicol. 40(5), 280-283.
<i>Kalanchoe pinnata</i> Lam.) Pers. (Byophyllum calycinum Salib.)	Crassulaceae	Leaf	Bulbiferolides: e.g. thyophyllin A and C		EMA HMPC/2010. Assessment report on <i>Juniperus communis</i> L., aetherioleum. EMA/HMPC/12410/2010.
<i>Kalanchoe pinnata</i> Lam.) Pers. (Byophyllum calycinum Salib.)	Crassulaceae	Leaf	Bulbiferolides: e.g. thyophyllin A and C		Koruk U. et al. 2005. Juniper leaf poisoning. Clin. Toxicol. 1, 47-48.
<i>Kalanchoe pinnata</i> Lam.) Pers. (Byophyllum calycinum Salib.)	Crassulaceae	Leaf	Bulbiferolides: e.g. thyophyllin A and C		Tisserand, R. and Balaska, T., 1995. Essential oil safety. A Guide for Health Care Professionals. Churchill Livingstone, Edinburgh. ISBN: 0443052803.
<i>Kalanchoe pinnata</i> Lam.) Pers. (Byophyllum calycinum Salib.)	Crassulaceae	Leaf	Bulbiferolides: e.g. thyophyllin A and C		Barneo, A.F. et al. 2004. Oxygenated diterpenes and other constituents from Moroccan <i>Juniperus phoenicea</i> and <i>Juniperus thurifera</i> var. <i>athabasca</i> . Phytochemistry, 65(17), 2507-2515.
<i>Kalanchoe pinnata</i> Lam.) Pers. (Byophyllum calycinum Salib.)	Crassulaceae	Leaf	Bulbiferolides: e.g. thyophyllin A and C		James D.A. et al. 1980. Plant anticancer agents. X. Lignans from <i>Juniperus phoenicea</i> . J. Nat. Prod. 43, 495-497.
<i>Kalanchoe pinnata</i> Lam.) Pers. (Byophyllum calycinum Salib.)	Crassulaceae	Leaf	Bulbiferolides: e.g. thyophyllin A and C		Sawi S.A.E. and Molave H.M. 2008. Labdane, pimarane and abietane diterpenes from the fruits of <i>Juniperus phoenicea</i> L. grown in Egypt and their activities against human liver carcinoma. Canadian Journal of Pure and Applied Sciences 2(1), 115-122.
<i>Kalanchoe pinnata</i> Lam.) Pers. (Byophyllum calycinum Salib.)	Crassulaceae	Leaf	Bulbiferolides: e.g. thyophyllin A and C		Tisserand, R. and Balaska, T., 1995. Essential oil safety. A Guide for Health Care Professionals. Churchill Livingstone, Edinburgh. ISBN: 0443052803.
<i>Kalanchoe pinnata</i> Lam.) Pers. (Byophyllum calycinum Salib.)	Crassulaceae	Leaf	Bulbiferolides: e.g. thyophyllin A and C		Barneo, A.F. et al. 2004. Oxygenated diterpenes and other constituents from Moroccan <i>Juniperus phoenicea</i> and <i>Juniperus thurifera</i> var. <i>athabasca</i> . Phytochemistry, 65(17), 2507-2515.
<i>Kalanchoe pinnata</i> Lam.) Pers. (Byophyllum calycinum Salib.)	Crassulaceae	Leaf	Bulbiferolides: e.g. thyophyllin A and C		James D.A. et al. 1980. Plant anticancer agents. X. Lignans from <i>Juniperus phoenicea</i> . J. Nat. Prod. 43, 495-497.
<i>Kalanchoe pinnata</i> Lam.) Pers. (Byophyllum calycinum Salib.)	Crassulaceae	Leaf	Bulbiferolides: e.g. thyophyllin A and C		Sawi S.A.E. and Molave H.M. 2008. Labdane, pimarane and abietane diterpenes from the fruits of <i>Juniperus phoenicea</i> L. grown in Egypt and their activities against human liver carcinoma. Canadian Journal of Pure and Applied Sciences 2(1), 115-122.
<i>Kalanchoe pinnata</i> Lam.) Pers. (Byophyllum calycinum Salib.)	Crassulaceae	Leaf	Bulbiferolides: e.g. thyophyllin A and C		Tisserand, R. and Balaska, T., 1995. Essential oil safety. A Guide for Health Care Professionals. Churchill Livingstone, Edinburgh. ISBN: 0443052803.
<i>Kalanchoe pinnata</i> Lam.) Pers. (Byophyllum calycinum Salib.)	Crassulaceae	Leaf	Bulbiferolides: e.g. thyophyllin A and C		Barneo, A.F. et al. 2004. Oxygenated diterpenes and other constituents from Moroccan <i>Juniperus phoenicea</i> and <i>Juniperus thurifera</i> var. <i>athabasca</i> . Phytochemistry, 65(17), 2507-2515.
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<i>Kalanchoe pinnata</i> Lam.) Pers. (Byophyllum calycinum Salib.)	Crassulaceae	Leaf	Bulbiferolides: e.g. thyophyllin A and C		James D.A. et al. 1980. Plant anticancer agents. X. Lignans from <i>Juniperus phoenicea</i> . J. Nat. Prod. 43, 495-497.
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<i>Kalanchoe pinnata</i> Lam.) Pers. (Byophyllum calycinum Salib.)	Crassulaceae	Leaf	Bulbiferolides: e.g. thyophyllin A and C		Tisserand, R. and Balaska, T., 1995. Essential oil safety. A Guide for Health Care Professionals. Churchill Livingstone, Edinburgh. ISBN: 0443052803.
<i>Kalanchoe pinnata</i> Lam.) Pers. (Byophyllum calycinum Salib.)	Crassulaceae	Leaf	Bulbiferolides: e.g. thyophyllin A and C		Barneo, A.F. et al. 2004. Oxygenated diterpenes and other constituents from Moroccan <i>Juniperus phoenicea</i> and <i>Juniperus thurifera</i> var. <i>athabasca</i> . Phytochemistry, 65(17), 2507-2515.
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<i>Kalanchoe pinnata</i> Lam.) Pers. (Byophyllum calycinum Salib.)	Crassulaceae	Leaf	Bulbiferolides: e.g. thyophyllin A and C		Sawi S.A.E. and Molave H.M. 2008. Labdane, pimarane and abietane diterpenes from the fruits of <i>Juniperus phoenicea</i> L. grown in Egypt and their activities against human liver carcinoma. Canadian Journal of Pure and Applied Sciences 2(1), 115-122.
<i>Kalanchoe pinnata</i> Lam.) Pers. (Byophyllum calycinum Salib.)	Crassulaceae	Leaf	Bulbiferolides: e.g. thyophyllin A and C		Tisserand, R. and Balaska, T., 1995. Essential oil safety. A Guide for Health Care Professionals. Churchill Livingstone, Edinburgh. ISBN: 0443052803.
<i>Kalanchoe pinnata</i> Lam.) Pers. (Byophyllum calycinum Salib.)	Crassulaceae	Leaf	Bulbiferolides: e.g. thyophyllin A and C		Barneo, A.F. et al. 2004. Oxygenated diterpenes and other constituents from Moroccan <i>Juniperus phoenicea</i> and <i>Juniperus thurifera</i> var. <i>athabasca</i> . Phytochemistry, 65(17), 2507-2515.
<i>Kalanchoe pinnata</i> Lam.) Pers. (Byophyllum calycinum Salib.)	Crassulaceae	Leaf	Bulbiferolides: e.g. thyophyllin A and C		James D.A. et al. 1980. Plant anticancer agents. X. Lignans from <i>Juniperus phoenicea</i> . J. Nat. Prod. 43, 495-497.
<i>Kalanchoe pinnata</i> Lam.) Pers. (Byophyllum calycinum Salib.)	Crassulaceae	Leaf	Bulbiferolides: e.g. thyophyllin A and C		Sawi S.A.E. and Molave H.M. 2008. Labdane, pimarane and abietane diterpenes from the fruits of <i>Juniperus phoenicea</i> L. grown in Egypt and their activities against human liver carcinoma. Canadian Journal of Pure and Applied Sciences 2(1), 115-122.
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Botanical name	Family	parts of plants of possible concern	Chemical of concern	Remarks on toxic/adverse effects(s) not known to be related to the identified chemical(s) of concern	Specific References
<i>Kalmia latifolia</i> L.	Ericaceae	Leaf	Hydroquinone; e.g. arbutin and diisoprenes; e.g. andromedotoxin		Verlangien A.J. et al. 1976. Acute toxicity of <i>Kalmia angustifolia</i> , (sheep burea) extracts in the rat. Vet. Toxicol. 18, 122-124. Frohe D. and Prander H.J. 1997. Giftpflanzen. Ein Handbuch für Apotheker, Ärzte, Toxikologen und Biologen. Wissenschaftliche Verlagsgesellschaft mbH. ISBN 3-8047-1468-8. WICRI M (2002). Teedrogen und Phytopharmaka. Wissenschaftliche Verlagsgesellschaft mbH. Hagers Handbuch der Pharmazeutischen Praxis 1998. Springer Verlag. ISBN 3-540-52688-9. Hagers Handbuch der Pharmazeutischen Praxis 1998. Springer Verlag. ISBN 3-540-52688-9.
<i>Krameria lapacea</i> (Domby) Burdet & B. Simpson (Kameria <i>haradia</i> Ruiz & Pav.)	Kameriaceae	Root		Root: 8-18% tannin-proanthocyanidins, root bark: 18-42% tannin-proanthocyanidins.	
<i>Laburnum anagyroides</i> Medik. (<i>Laburnum vulgare</i> J. Presl., <i>Cytisus abrotanum</i> L.)	Leguminosae (Fabaceae)	Whole plant	Quinoline alkaloids; e.g. cytisine		Bestari S. et al. 2009. Wild lettuce (<i>Lactuca virosa</i>) toxicity. Brit. Med. J. Case Reports. doi:10.1136/bcr.2008.0134. Frohe D. and Prander H.J. 1997. Giftpflanzen. Ein Handbuch für Apotheker, Ärzte, Toxikologen und Biologen. Wissenschaftliche Verlagsgesellschaft mbH. ISBN 3-8047-1468-8. Kudoczka K.H. and Schutze W. 1987. Biology and chemistry of conifer oils. Flavour Fragr. J. 2, 137-148. Valentine J.L. et al. 1994. Gas chromatographic determination of nordihydroguajaretic acid in <i>Lactuca divaricata</i> . Anal. Lett. 17, 1617-1628. Arreaga S. et al. 2005. <i>Larrea tridentata</i> (Creosote bush), an abundant plant of Mexican and US-American deserts and its metabolite nordihydroguajaretic acid. J. Ethnopharmacol. 98, 231-239. Sheikh N.M. et al. 1996. Chaparral-associated hepatotoxicity. Arch. Intern. Med. 157, 913-919. Cheske P.R. (Ed.). 1989. Toxicants of plant origin. Volume III. Proteins and amino acids. CRC Press, Inc., ISBN: 0-8493-6692-4. Ludolph A.C. and Spenser P.S. 1996. Toxic models of upper motor neuron disease. Journal of the Neurological Sciences, 139 (Suppl), S3-S9. Spencer P.S. et al. 1988. Lathyrism: evidence for role of the neuroexcitatory amino acid BONA. Lancet. November 8, 1068-1067. Yan Z.Y. et al. 2006. <i>Lathyrus sativus</i> (grass pea) and its neurotoxin ODA-P. Phytochemistry, 67, 1071-121.
<i>Larrea divaricata</i> Cav.	Zygophyllaceae	Aerial part	Essential oil from bark; monoterpene etheroxide: 1,8-cineole (2.09%) Essential oil from leaf		
<i>Larrea tridentata</i> (Ses. & Moc. ex DC.) Cav.	Zygophyllaceae	Aerial part	Lignans: e.g. nordihydroguajaretic acid		
<i>Lathyrus sativus</i> L.	Leguminosae (Fabaceae)	Seed	Amino acids (0.02-2.5% in dry seeds); e.g. β -N-oxalyl-DL-diaminopropionic acid (B-ODAP), β -N-oxalylamino-L-alanine (BOAA)		
<i>Laurus nobilis</i> L.	Lauraceae	Fruit and leaf	Essential oil from leaf; phenylpropanoids: e.g. methyl Eugenol (17-11.8%) and monoterpene etheroxide: 1,8-cineole (34-53%)		Council of Europe. 2008. Natural sources of Flavonoids. Report No. 3. Council of Europe Publishing. ISBN: 978-92-871-4422-3. García-Valejo M.C. et al. 1989. Essential oils of genus <i>Lavandula</i> in Spain. Proc. Int. Congr. Essential Oil Fragrances. Havours, 4, 15-26. Närf R. and Mörs A.F. 1992. [Lavender-avandula (a comparison)]. Rivista Italiana Etopos (special edition), 364-377. Peter K.V. 2004. Handbook of Herbs and Spices vol 2. Woodhead Publishing Limited, ISBN: 1-85573-721-3.
<i>Lavandula latifolia</i> Medik. (<i>Lavandula spica</i> auct., non L.)	Lamiaceae (Labiatae)	Aerial part	Essential oil from aerial part; monoterpene etheroxide: 1,8-cineole (33%), bicyclic monoterpenes: e.g. camphor (5%) Essential oil from flower; monoterpene etheroxide: 1,8-cineole (23-48%), bicyclic monoterpenes: e.g. camphor (11-18%) Essential oil from leaf; monoterpene etheroxide: 1,8-cineole (47-55%), bicyclic monoterpenes: e.g. camphor (32-44%)		Munoz-Betorreu J. et al. 2007. Essential oil variation within and among natural populations of <i>Lavandula latifolia</i> and its relation to their ecological areas. Biochem. Syst. Ecol. 35, 479-488.
<i>Lavandula stoechas</i> L.	Lamiaceae (Labiatae)	Aerial part	Essential oil from leaf; bicyclic monoterpenes: e.g. thujones, camphor (0.99%) and monoterpene etheroxide: 1,8-cineole (3.32-30%) Essential oil from fresh flower; bicyclic monoterpenes: e.g. camphor (13.32%) and monoterpene etheroxide: 1,8-cineole (5.81%)		Argüot A. et al. 2006. Chemical composition, seasonal variability, and antifungal activity of <i>Lavandula stoechas</i> L. ssp. <i>stoechas</i> essential oils from stemleaves and flowers. J. Agric. Food Chem. 54, 4364-4370. Kırmızıbekmez H. et al. 2009. Chemical composition and antimicrobial activity of the essential oils of <i>Lavandula stoechas</i> L. ssp. <i>stoechas</i> growing wild in Turkey. NatProd Commun. 4(7):1001-6 Tzakou O. et al. 2009. Essential oil composition and enantiomeric distribution of thujone and camphor of <i>Lavandula canariensis</i> and <i>L. stoechas</i> ssp. <i>stoechas</i> grown in Greece. Natl Prod. Commun. 2009, 4(9), 1103-1106. SCONFP (2002). Opinion on the Scientific Committee on Cosmetic Products and Non-Food Products Intended for Consumers concerning <i>Lavandula vera/mix</i> , herein (adopted by the SCONFP during the 21st Plenary meeting of 17 September 2002. Frohe D. and Prander H.J. 1997. Giftpflanzen. Ein Handbuch für Apotheker, Ärzte, Toxikologen und Biologen. Wissenschaftliche Verlagsgesellschaft mbH. ISBN: 3-8047-1468-8.
<i>Levensonla nemris</i> L. (<i>Lavosonia alba</i> Lam.)	Lythraceae	Leaf	Naphthoquinone; e.g. lawsone (dried leaf content 1-2%)		European Medicines Agency. 2010. Assessment report on <i>Leonurus cardica</i> L., herba. Draft. Newall C.A., Anderson L.A., Phillipson J.D. 1996. Herbal medicines: a guide for health care professionals. The Pharmaceutical Press. ISBN: 0853232590.
<i>Leonurus palustris</i> L.	Erdaceae	Whole plant	Diisoprenes: e.g. acetylcholinesterase inhibitor		Chen Z. et al. 2004. Determination of staerydine and leonurine in Herba Leonuri by on-par reversed-phase high-performance liquid chromatography. Di Yi Yun Yi Da Xue Xue Bao. 24 (11), 1223-1226. Chen Z. et al. 2010. Development and validation of an UPLC-DAD-MS method for the determination of leonurine in Chinese medicine Herba Leonuri (Leonurus japonicus). J. Chromatogr. Sci. 48 (10), 802-6.
<i>Leonurus japonicus</i> Houtt. (<i>Leonurus heterophyllus</i> Svedel)	Lamiaceae (Labiatae)	Aerial part	Pyridoline alkaloids: e.g. staerydine (0.1-0.2%) and and cyclic peptide: cycloleucunurine		Hong S.S. 2001. Isolation and quantitative analysis of leonurine from Leonuri herba]. Sengnyak Hakkochei. 32(4), 322-326.
<i>Leonurus sibiricus</i> L.	Lamiaceae (Labiatae)	Aerial part	Pyridoline alkaloids: e.g. staerydine and and cyclic peptide: cycloleucunurine Diisoprenes: e.g. lescibiricin		

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<i>Lepidium meyenii</i> Walp. (<i>Lepidium peruvianum</i> Chacon)	Brassicaceae	Root	Indazole alkaloids (0.0016-0.0123% in the dried root); e.g. lepidiline A, B and C		Cicero A.F.G. et al. 2001. <i>Lepidium meyenii</i> Walp. improves sexual behaviour in male rats independently from its action on spontaneous locomotor activity. <i>J. Ethnopharmacol.</i> 75, 225-229. Jin W. et al. 2007. Identification of <i>Lepidium meyenii</i> (Walp.) based on spectra and chromatographic characteristics of its principal functional ingredients. <i>J. Sci. Food. Agric.</i> 87, 2251-2258. McColellan M.M. et al. 2005. Analysis of macemides in samples of maca (<i>Lepidium meyenii</i>) by HPLC-DV-MS/MS. <i>Phytochem. Anal.</i> 16, 463-469. Oshima et al. 2003. Identification of <i>Lepidium meyenii</i> (Walp.) based on spectra and chromatographic characteristics of its principal functional ingredients. <i>J. Vet. Med. Sci.</i> 65(10), 1145-1146. Zheng et al. 2000. Effect of a lipidic extract from <i>Lepidium meyenii</i> on sexual behavior in mice and rats. <i>Urology</i> , 55, 599-602.
<i>Leucanthemum vulgare</i> Lam. (<i>Chrysanthemum leucanthemum</i> L.)	Compositae (Asteraceae)	Flower	Unsaturated pyridizidine alkaloids: e.g. platyphylline, senebionine		Sagarshini V.T.G. 2000. Alkaloids of <i>Leucanthemum vulgare</i> . <i>Chem. Nat. Compd.</i> 36(3), 327.
<i>Leucosium vernum</i> L.	Amaryllidaceae	Bulb and leaf	Isoquinoline alkaloids (Amaryllidaceae alkaloids) e.g. lycorine, homolycorine, 2-O-acetyllycorine		Roth L., Daurer M. and Kormanik K. 1994. Giftpflanzen – Pflanzenstoffe. Nikol Verlagsgesellschaft mbH & Co. KG, Hamburg. ISBN 3-933202-31-7. Schwark L. 2004. Alkaloids from <i>Leucosium vernum</i> and antitumor activity of amaryllidaceae alkaloids. <i>Pharmazie</i> 70, 871-873.
<i>Levisticum officinale</i> W.A.D.Koch	Apiaceae (Umbelliferae)	Whole plant	Furocoumarins in root and seeds: e.g. lipoatoenin (12.82 mg/kg), isosoralen (3.8 mg/kg), 8-methoxypsoralen (0.5 mg/kg), furcoumarin in leaf: e.g. 5-methoxypsoralen. In stem: monoterpenes: bicyclic monoterpenes: e.g. alpha and beta thuyones, and monoterpene etheroxide: 1,8-cineole		Lin L.S.S.A. 2007. Gel based bioassay for the isolation and characterization of novel phytoprogens from <i>Ligusticum chinuixiong</i> and the detection of xerostrogens androgens from Singapore's marine environment. PhD. thesis, National University of Singapore. Lin L.S. et al. 2006. Dynamics of progestogenic activity in serum following administration of <i>Ligusticum chinuixiong</i> . <i>Life Sci.</i> 79, 1724-1780. Zhang C et al. 2007. Analysis of the volatile compounds in <i>Ligusticum chinuixiong</i> Hort. <i>Jena HSSPME-GC/MS</i> . <i>J. Pharmazie/Biomed.</i> 44, 464-470. Roth L., Daurer M. and Kormanik K. 1994. Giftpflanzen – Pflanzenstoffe. Nikol Verlagsgesellschaft mbH & Co. KG, Hamburg. ISBN 3-933202-31-7.
<i>Ligustrum vulgare</i> L.	Oleaceae	Bark, fruit and leaf	Secoiridoid glucosides (8.85% in ripe fruits)		Mimaki Y. and Sashida Y. 1990. Steroidal saponins and alkaloids from the barks of <i>Lilium brownii</i> var. <i>colchense</i> . <i>Chem. Pharm. Bull.</i> 38, 3095-3099. Wang H. et al. 2002. Isolation of lirin, a novel arginine- and glutamate-rich protein with potent antitumor and mitogenic activities from lily bulbs. <i>Life. Sci.</i> 70, 1075-1094.
<i>Lilium brownii</i> F.E.Br. ex Meisner	Liliaceae	Bulb	Bulb reported to contain steroidal saponins and steroidal alkaloids and a protein: e.g. lirin,		Arai C.K. 1982. Chemistry and pharmacology of vasidone - a new oxynone and abortifacient. <i>J. Ethnopharmacol.</i> 6(1), 125-126. Hua H. et al. 2002. A new polyquinazolinone alkaloid from <i>Linaria vulgaris</i> . <i>Chem. Pharm. Bull.</i> 50(10), 1393-1394.
<i>Linaria vulgaris</i> Mill.	Plantaginaceae	Aerial part	Quinoline alkaloids: e.g. vasicine		Council of Europe. 2008. Natural sources of Flavonings. Report No. 3. Council of Europe Publishing. ISBN 978-92-871-6422-3. Haque M.R. and Bradbury J.H. 2002. Total cyanide determination of plants and foods using the picrate and acid hydrolysis methods. <i>Food Chem.</i> 77, 107-114. Niedzwiedz-Sieglen I. 1998. Cyanogenic glucosides in <i>Linum usitatissimum</i> . <i>Phytochemistry</i> , 49, 59-63. Orman B.D. et al. 1992. Cyanogenic compounds in linsseed. <i>J. Agric. Food Chem.</i> 40, 1346-1348. Schlichter H. von and Wilkens-Sailer M. 1996. Quantitative Bestimmung cyanogener Glykoside in <i>Linum usitatissimum</i> mit Hilfe der HPLC. <i>Fel. Wiss. Technol.</i> 88, 287-290.
<i>Linum usitatissimum</i> L.	Linaceae	Seed	Cyanogenic glycosides: e.g. diglucosides linostatin and neolinostatin (2.6 resp. 3.5 mg/kg) and traces of linamarin monoglucoside. Lignan: prosteroiddiglucosid		EMEA-HMP-PC. 2004. Final position paper on the use of herbal medicinal products containing methyleugenol. EMEA/HMP/WP/357/03 Juliani H.K. et al. 2002. Intra-specific variation in leaf oils of <i>Lippia junelliana</i> (Mold.) Tronc. <i>Biochim. Syst. Ecol.</i> 30, 163-170. EMEA-HMP-PC. 2004. Final position paper on the use of herbal medicinal products containing methyleugenol. EMEA/HMP/WP/357/03 Dusaratay C. et al. [Composition of essential oils from <i>Lippia junelliana</i> , <i>Lippia integrifolia</i> and <i>Lippia tubulata</i> from San Luis Province (Argentina)]. <i>Revista Colombiana de Quimica</i> 27(2), 9-16. EMEA-HMP-PC. 2004. Final position paper on the use of herbal medicinal products containing methyleugenol. EMEA/HMP/WP/357/03
<i>Lippia integrifolia</i> Hieron.	Verbenaceae	Unspecified	Essential oil reported to contain methyleugenol in unspecified quantities		Fernandez X. et al. 2005. Chemical composition of the essential oils from Turkish and Honduran styrax. <i>Flavour Fragr. J.</i> 20, 70-73. International Agency for Research on Cancer. 2002. Styrene. IARC Summary & Evaluation 82.
<i>Lippia junelliana</i> (Moldenke) Tronc.	Verbenaceae	Leaf	Essential oil from leaf: phenylpropanoid: methyleugenol (0.1-2.9%)	Methyleugenol content in leaf essential oil varied from 0.1-2.9% in samples from 16 regions in Argentina	Fernandez X. et al. 2005. Chemical composition of the essential oils from Turkish and Honduran styrax. <i>Flavour Fragr. J.</i> 20, 70-73. International Agency for Research on Cancer. 2002. Styrene. IARC Summary & Evaluation 82.
<i>Lippia laxiflora</i> Hieron.	Verbenaceae	Unspecified	Essential oil reported to contain methyleugenol in unspecified quantities		Fernandez X. et al. 2005. Chemical composition of the essential oils from Turkish and Honduran styrax. <i>Flavour Fragr. J.</i> 20, 70-73. International Agency for Research on Cancer. 2002. Styrene. IARC Summary & Evaluation 82.
<i>Lippia turbinata</i> Griseb.	Verbenaceae	Aerial parts	Essential oil from aerial part: monoterpene etheroxide: 1,8-cineole (14.7%) Essential oil reported to contain methyleugenol in unspecified quantities		Fernandez X. et al. 2005. Chemical composition of the essential oils from Turkish and Honduran styrax. <i>Flavour Fragr. J.</i> 20, 70-73. International Agency for Research on Cancer. 2002. Styrene. IARC Summary & Evaluation 82.
<i>Liquidambar orientalis</i> Mill.	Astringaceae	Bark	Resin: 0.55% styrene. Essential oil from balsam: 70% styrene (vinylbenzene)	International Agency for Research on Cancer has evaluated that styrene is possible carcinogenic to humans (Group 2 B)	Fernandez X. et al. 2005. Chemical composition of the essential oils from Turkish and Honduran styrax. <i>Flavour Fragr. J.</i> 20, 70-73. International Agency for Research on Cancer. 2002. Styrene. IARC Summary & Evaluation 82.
<i>Liquidambar styraciflua</i> L.	Astringaceae	Bark	Essential oil from balsam: 31% styrene	International Agency for Research on Cancer has evaluated that styrene is possible carcinogenic to humans (Group 2 B)	Fernandez X. et al. 2005. Chemical composition of the essential oils from Turkish and Honduran styrax. <i>Flavour Fragr. J.</i> 20, 70-73. International Agency for Research on Cancer. 2002. Styrene. IARC Summary & Evaluation 82.

COMPENDIUM OF BOTANICALS REPORTED TO CONTAIN NATURALLY OCCURRING SUBSTANCES OF POSSIBLE CONCERN FOR HUMAN HEALTH WHEN USED IN FOOD AND FOOD SUPPLEMENTS

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<i>Lithospermum</i> spp.	Boraginaceae	Whole plant	Genus in which species may contain unsaturated pyridizidine alkaloids e.g. lithosamine, lincramidine, lycopsamine		Mrozczak T. et al. 2004. Screening for pyridizidine alkaloids in plant materials by electron ionization RP-HPLC-MS with thermobeam interface. Biomed. Chromatogr. 18, 745–751. Kern L. et al. 1994. Pyridizidine alkaloids from <i>Lithospermum officinale</i> . Phytochemistry, 37(1), 275-277.
<i>Litsea cubeba</i> (Lour) Pers.	Lauraceae	Bark and stem	Isoquinoline alkaloids (phenanthrene type); e.g. litetamine		Huang C.H. et al. 2008. Litetamine, a phenanthrene alkaloid from the wood of <i>Litsea cubeba</i> , inhibits rat smooth muscle cell adhesion and migration on collagen. European J. Pharmacol. 596: 28-31.
<i>L. cubeba</i> spp.	Campanulaceae	Whole plant	Genus in which species may contain piperidine alkaloids e.g. labeine		Frome D., Pfänder H.J. and Anton R. 2009. Plantes à risques. Ed. Tec et Doc-Lavoisier. ISBN: 978-2-7430-0907-1
<i>Lolium temulentum</i> L.	Poaceae	Seed		Reported toxicity (on livestock) but the nature of the involved substances has not been established.	Frome D. and Pfänder H.J. 1997. Giftpflanzen. Ein Handbuch für Apotheker, Ärzte, Toxikologen und Biologen. Wissenschaftliche Verlagsgesellschaft mbH. ISBN 3-8047-1468-8. Cooper M.R. and Johnson A.W. 1998. Poisonous plants and fungi in Britain. Animal and human poisoning. The Stationery Office. ISBN 0-11-242981-5. Schiard C.L. et al. 2007. Loline alkaloids. Cures of multitalism. Phytochemistry, 68(7), 980-996
<i>Lonicera</i> spp.	Caprifoliaceae	Fruit	Genus in which species may contain terpenoid saponins and traces of pyridinium alkaloid-coupled scordifidols	Fruits from <i>L. loncera</i> species tested in mice. Immature fruits more toxic than mature ones and pericarpis more toxic than seeds when tested in mice. Vomiting, diarrhoea and lethargy have been reported in dogs.	Burton J. 2009. Pharmacognosie, (Phytochimie, Plantes médicinales), Ed. Tec & Doc-Lavoisier. Paris. 4ème édition. 1269 pages. ISBN: 978-2-7430-1188-8 Cooper M.R. and Johnson A.W. 1998. Poisonous plants and fungi in Britain. Animal and human poisoning. The Stationery Office. ISBN 0-11-242981-5. Léveau A.M. et al. 1977. The toxicity of fruits of various <i>Lonicera caprifoliaceae</i> . Plantes Médicinales et Phytothérapie, 11, 2, 94-105. Song W. et al. 2008. Pyridinium alkaloid-coupled scordifidols from the flower buds of <i>Lonicera japonica</i> . J. Nat. Prod. 71, 922-925.
<i>Lophophora williamsii</i> (Salm-Dyck)	Cactaceae	Whole plant	Phenylethylamine alkaloids e.g. mescaline		Frome D., Pfänder H.J. and Anton R. 2009. Plantes à risques. Ed. Tec et Doc-Lavoisier. ISBN: 978-2-7430-0907-1
<i>Luffa</i> spp.	Cucurbitaceae	Aerial part	Genus in which species may contain oxygenated tetracyclic terpenes (cucurbitacins) and ribosome-inactivating proteins. e.g. Luffin A and B, Luffacin	Abortive effect reported in ruminants. <i>Luffa</i> species used by women as a abortifacient. Adverse effect on gestation observed on laboratory animals.	Fernandes L.C.B. et al. 2010. <i>Luffa acutangula</i> Roxb.: tea promotes developmental toxicity in rats. Journal of Animal and Veterinary Advances 9(8), 1255-1258. Lanni J. et al. 2009. Natural and the herb's free at risks: adverse effects, poisonings and other problems related to medicinal herbs by "valerose" in Diademata SP. Rev. Bras. Farmacogn. 19,121-129 Medelir M.G. and Takahashi C.S. 1987. Action of <i>Luffa operculata</i> (Cucurbitaceae) on the chromosomes of <i>Vicia</i> rat. Cytologia 52, 261-285.
<i>Lupinus</i> spp.	Leguminosae (Fabaceae)	Seed	Genus in which species may contain quinolizidine alkaloids - e.g. anagyricine		Lee ST et al. 2007. Lupine induced "crooked calf disease" in Washington and Oregon: identification of the alkaloid proline in <i>Lupinus sativus</i> , <i>Lupinus leucophyllus</i> , and <i>Lupinus sericeus</i> . J. Agric. Food Chem 55(26), 10649–10655. Pilegaard K. and Gryn J. Alkaloids in edible lupin seeds. A toxicological review and recommendations. Temandord 2008. 605. Nordic Council of Ministers. ISBN: 978-92-939-1802-0
<i>Lycium</i> spp.	Solanaceae	Whole plant	Genus in which species may contain tropane alkaloids and/or steroidal alkaloid glycosides.		Adams M. et al. 2006. HPLC-MS/MS trace analysis of atropine in <i>Lycium barbarum</i> berries. Phytochem. Anal. 17, 279-283. Fujyama S. et al. 1995. Kukorimine B, a spermine alkaloid from <i>Lycium chinense</i> . Phytochemistry 38, 1529-1531. Wang K. et al. 2011. Two novel steroidal alkaloid glycosides from the seeds of <i>Lycium barbarum</i> . Chem. Biodivers. 8(12), 2277-2284.
<i>Lycopodium clavatum</i> L.	Lycopodiaceae	Whole plant	<i>Lycopodium</i> alkaloids (0-14.4%), e.g. lycopodine		Wierl M. 2002. Tiedrogen und Phytofarmaka. Ein Handbuch für die Praxis auf wissenschaftlicher Grundlage. Ed. Wissenschaftliche Verlagsgesellschaft mbH. ISBN: 3-8042-1894-X
<i>Lycopodium saururus</i> Lam. (<i>Huperzia saurus</i> (Lam.) Trevis.)	Lycopodiaceae	Aerial part	<i>Lycopodium</i> alkaloids : e.g. sauroxine, lycopine, lycopodine, clavonine	Depending on concentration, decocts of the plant has been the cause of severe adverse effects such as vomiting, diarrhea, convulsions and even death	Cigagna C., Laborde A. 2003. Herbal infusions used for induced abortion. J. Toxicol Clin Toxicol. 41, 235-239. Ortega M.G. et al. 2004. Anticholinesterase activity in an alkaloid extract of <i>Huperzia saururus</i> . Phytomed. 11, 539-543 Ortega M.G. et al. 2006. <i>Huperzia saururus</i> , actively on synaptic transmission in the hippocampus. J. Ethnopharmacol. 104, 374-378.
<i>Lycopodium selago</i> L. See <i>Huperzia selago</i> (L.) Schrank & Mart.					
<i>Lycopus</i> spp.	Lamiaceae (Labiatae)	Leaf		Genus in which species may show antithrombotic effect on thyroid hormones, possibly at the hypophysial level. Other mechanisms may be involved.	Lee W.S. et al. 2006. Human ACOAT-1 and ACOAT-2 inhibitory activities of pentacyclic terpenes from the leaves of <i>Lycopus lucidus</i> (TURCZ. Biol Pharm Bull 28(12),382-384. Beer A.M. et al. 2008. <i>Lycopus europaeus</i> (Spreng.) effects on the thyroidal parameters and symptoms associated with thyroid function. Phytomed. 15(1-2),16-22. Sorgens H. et al. 1992. Antithrombotic effects of plant extracts. Planta Med 48(6),78-86. Aurinko M. et al. 1994. Antithrombotic effects of plant extracts: Piddlythorne deacetylase of rat liver is inhibited by extracts and secondary metabolites of plants. Hom Metab Res. 16(4), 188-92.

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<i>Lycoreis</i> spp.	Amarillidaceae	Whole plant	Genus in which species may contain isoquinoline alkaloids (Amarillidaceae alkaloids); e.g. lycorine		Myasaka K. and Hiratsuyu Y. 1980. Pharmacological studies of lycorine, an alkaloid of <i>Lycoreis radialis</i> herb. . II. Effects of blood pressure in rats and dogs and the mechanism of tachyphylaxis to the vasodpressor action of lycorine in rats. <i>Japan J Pharmacol.</i> 30:655-664. Mu H.M. et al. 2010. Alkaloid accumulation in different parts and ages of <i>Lycoreis chinensis</i> . <i>Z Naturforsch C.</i> 65(7-8):459-462. Jitsuno M. et al. 2011. Chemical constituents of <i>Lycoreis albiflora</i> and their cytotoxic activities. <i>Natl Prod Commun</i> 6(2):187-192. Fornie D., Pflander H. J. et Anton R. 2009. <i>Plantas a rasques</i> . Ed. Tec et Doc-Lavoisier. ISBN: 978-2-7430-0907-1
<i>Lyonia</i> spp.	Ericaceae	Whole plant	Genus in which species may contain toxic diterpenes; e.g. andromedotoxin (acetyl-andromedol)		Hardman R. (Ed). 2002. <i>Medicinal and Aromatic Plants—Industrial Profiles</i> . D. Sarker and Viji Manjeyam. Magnolia, the genus <i>Magnolia</i> . Taylor & Francis. ISBN 0-415-28294-5 Nagase H. et al. Inhibitory effect on magnoliol and honokiol from <i>Magnolia obovata</i> on human thiosarcosina HT-1080 invasiveness in vitro. 2001. <i>Planta Med</i> 67, 705-709. Cordell G.A. 2000. Alkaloids of the Menispermaceae. University of Illinois. ISBN 0099-9698/00
<i>Magnolia</i> spp.	Magnolaceae	Whole plant	Genus in which species may contain lignans; e.g. honokiol, magnolol and benzylisoquinoline alkaloids; e.g. magnoflorine and quaternary ammonium; e.g. magnocouramine.	Because of the quaternary ammonium structure of the magnocouramine, absorption is unlikely.	Genakova M. and Kostalova D. 2002. Antimicrobial activity of berberine - a constituent of <i>Magnolia aquilifolium</i> . <i>Folia Microbiol (Praha)</i> , 47(4), 375-378 Fornie D., Pflander H. J. et Anton R. 2009. <i>Plantas a rasques</i> . Ed. Tec et Doc-Lavoisier. ISBN: 978-2-7430-0907-1 Roh L., Daurer M. and Korman K. 1984. Giftphalanz - Pflanzenzengle. <i>Vorkommen Wirkuna Therapie</i> . ecomed ISBN 3-609-04810-4 Klinger JT et al. 2007. Rotenon causes pulmonary edema in vivo: a possible role for PKCdelta. <i>J Appl Physiol.</i> 102(6), 2004-2004 Thakur S.C. et al. 2005. An ethereal extract of <i>Kamela (Malolus philippensis)</i> (Mol. Arg.) Lam.) induce adverse effects on reproductive parameters of female rats. <i>Reprod Tox.</i> 20, 148-156.
<i>Magnolia aquilifolium</i> (Pursh) Nutt.	Berberidaceae	Root and stem bark	Isoquinoline alkaloids; e.g. magnoflorine, isobellane and isopyridine, berberine, oxycarpine		Bruneton J. 2009. <i>Pharmacognosie, (Phytochimie, Plantes médicinales)</i> . Ed. Tec & Doc-Lavoisier. Paris, 4ème édition. ISBN: 978-2-7430-1188-8 Bruneton J. 2005. <i>Plantes toxiques (Végétaux dangereux pour l'homme et les animaux)</i> . Ed. Tec & Doc-Lavoisier. Paris, 2ème édition. ISBN: 2-7430-0906-7 Council of Europe. 2005. <i>Active principles (constituents of chemical concern) contained in natural sources of flavonings</i> . Ed. Council of Europe Publishing. http://www.coe.int/t/escsoai_consensoc-sopublic/NeatlyFlavoungs_substancesActive%20principles.pdf Padmaja G. 1995. Cyanide detoxification in cassava for food and feed uses. <i>Chl Rev Food Sci Nutr.</i> 31(4), 299-339 Berger S. et al. 1984. Structural revision of pregnane ester glycosides from <i>condurango cortex</i> and new compounds. <i>Phytochemistry</i> 21(5), 1451-1459 Slovak J. Stavkova L. 2009. Alkaloids of <i>Meconopsis cambrica</i> (L.) VIG. and <i>M. robusta</i> HOOK. f. et THOM. <i>Collection Czechoslovak Chemical Communications</i> 61, 12, 1815-1822
<i>Mandragora officinarum</i> L. (M. autumnalis Bertol., M. acaulis Gaertn., M. vernalis Bertol.)	Scianaceae	Whole plant	Root: 0.4% tropane alkaloids; e.g. scopolamine, L-hyoscyamine		Barnes J., Anderson L. A., Phillips J.D. 2007. <i>Herbal Medicines</i> . 3rd ed. Ed. Pharmaceutical Press. ISBN 978-0-35369-623-0 Bruneton J. 2009. <i>Pharmacognosie, Phytochimie, Plantes médicinales</i> . Ed. Tec & Doc-Lavoisier. Paris, 4ème édition. ISBN: 978-2-7430-1188-8 Natural Sources of Flavonings Report No. 2, 2007. Ed. Council of Europe Publishing. ISBN 978-92-87-14156-7 Natural Sources of Flavonings Report No. 3, 2008. Ed. Council of Europe Publishing. ISBN 978-92-87-14422-3 Olefiants P.B. et al. 1983. Toxic tetranortriterpenes of the fruits of <i>M. azedarach</i> . <i>Phytochemistry</i> 22 (2), 531-534. Del C. et al. 2002. <i>Inotoxicacao experimental pelis fofhas de M. azedarach em bovinos</i> . <i>Pesqui Vet. Brasil</i> 22 (1), 19-24.
<i>Meconopsis</i> spp.	Papaveraceae	Whole plant	Genus in which species may contain isoquinoline alkaloids; e.g. meconbrine		Wicrli M. and Anton R. 2003. <i>Plantes tetrapépiques (Tradition, pratique officinale science et thérapeutique)</i> . Ed. Tec & Doc-Lavoisier. Paris, 2ème édition. ISBN : 2-7430-0631-5 Puschner B. et al. 1998. Sweet clover poisoning in dairy cattle in California. <i>J Am Vet Med Assoc</i> 15, 21(6):387-389 Marino E. et al. 2006. Microwave assisted extraction of coumarin and related compounds from <i>Melilotus officinalis</i> (L.) Palas as an alternative to Soxhlet and ultrasound-assisted extraction. <i>J Chromatogr A.</i> 1129, 147-151 De Vincenzi M. et al. 1997. Monographs on botanical flavonoid substances used in food. <i>Fitoterapia</i> , 68, 50-1 Magri F. et al. 2011. HPLC quantification of coumarin in bastard balm (<i>Melilotis melissophyllum</i> L., Lamiales). <i>Fitoquímica</i> 62, 121-5-1221.
<i>Melilotus</i> spp.	Leguminosae (Fabaceae)	Aerial part	Genus in which species may contain coumarin glycosides (e.g. melilotoside)	Coumarin may be formed from melilotoside upon drying, up to 0.4-0.9%. (E.g. 4mg of coumarin after drying flowering tops of <i>M. officinalis</i>). Improper drying may yield dihydrocoumarol, a fungal metabolite from coumarin, which may give rise to hemostatic dysfunction.	
<i>Melilotus melissophyllum</i> L.	Lamiaceae (Labiatae)	Aerial part	Coumarin (2.6-7.0 g/kg in fresh leaves and 0.3-2.5 g/kg in dry leaves)		De Vincenzi M. et al. 1997. Monographs on botanical flavonoid substances used in food. <i>Fitoterapia</i> , 68, 50-1 Magri F. et al. 2011. HPLC quantification of coumarin in bastard balm (<i>Melilotis melissophyllum</i> L., Lamiales). <i>Fitoquímica</i> 62, 121-5-1221.
<i>Menispermum canadense</i> L.	Menispermaceae	Fruit and root	Isoquinoline alkaloids		Maniske R. H. et al. 1985. Studies on the alkaloids of menispermaceous plants. <i>CCXX</i> . <i>Daunche</i> from <i>Menispermum canadense</i> L. <i>Chem Pharm Bull (Tokyo)</i> 13 (12), 1476-7.
<i>Menispermum dauricum</i> DC.	Menispermaceae	Aerial part	Bisbenzyltetrahydroisoquinoline alkaloids; e.g. dauricine		Bruneton J. <i>Pharmacognosy, Phytochemistry, Medicinal Plants</i> . 2nd ed. 1999. Ed. Interleaf Ltd. ISBN: 1-58928-653-7
<i>Mentha canadensis</i> L. (M. <i>arvensis</i> var. <i>obovata</i> Muhlrv. ex Hornem.)	Lamiaceae (Labiatae)	Aerial part	Essential oil: monoterpene etheroxide: 1,8-cineole (2.4-18.5%), monocylic monoterpene ketones: e.g. pulegone and bicyclic monoterpene ketones: e.g. pulegone (0.1-5.4%), bicyclic monoterpene mentholidian (0.1-7.4%), and coumarin		Natural Sources of Flavonings Report No. 3, 2008. Ed. Council of Europe Publishing. ISBN 978-92-87-14422-3
<i>Mentha piperita</i> L.	Lamiaceae (Labiatae)	Aerial part	Essential oil: monoterpene etheroxide: 1,8-cineole (2.4-18.5%), monocylic monoterpene ketones: e.g. pulegone (0.1-5.4%), bicyclic monoterpene mentholidian (0.1-7.4%), and coumarin		Natural Sources of Flavonings Report No. 3, 2008. Ed. Council of Europe Publishing. ISBN 978-92-87-14422-3

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<i>Mentha pulegioides</i> L.	Lamiaceae (Labiatae)	Aerial part	Essential oil: monoterpenic monoterpene ketones: e.g. pulegone (71.3-90%), bicyclic monoterpenes: menthulone, thujones and monoterpene etheroxide: 1,8-cineole		Natural Sources of Flavonoids Report No. 3, 2008. Ed. Council of Europe Publishing. ISBN 978-92-87-14422-3
<i>Mentha spicata</i> L. (<i>Mentha viridis</i> (L.) L.)	Lamiaceae (Labiatae)	Aerial part	Essential oil: monoterpenic monoterpene ketone: e.g. pulegone (1.7-1.9%) and monoterpene etheroxide: 1,8-cineole (6-6.5%) Essential oil: chemo-type carvone: 1,8-cineole (0.5%) Essential oil: chemo-type dihydrocarvyl acetate: 1,8-cineole (2.2%)		Natural Sources of Flavonoids Report No. 3, 2008. Ed. Council of Europe Publishing. ISBN 978-92-87-14422-3
<i>Menthaefolia</i> L.	Menyanthaceae	Leaf	Anthraquinones: e.g. emodin, alcea-emodin, chrysophanol; Coumarins: e.g. coumarin, scopoletin	Reports on monoterpene alkaloids gentianin and gentianidin may be artefacts (CofE, 2007)	Capasso R. et al. 2000. Phytotherapy and quality of herbal medicines. <i>Fitoterapia</i> 71(S), 56-65 Natural Sources of Flavonoids Report No. 2, 2007. Ed. Council of Europe Publishing. ISBN 978-92-87-14516-7
<i>Mercurialis</i> spp.	Euphorbiaceae	Whole plant	Genus in which species may contain cocarbinogenic diterpenes: e.g. ingenol esters		Frome D., Pfander H.J. and Anton R. 2009. <i>Plantes à risques</i> . Ed. Ter et Doc. Lavoisier. ISBN: 978-2-7430-0097-1
<i>Mesembryanthemum</i> spp.	Aizoaceae	Aerial part	Genus in which species may contain iridoid alkaloids: e.g. mesembryne, and oxalic acid	<i>Mesembryanthemum</i> spp. are now named <i>Scolecium</i> spp. <i>Mesembryne</i> in <i>S. expansum</i> , <i>S. tortuosum</i> and <i>S. anatumicum</i>	Brurton J. Pharmacognosy. Phytochemistry. Medicinal Plants, 2nd ed. 1999. Ed. Intersci Ltd. ISBN: 1589298-63-7
<i>Michelia hedyosperma</i> V.W.Law	Magnoliaceae	Unspecified	Essential oil reported to contain the phenylpropanoid methyl Eugenol in unspecified quantities		EMEA-HMPc. 2004. Final position paper on the use of herbal medicinal products containing methyl Eugenol. EMEA/HMPc/232703
<i>Milletia gibbifera</i> Adema	Leguminosae (Faboaceae)	Root	Rotenoids: e.g. rotenone		Brurton J. Pharmacognosy. Phytochemistry. Medicinal Plants, 2nd ed. 1999. Ed. Intersci Ltd. ISBN: 1589298-63-7
<i>Mimosa</i> spp.	Leguminosae (Faboaceae)	Aerial part	Genus in which species may contain non-protein amino acids: e.g. mimosine and mimosides	Teratogenic effect described for <i>M. tenuiflora</i> (Willd.) Pol.	Medeiros R.T.M. et al. 2008. Teratogenicity of <i>Mimosa tenuiflora</i> seeds to pregnant rats. <i>Toxicol</i> 51: 316-319. Jiang Y. et al. 1992. Effects of saponins from <i>M. tenuiflora</i> on lymphoma cells and lymphocytes. <i>Phytother. Res.</i> 6 (6): 310-313.
<i>Milragya speciosa</i> Korh.	Rubiaceae	Whole plant	Iridoid-monoterpene alkaloids in leaf: e.g. mitraglyne (accounting for 2/3 of alkaloids present) and 7-hydroxymitraglyne		Kumaratil E. et al. 2008. Acute and long-term effects of alkaloid extracts of <i>Milragya speciosa</i> on food and water intake and body weight in rats. <i>Fitoterapia</i> 77: 338-345.
<i>Momordica charantia</i> L. (<i>M. chinensis</i> , <i>M. edulis</i> , <i>M. indica</i> , <i>M. opoculata</i> , <i>M. stenosia</i>)	Cucurbitaceae	Aerial part	Cucurbitane triterpenoids: momordicosides and momordicins; Seeds: a lectin: momodin	Some seed extracts showed antispermatogenic activity in rats	Chiang C.-I. 2008. Cucurbitane-type triterpenoids from <i>Momordica charantia</i> . <i>J Nat Prod</i> 71: 1327-1330. Fátore M.O. 1990. New cucurbitane triterpenoids from <i>Momordica charantia</i> . <i>J Nat Prod</i> 53: 1491-1497. Naseem M.Z. et al. 1998. Antispermatogenic and androgenic activities of <i>Momordica charantia</i> (Kasturi). <i>Indian J. Ethnopharmacol</i> 61: 59-66. Barnett J.W. and Kirch M. 2003. <i>Mycobionics</i> . <i>Clinical Microbiology Reviews</i> 16(3): 497-516.
<i>Morus purpureus</i>	Moraceae	Microfungi	May produce citrinin (mycotoxin)		Pedersen M.E. et al. 2008. Effects of South African traditional medicine in animal models for depression. <i>J Ethnopharmacol</i> 119: 542-548. Patrian R. et al. 2005. A chlorinated coumarinignan from the African medicinal plant, <i>Morinda whitei</i> . <i>Phytochemistry</i> 66:683-686. Kubo I. and Kuri-Hara I. 1999. 2-Hydroxy-4-methoxybenzaldehyde: a potent tyrosinase inhibitor from African medicinal plants. <i>Plant Med</i> 1999: 66(1):19-22 Valcho P. et al. 2004. Androgenic effects of the aqueous extract of the roots of <i>Morinda whitei</i> in rats. <i>Asian J. Androl</i> 6(3): 269-72
<i>Morinda whitei</i> (Hook.) Skeels	Apocynaceae	Whole plant	Root: chlorinated coumarinignan: 5-chloropropan and phenolic: 2-hydroxy-4-methoxybenzaldehyde (=p-anisaldehyde)	Root given orally had androgenic effect on male rats.	Chenke P.R. 1989. <i>Toxicants of Plant Origin</i> . Volume IV. <i>Phenolics</i> . CRC Press Inc. ISBN 0-8493-6693-2. Lundgren B.M. et al. 1979. Clinical effect of orally administered extracts of <i>Morinda tomentosa</i> in early human pregnancy. <i>Toxicol</i> 135: 480-484. Robiez-Zepeda R.E. et al. 2009. <i>Morinda tomentosa</i> glandular trichomes containing kaurenoic acids: chemical profile and distribution. <i>Flotetaria</i> 80: 12-17.
<i>Morinda tomentosa</i> Cav.	Compositae (Asteraceae)	Whole plant	Oxepane diterpenoids in leaf: e.g. zoapatanol and morindol and kaurenoic acids		Shukla S. et al. 1988. Antifertility profile of the aqueous extract of <i>Moringa oleifera</i> roots. <i>J Ethnopharmacol</i> 22 (1): 51-62. Dangi S.Y. et al. 2002. Antihypertensive activity of the total alkaloids from the leaves of <i>Moringa oleifera</i> . <i>Pharm. Biol</i> 40(2): 144-148 Mifru-Solez Z. et al. 2010. Chronic benzoxazine administration in the drinking water improves glucose tolerance, reduces body weight gain and circulating cholesterol in high-fat diet-fed mice. <i>Pharmazie Res</i> 61(14): 355-363.
<i>Moringa oleifera</i> Lam.	Moringaceae	Root and wood	Alkaloids (benzylamines) in root bark (0.1%), e.g. moringine (synonym of benzylamine) and moringine	Antifertility effects of aqueous extract of roots have been reported	Infrate M.E. et al. 1990. Outcome of acute toxic psychosis attributed to <i>Mazuna pruriens</i> . <i>Lancet</i> 336: 1125. Miral L. and Wagner H. 2004. Alkaloid constituents of <i>Mazuna pruriens</i> seeds. <i>Phytochemistry</i> 65: 2565-2567.
<i>Mucuna pruriens</i> (L.) DC.	Leguminosae	Whole plant	Seeds: amino acids: L-dopamine (3.6-5.4%) Iridoid alkaloids (tyrptamine derivatives): e.g. NN-dimethyltyrptamine, bufotenine, 5-methoxy-NN-dimethyltyrptamine.		Raposo J.B. et al. 1998. Experimental intoxication by <i>Myzoreum laetum</i> in sheep. <i>Vet Human Toxicol</i> 40:132-135.
<i>Myzoreum laetum</i> G.Forst.	Scrophulariaceae	Leaf	Essential oil: furanoid sesquiterpene ketones: gonalone		Brurton J. Pharmacognosy. Phytochemistry. Medicinal Plants, 2nd ed. 1999. Ed. Intersci Ltd. ISBN: 1589298-63-7
<i>Mycosila</i> spp.	Boraginaceae	Aerial part	Genus in which species may contain unsaturated pyrrolizidine alkaloids.		Resch J.F. et al. 1992. Biologically active pyrrolizidine alkaloids from the foggy-needle <i>Mycosila scoroidalis</i> . <i>J Nat Prod</i> 5(3):358-62
<i>Myristica fragrans</i> Hook. (<i>M. mosata</i> Thunb., <i>M. officinalis</i> L.)	Myristicaceae	Mace and seed	Essential oil from seed: phenylpropanoids: e.g. eilmnon (up to 7.5%), myristicin (1.3% in the seed and 2.7% in mace), safrole		Council of Europe. 2005. <i>Active principles (constituents of chemical concern) contained in natural sources of flavourings</i> . Ed. Council of Europe Publishing. http://www.coe.int/t/09social_consensus/active_principles/health/flavourings_substances/Active%20principles.pdf EC SCF. 2001. Opinion of the EC Scientific Committee on Food on the safety of the presence of safrole in flavourings and other food ingredients with flavouring properties. http://europe.eu.int/comm/food/fs/sc/sf/index_en.html

COMPENDIUM OF BOTANICALS REPORTED TO CONTAIN NATURALLY OCCURRING SUBSTANCES OF POSSIBLE CONCERN FOR HUMAN HEALTH WHEN USED IN FOOD AND FOOD SUPPLEMENTS

This compendium lists in alphabetical order botanicals without any judgment on whether these are suitable or not suitable for food applications in Europe. This compendium is part of a preliminary work undertaken by EFSA to harmonise the methodology across its panels for assessing the safety of botanicals and botanical preparations used in food and food supplements. Without prejudice to the existing legal framework, such compendium has no legal status and may not be used as support or evidence in any disagreement or dispute pertaining to the legal classification of products or substances. This compendium is a living document and is therefore open for additional contributions and comments.

Botanical name	Family	Parts of plants of possible concern	Chemical of concern	Remarks on toxic/adverse effect(s) not known to be related to the identified chemical(s) of concern	Specific References
<i>Myrrhis odorata</i> (L.) Scop.	Apiaceae (Umbelliferae)	Whole plant	Essential oil from fruit: phenylpropanoids: e.g. trans-anethole (76-85%), methyl Eugenol, methylchavicol (1.2-1.7%) Essential oil from leaf: e.g. trans-anethole (82-85%)		Council of Europe. 2005. Active principles (constituents of chemical concern) contained in natural sources of flavonoids. Ed. Council of Europe Publishing http://www.coe.int/t/e/scp/consensus/ncp/synoptic_health/Flavonoid_substances/Active%20principles.pdf Natural Sources of Flavonoids Report No. 2. 2007. Ed. Council of Europe Publishing. ISBN 978-92-871-4516-7
<i>Myrtus communis</i> L.	Myrtaceae	Aerial part	Essential oil: phenylpropanoids: methylchavicol (58-88 ppm), methyl Eugenol (0.2%-6%)		Teuscher E., Anton R. et al. 2005. <i>Plantas aromáticas</i> (Epicés, aromáticas, condimentos et hules essentiales). Ed. Tec et Doc-Lavoisier. ISBN : 2-7430-0720-6
<i>Narcissus</i> spp.	Amaryllidaceae	Whole plant	Genus in which species may contain isouranoline alkaloids (Amaryllidaceae alkaloids): e.g. lycorine, galanthamine, homonocorine, haemanthamine.		Burleton J. Pharmacognosy. Phytochemistry. Medicinal Plants 2nd ed. 1999. Ed. Interscienc Ltd. ISBN 1-898292-63-7
<i>Nepeta cataria</i> L.	Lamiaceae (Labiatae)	Aerial part	Essential oil: bicyclic monoterpenes: e.g. camphor		Natural Sources of Flavonoids Report No. 1. 2000. Ed. Council of Europe Publishing. ISBN 92-871-4324-2
<i>Nerium</i> spp.	Apocynaceae	Whole plant	Genus in which species may contain cardenolide glycosides: e.g. strophanthidin, oleandrin, ...		Burleton J. 2005. <i>Plantes toxiques (Végétaux dangereux pour l'homme et les animaux)</i> . Ed. Tec & Doc. Lavoisier. Paris, 3ème édition. 618 pages. ISBN: 2-7430-0806-7
<i>Nicotiana</i> spp.	Solanaceae	Whole plant	Genus in which species may contain pyridine alkaloids: e.g. nicotine and anabasine	In <i>Nicotiana glauca</i> , 99% of the alkaloids is anabasine	PDR for Herbal Medicines. 2004 Thomson ed. ISBN: 1-56363-5125-7
<i>Nierembergia aristata</i> D.Don	Scitaneaceae	Whole plant	Cardenolides: e.g. 17-epi-11 alpha-hydroxy-5,7-dehydrostrophanthidin-3-O-beta-D-hominyranoside, 6,7-dehydrostrophanthidin-3-O-beta-D-hominyranoside, 6,7-dehydrostrophanthidin-3-O-beta-D-glucopyranoside		Burleton J. 2005. <i>Plantes toxiques (Végétaux dangereux pour l'homme et les animaux)</i> . Ed. Tec & Doc. Lavoisier. Paris, 3ème édition. 618 pages. ISBN: 2-7430-0806-7
<i>Nierembergia velutina</i> Berkley ex Hook.	Solanaceae	Whole plant	Proto-alkaloid: e.g. damascenine (=ingelone)		GH R. R. et al. 1995. Cardenolides from <i>Nierembergia aristata</i> . J. Natl. Prod. 58(6): 848-56
<i>Nigella damascena</i> L.	Ranunculaceae	seed			Riet-Correa F. et al. 1993. Ergolic calcosin in sheep: experimental reproduction with <i>N. velutina</i> . <i>Presq. Vét. Bras.</i> 13(1,2): 21-24.
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
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<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
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<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
<i>Nigella arvensis</i> L.	Ranunculaceae	seed			Morrell A. et al. 2004. Essential Oils of <i>Nigella arvensis</i> L. and <i>Nigella damascena</i> L. Seed J Ess Oil Res. May/Jun. http://dx.doi.org/10.1002/essoc.10049
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Botanical name	Family	parts of plants of possible concern	Chemical of concern	Remarks on toxic/adverse effect(s) not known to be related to the identified chemical(s) of concern	Specific References
<i>Ocimum sancto</i> Benth.	Lamiaceae (Labiatae)	Aerial part	Essential oil (51.1%); phenylpropanoids e.g. methylchavicol (94.95% in essential oil from the leaf), 92.54% in essential oil from flower)		Martini M.G. Et al. 2011. Chemical composition and antimicrobial activities of the essential oils from <i>Ocimum sancto</i> and <i>Hesperozygia myrtoides</i> . <i>Nat Prod Commun.</i> 6(7), 1027-1030. Nasrimento J.C. et al. 2011. Chemical composition and antimicrobial activity of essential oils of <i>Ocimum canum</i> Sims. and <i>Ocimum sancto</i> Benth. <i>An Acad Bras Cienc.</i> 83(3):767-799.
<i>Ocimum suave</i> Willd.	Lamiaceae (Labiatae)	Aerial part	Essential oil (2%); phenylpropanoids: methylchavicol (65.49%; 66.18 in leaf and flower oil; 2.240ppm in bud)		EME/ANMP/138383/2005. 2005. Public statement on the use of herbal medicinal products containing methylchavicol. Available at: http://www.ema.europa.eu/docs/en_GB/document_library/Scientific_guideline/2010/04/WC500089961.pdf
<i>Ocimum tenuiflorum</i> L. (<i>Ocimum sanctum</i> L.)	Lamiaceae (Labiatae)	Whole plant	Essential oil: phenylpropanoids: methylchavicol (39.950 ppm in leaf), methylchavicol (15-100 ppm in plant and 50ppm in leaf)		EFSA Scientific Cooperation (ESCO) Working Group on Botanicals and Botanical Preparations. EFSA guidance document for the safety assessment of botanicals and botanical preparations intended for use as food supplements, based on real case studies on request of EFSA. EFSA Journal 2009; 7(9):280
<i>Ocotea odorifera</i> (Vell.) Rohmer (<i>Ocotea pretiosa</i> (Nees) Mez.)	Lauraceae	Wood	Essential oil: phenylpropanoids e.g. methylchavicol (0.1-78%), safrole		EME/ANMP/138383/2005. 2005. Public statement on the use of herbal medicinal products containing methylchavicol. Available at: http://www.ema.europa.eu/docs/en_GB/document_library/Scientific_guideline/2010/04/WC500089961.pdf Perera E.F. et al. 1989. Anti-inflammatory properties of new bioisosteres of indomethacin synthesized from safrole which are sulfinate analogues. <i>Braz J Med Biol Res.</i> 22(11), 1415-9.
<i>Oenanthe aquatica</i> (L.) Poit.	Apiaceae (Umbelliferae)	Fruit and root	Polyacetylene derivatives: e.g. oenanthaloxin. In fruit: phenylpropanoids: e.g. myristicin		Frohe D., Pfander H.J. and Anton R. 2009. <i>Plantae 3</i> inquest. Ed: Tec et Doc-Lavoisier, ISBN: 978-2-7430-0907-1 Cooper M.R. and Johnson A.W. 1998. <i>Poisonous plants and fungi in Britain. A manual and human poisoning.</i> The Stationery Office, ISBN 0-11-242814-5. Vincet F.F. et al. 1976. Composition of the <i>Oenanthe aquatica</i> essential oil. <i>Planta Med</i> 29, 101-112.
<i>Oenanthe crocata</i> L.	Apiaceae (Umbelliferae)	Whole plant	Polyacetylene derivatives: e.g. oenanthaloxin, 1-oenanthenol and 14-desac-oenanthaloxin		Burtonet J. 2005. <i>Plantae toxiques (Végétaux dangereux pour l'homme et les animaux)</i> . Ed: Tec & Doc, Lavoisier, Paris, 3ème édition, ISBN : 2-7430-0806-7 Jens Jacob J. 2009. <i>Wundheilerforschung in der Kulturtradition - Grundlagen und Anwendung.</i> Julius Kühn-Institut Bundesforschungsanstalt für Kulturpflanzen, Arno Bryda GmbH, Berlin, ISBN 978-3-930037-58-2. Schlep L.J. et al. 2009. <i>Poisoning due to water hemlock.</i> <i>Clin Tox</i> 47: 270-278.
<i>Oenanthe luteola</i> L.	Apiaceae (Umbelliferae)	Whole plant			Gran L. et al. 2008. Cyclic peptides from <i>Oenanthe affinis</i> DC. Molecular and biological properties. <i>Chem & Biodiversity</i> 5:2014-2022. Gran L. et al. 2000. <i>Oleandra affinis</i> (R&S) DC. A plant containing terpenoid peptides used in african traditional medicine. <i>Journal Ethnopharmacol.</i> 70(3):197-203 Crak D. J. et al. 2010. Cyclic dipeptides: macrocyclic peptides with applications in drug design and agriculture. <i>Cell Mol Life Sci.</i> 67:9-16. Dornenburg H. 2010. Cyclic dipeptide synthesis and supply: From plant to bioprocess. <i>Biopolymers.</i> 94(5), 602-10.
<i>Oenocaulis macrotarpa</i> (L.) Urb. (<i>Oenocaulis operculata</i> (Gomès) Mart., <i>Merremia macrotarpa</i> (L.) Robinson)	Convolvulaceae	Root	Glycoresin (10%); e.g. oenocaulinic acid C		Ono M. et al. 2009. Components of ether-insoluble resin glycosides (pharmacovulfin) from <i>hizoma</i> (japanese brazilianis. <i>Chem. Pharm. Bull (Tokyo)</i> 57(3): 282-288
<i>Oenocaulis lupaninum</i> (L.) S.Manso	Convolvulaceae	Root	Glycoresin (4%); e.g. lupanin		Burtonet J. 2005. <i>Plantae toxiques (Végétaux dangereux pour l'homme et les animaux)</i> . Ed: Tec & Doc, Lavoisier, Paris, 3ème édition, ISBN : 2-7430-0806-7
<i>Oenanthe ssp.</i>	Apiaceae (Umbelliferae)	Whole plant	Genus in which species may contain tannocoumarins		Aperntino G. et al. 2004. Coumarins from <i>Oenanthe chinensis</i> New dityroleanocoumarins and differential induction of apoptosis by imipenem and berberetin. <i>J Nat Prod.</i> 67(2):532-536.
<i>Oeniganum majonana</i> L.	Lamiaceae (Labiatae)	Aerial part	Essential oil: bicyclic monoterpenes: e.g. camphor (2%) and phenylpropanoids: e.g. methylchavicol (95-550 ppm).		Council of Europe. 2005. Active principles (constituents of chemical concern) contained in natural sources of flavonoids. Ed: Council of Europe Publishing. http://www.coe.int/t/cesoc/active%20principles.pdf
<i>Oeniganum vulgare</i> L.	Lamiaceae (Labiatae)	Aerial part	Essential oil: bicyclic monoterpene: beta-thujone (0-0.6%), monoterpene etheroxide: 1-8-cineole (0-6.5%)		Verna R.S. et al. 2010. Chemical diversity in <i>Indian oregano</i> (<i>Oeniganum vulgare</i> L.). <i>Chemistry and Biodiversity.</i> 7: 2054-2064.
<i>Orthostegium ssp.</i>	Asparagaceae	Whole plant	Genus in which species may contain cardenolides: e.g. samentholoside		Ghanmany V. et al. 1987. Cardenolides from <i>O. bouchéanum</i> . <i>Planta med.</i> 53(2): 172-178.
<i>Orobanchaceae ssp.</i>	Orobanchaceae	Whole plant	Genus in which species may contain oxalates		Frohe D. and Pfander H.J. 1997. Giftpflanzen. Ein Handbuch für Apotheker, Ärzte, Toxikologen und Biologen. Wissenschaftliche Verlagsgesellschaft mbH, ISBN 3-8047-1465-8. Roth L., Daubert M. and Kormanik K. 1994. Giftpflanzen - Pflanzengifte. Vorkommen, Wirkung, Therapie, edited. ISBN 3-809-54810-4. Medicinal and Aromatic Plants XI. <i>Vagaria</i> . Toshiyuki, Ebizuka, Y. (Eds.). Springer Verlag 2002. ISBN 978-3-540-41695-9 PDR for Herbal Medicines. 2004. Thomson ed. ISBN: 1-56363-5126-7
<i>Oxalis ssp.</i>	Oxalidaceae	Aerial part	Genus in which species may contain oxalates		Parasitic plant using sap from host plant. If toxic compounds are present in the host, they may also be found in <i>Oxobancha</i> .
<i>Pachera aurea</i> (L.) A. Löve & D. Löve (<i>Seneo aureus</i> (L.)	Compositae (Asteraceae)	Aerial part	Unsaturated pyridizidine alkaloids: e.g. senecholine.		Röder E. et al. 1983. Pyridizidine alkaloids aus <i>Seneo aureus</i> . <i>Planta Med.</i> 49 (9): 57-59.
<i>Papaaver ssp.</i>	Papaveraceae	Whole plant	Genus in which species may contain isoquinoline alkaloids (morphinanes); e.g. morphine, codeine, thebaine		Frohe D., Pfander H.J. and Anton R. 2009. <i>Plantae 3</i> inquest. Ed: Tec et Doc-Lavoisier, ISBN: 978-2-7430-0907-1
<i>Paris quadrifolia</i> L.	Melasthaceae	Whole plant	Steroid and spirostane saponins: e.g. hemogenin tetraglycoside		Cooper M.R. and Johnson A.W. 1998. <i>Poisonous plants and fungi in Britain. A manual and human poisoning.</i> The Stationery Office, ISBN 0-11-242814-5. Frohe D., Pfander H.J. and Anton R. 2009. <i>Plantae 3</i> inquest. Ed: Tec et Doc-Lavoisier, ISBN: 978-2-7430-0907-1

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<i>Peucedanum quinquiflorum</i> (L.) Panich.	Vitaceae	Leaf	Calciumoxalate raphides (up to 2%)	Cases have been reported of illness or death resulting after berries of Juice from leaves of Virginia creeper were ingested. Modern reports view these cases as circumstantial	Fuller, T. G., McChickok, E. 1986. Poisonous plants of California. Univ. California Press, Berkeley, Calif., USA. ISBN: 0-520-05669-1
<i>Psyllium cuspana</i> Kunth	Sapindaceae	Seed	Methylated xanthine derivatives: e.g. caffeine (3.0-4.8% dry weight), Essential oil: phenylpropanoids: e.g. methylchavicol, anethole		Andersson H. C., Hallstrom H., Kihlman B. A. 2004. Intake of caffeine and other methylxanthines during pregnancy and risk for adverse effects in pregnant women and their foetuses. <i>Terrahord</i> 2004:565. Nordic Council of Ministers. ISBN 92-893-1098-7 Natural Sources of Flavonoids Report No. 2. 2007. Ed. Council of Europe Publishing. ISBN 978-92-871-4516-7 Crozier A., Ashihara H., Tomás-Barbáñan F. 2012. 'Teas, cocoa and coffee. Part 1: secondary metabolites and health'. Blackwell Publishing Ltd. ISBN: 1-3: 978-1-4443-3441-8. ISBN-10: 1-4443-3441-7.
<i>Psylliostictia jolimbhe</i> (K. Schum.) Pierre ex Belle (Coryphantha jolimbhe K. Schum.)	Rubiaceae	Whole plant	Indole alkaloids (yohimbanes) in bark: e.g. yohimbine (=corynine, quebrachine), alpha-yohimbine (=corynanthidine, isoyohimbine), beta-yohimbine, delta-yohimbine (=(-)-gimnaine), corynanthine, corynanthene, dihydrocorynanthene, allo-yohimbine (=dihydroyohimbine), pseudo-yohimbine and tetrahydrocorynanthene.		Mongkolkeha W. and Suttivayakul S. 2007. Antimalarial and antidiabetic poly-O-acetylated jatrophone diterpenoids from <i>Pedilanthus tithymaloides</i> . <i>J Nat Prod</i> . 70(9). 1434-1438. Bruneton J. 2005. <i>Plantes toxiques (Végétaux dangereux pour l'homme et les animaux)</i> . Ed. Tec & Doc, Lavoisier, Paris, 3ème édition, 618 pages. ISBN : 2-7430-0806-7 Massoud M. et al. 2002. Toxicity of <i>Pegannum harmala</i> : review and a case report. <i>Iranian J Pharm Therap</i> . 1: 1-4. Kaiser O. et al. 1998. Composition of the essential oils of <i>Palaegonium stictodes</i> DC. and <i>Palaegonium reniforme</i> Curt. <i>Flavour Frag J</i> . 13(3). 203-213. Kozuka Y. et al. 1985. An antispasmodic substance from <i>Perilla frutescens</i> and its mechanism of action. <i>Phanta Med</i> . 6:480-482. Seto T.A. and Keup W. 1969. Effects of alkylmethoxybenzene and alkylmethylendioxybenzene essential oils on pentobarbital and ethanol sleeping time. <i>Arch. Int. Pharmacodyn</i> . 180:323-240. Bruneton J. 2009. <i>Pharmacognosie. (Phytochimie, Plantes médicinales)</i> . Ed. Tec & Doc, Lavoisier, Paris, 4ème édition. ISBN: 978-2-7430-1188-8
<i>Pedilanthus</i> spp.	Euphorbiaceae	Whole plant	Genus in which species may contain cytotoxic diterpenes : e.g. oxygenated jatrophone diterpenes, phenolsters,...		
<i>Pegannum harmala</i> L.	Nitifariaceae	Whole plant	Indole alkaloids (beta-carbolines): e.g. harmine, harmaline, and quinoline alkaloids: e.g. vasicine, vasicinone		
<i>Palaegonium stictodes</i> DC.	Geraniaceae	Leaf	Essential oil: phenylpropanoids: e.g. methyl Eugenol (4.3%) and elemicin (3.6%)		
<i>Perilla frutescens</i> Britton	Lamiaceae (Labiatae)	Leaf and seed	The phenylpropanoid chemo-type contains myristicin	Part must be properly dried to avoid the appearance of the toxic perilla ketone	
<i>Persea americana</i> Mill (Persea edulis) Schubl. & Cham)	Lauraceae	Leaf	Essential oil: phenylpropanoids: e.g. methyl Eugenol (3-8%)		
<i>Pezales</i> spp.	Compositae (Asteraceae)	Whole plant	Genus in which species may contain unsaturated pyrolicidic alkaloids		
<i>Peucedanum crispum</i> (Mill.) A.W.Hill	Apiaceae (Umbelliferae)	Whole plant	Furocoumarins in leaf: e.g. psoralen (3.2-10.5%), bergapten (6.4-14.7%), 8-methoxyoxoralan (0.53-5.3%), isopimpinellin (1.6-8.0%), Parley leaf oil: phenylpropanoids: e.g. myristicin (1.5-14%), apiole (0.9-8.1%) Common parsley seed oil: phenylpropanoids: e.g. myristicin (2.4-37%), elemicin (8.8%), apiole (1.1-67%) Italian parsley seed oil: phenylpropanoids: e.g. myristicin (0.7-40%), elemicin (0.2%), apiole (30-68%) Curly parsley seed oil: phenylpropanoids: myristicin (45-82%), elemicin (0-12.2%), apiole (0-7.2%)	Fruit has been used to induce abortion.	PDR for Herbal Medicines. 2004 Thomson ed. ISBN: 1-56933-5126-7 Natural Sources of Flavonoids Report No. 2. 2007. Ed. Council of Europe Publishing. ISBN 978-92-871-4516-7
<i>Peunia violacea</i> Lindl.	Solanaceae	Unspecified		Reported hallucinogenic properties. Compounds not defined	Butler EG. Et al. 1981. <i>Peunia violacea</i> , hallucinogen or not? <i>J. Ethnopharmacol</i> . 4(1):111-114. Scribnikov Z. A. et al. 2003. Stritulin: An antinociceptive coumarin from the roots of <i>Peucedanum ostruthium</i> . <i>Phanta Med</i> . 69(4). 369-71. Council of Europe. 2008. Natural sources of flavonoids. Report No. 3. Council of Europe Publishing. ISBN 978-92-871-8422-3
<i>Peucedanum ostruthium</i> (L.) W.Koch (Umbelliferae)	Apiaceae (Umbelliferae)	Whole plant	Furocoumarins in root: e.g. psocadin, imperatorin, oxypeucedanin		Bruneton J. 2009. <i>Pharmacognosie. (Phytochimie, Plantes médicinales)</i> . Ed. Tec & Doc, Lavoisier, Paris, 4ème édition. ISBN: 978-2-7430-1188-8 del Vallea J.M. et al. 2005. Extraction of boldo (<i>Peumus boldus</i> M.) leaves with supercritical CO ₂ and hot pressurized water. <i>Food Res Int</i> . 38(2). 203-213. Council of Europe. 2005. Active principles (constituents of chemical concern) contained in natural sources of flavonoids. Ed. Council of Europe Publishing. http://www.coe.int/t/09/active_principles/active_20principles.pdf Sparvoli F., et al. 2001. Lectin and lectin-related proteins in <i>Lima bean</i> (<i>Phaseolus lunatus</i> L.) seeds: biochemical and evolutionary studies. <i>Plant Molecular Biology</i> 45: 587-597.
<i>Peumus boldus</i> Molina	Moniiniaceae	Leaf	Isocoumarin alkaloids: e.g. boldine, ... Essential oil: phenylpropanoids: e.g. methyl Eugenol (1.19%)		Council of Europe. 2005. Active principles (constituents of chemical concern) contained in natural sources of flavonoids. Ed. Council of Europe Publishing. http://www.coe.int/t/09/active_principles/active_20principles.pdf Sparvoli F., et al. 2001. Lectin and lectin-related proteins in <i>Lima bean</i> (<i>Phaseolus lunatus</i> L.) seeds: biochemical and evolutionary studies. <i>Plant Molecular Biology</i> 45: 587-597.
<i>Phaseolus lunatus</i> L.	Leguminosae (Fabaceae)	Seed	Cyanogenic glycoside: licanarin (100 to 3000 mg HCN/kg of seed). Lectins		Chen M.L. et al. 2010. Chemical and biological differentiation of <i>Cortex Pteleodendri</i> <i>Chionensis</i> and <i>Cortex Pteleodendri Annurensis</i> . <i>Phanta Med</i> . 78(14):1530-5
<i>Phaseolus vulgaris</i> L.	Leguminosae (Fabaceae)	Seed	Cyanogenic glycoside: licanarin (20 mg/kg). Lectins		Petersen D.D. 2011. <i>Common plant toxicology: a comparison of national and southwest Ohio data trends on plant poisonings in the 21st century</i> . Toxcol Appl Pharmacol. 254(2):148-53.
<i>Pteleodendron amurense</i> Rupr.	Rutaceae	Bark	Isocoumarin alkaloids: e.g. berberine (major alkaloid, up to 8%), palmatine		
<i>Pteleodendron</i> spp.	Araceae	Whole plant	Genus in which species may contain oxalate raphides		

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Botanical name	Family	parts of plants of possible concern	Chemical of concern	Remarks on toxic/adverse effects(s) not known to be related to the identified chemical(s) of concern	Specific References
<i>Physalis alkekengi</i> L.	Solanaceae	Fruit, root	Tropane alkaloids in root (0.09-0.1%); e.g. 3-alpha-lygoyoxytropine, phytamine, cuscohygrine	Anti-estrogen activity of fruit.	Bassey K. and Woolley J.G.: 1973. Alkaloids of <i>Physalis alkekengi</i> . <i>Phytochem Rep</i> 12, 2557-2559. Bassey K. et al.: 1992. Phytamine, an alkaloid from <i>Physalis alkekengi</i> species. <i>Phytochem</i> , 31: 4173-4176. Montasserri A. et al.: 2007. Anti-fertility effects of <i>Physalis alkekengi</i> fruit on estrus cycle. <i>female rat</i> . <i>Iranian J. Reprod Med</i> 5: 13-16. Vessal M. et al.: 1991. Effects of an aqueous extract of <i>Physalis alkekengi</i> fruit on estrus cycle, reproduction and uterine creatine kinase BB-isoenzyme in rats. <i>J Ethnopharmacol</i> 34: 69-78.
<i>Physostigma venenosum</i> Bail.	Leguminosae (Fabaceae)	Seed	Indole alkaloids, e.g. physostigmine (eserine)		Burtonel J.: 2009. Pharmacognosie. (Phytochimie, Plantes médicinales). Ed. Tec & Doc. Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1188-8
<i>Phytolacca</i> spp.	Phytolaccaceae	Root, seed	Genus in which species may contain triterpenoid saponins: e.g. phytolaccatoxin and mitogenic lectins		Cooper M.R. and Johnson A.W.: 1998. Poisonous plants and fungi in Britain. <i>Animal and Human Poisoning: The Stationery Office</i> , ISBN 0-11-242981-5. Frohe D, Plander HJ and Anton R.: 2009. <i>Plantes à risques</i> . Ed. Tec et Doc-Lavoisier, ISBN: 978-2-7430-0907-1
<i>Picramnia antidemsa</i> Sw.	Picramniaceae	Unspecified	Anthraquinone derivatives: e.g. aloë emodin, aloë emodin anthrone and substituted hydroxyanthraquinones: e.g. picramninoside A, B, C		Hernandez-Medel M.R., Pereda-Miranda R.: 2002. Cytotoxic anthraquinone derivatives from <i>Picramnia antidemsa</i> . <i>Planta Med</i> 68(6):556-8.
<i>Pteris formosa</i> (Wall.) D. Don.	Ericaceae	Whole plant	Diterpenoids: gyanarotoxins		Hollands R.D., et al.: 1986. P. tomosorum poisoning in the goat. <i>Ver. Rec.</i> 118 (14): 407-408.
<i>Pteris japonica</i> (Thunb.) D. Don ex G. Don.	Ericaceae	Whole plant	Diterpenoids: gyanarotoxins		Zhang E.L., et al.: 2001. Study on the mechanism of action of <i>P. tomosorum</i> . <i>Ind. Vet. J.</i> 78 (12): 1098-1101.
<i>Plicocarpus</i> spp.	Rubiaceae	Whole plant	Genus in which species may contain indazole alkaloids, e.g. plicocarpine, plicocarpidine, plicisine,...	<i>Plicocarpus laborandi</i> Holmes known for its high plicocarpine content	Cooper M.R. and Johnson A.W.: 1998. Poisonous plants and fungi in Britain. <i>Animal and Human Poisoning: The Stationery Office</i> , ISBN 0-11-242981-5. Burtonel J.: 2005. <i>Plantes toxiques (Végétaux dangereux pour l'homme et les animaux)</i> . Ed. Tec & Doc, Lavoisier, Paris, 4ème édition, 618 pages, ISBN: 2-7430-0806-7
<i>Pimenta racemosa</i> (Mill.) J.W. Moore	Myrtaceae	Leaf	Essential oil: phenylpropanoids: methylchavicol (30-10.745 ppm), methyl Eugenol (4.31-14.65 ppm)		EMEA/HMPWP/339/03/2004. Final position paper on the use of herbal medicinal products containing estragole. Available at: http://www.emea.europa.eu/docs_en_GB/document_library/Position_statement/2009/1/2/WC500018033.pdf
<i>Pimpinella anisum</i> L.	Apiaceae (Umbelliferae)	Seed	Furocoumarins in traces. Essential oil: phenylpropanoids e.g. methylchavicol (1-5%)		Council of Europe: 2007. Natural sources of Flavonoids. Report No. 2. Council of Europe Publishing. ISBN 978-92-871-6156-7
<i>Pimpinella major</i> (L.) Huds.	Apiaceae (Umbelliferae)	Root	Furocoumarins: e.g. pimpinellin, sponidin		Council of Europe: 2008. Natural sources of Flavonoids. Report No. 3. Council of Europe Publishing. ISBN 978-92-871-6422-3
<i>Pimpinella saxifraga</i> L.	Apiaceae (Umbelliferae)	Whole plant	Furocoumarins in root (0.025%); e.g. angelicin, pimpinellin, sponidin, imperatorin, bergapten, isobergapten, isopimpinellin, pterocedarin, scopoletin, umbelliferon, umbelliprenin, xanthoxon		Han M.H., Yang X.W.: 2006. Phytochemical Study of the Rhizome of <i>Pimpinella tenuis</i> and Quantification of Phenylpropanoids in commercial <i>Pimpinella</i> Tubers by RP-LC. <i>J Chrom.</i> 64: 11-12
<i>Pimpinella turana</i> (Thunb.) Benthob. (P. tuberculata Ten.)	Apiaceae	Whole plant	Phenethylamine: L-ephedrine (0.0072% in tuber)		de Almeida R.R. et al.: 2009. Chemical variation in <i>Piper aduncum</i> and biological properties of its dihydropiperidine essential oil. <i>Chem Biodiv.</i> 6: 1427-1434.
<i>Piper aduncum</i> L.	Piperaceae	Aerial part	Essential oil: phenylpropanoids: e.g. dihydropiperidine (35-90%)	Formerly used as an abortifacient	Ralli et al.: 2007. Volatile chemical constituents of <i>Piper aduncum</i> L. and <i>Piper gubbilimum</i> C. DC (Piperaceae) from Papua New Guinea. <i>Molecules</i> , 12(3):389-94.
<i>Piper betle</i> L.	Piperaceae	Whole plant	Essential oil from the leaf (8%); phenylpropanoids: methylchavicol (1.02-4.0%), methyl Eugenol (4.1%)		Prakash B. et al.: 2010. Efficacy of chemically characterized <i>Piper betle</i> L. essential oil against fungal and aflatoxin contamination of some edible commodities and its antioxidant activity. <i>Int. J. Food Microbiol.</i> 142(1-2): 14-19
<i>Piper hispidum</i> Swingle (Piper asperitum Ric., Piper asperitulum Ruiz & Pav.)	Piperaceae	Leaf and stem	Butenolides: e.g. 9, 10-methyleneoxy-5,6-Z-hydroxyenolide, 5,6-Z-hydroxyenolide, piperolide	Estrogen agonistic effects reported for leaf extracts	Chakraborty J.B. et al.: 2011. Hydroxychavicol, a <i>Piper betle</i> leaf component, induces apoptosis of CML cells through mitochondrial reactive oxygen species-dependent JNK and endoplasmic reticulum stress activation and overrules intrinsic resistance. <i>Cancer Sci.</i> Epub.
<i>Piper methysicum</i> G. Forst.	Piperaceae	Whole plant			EMEA/HMPWP/339/03/2004. Final position paper on the use of herbal medicinal products containing estragole. Available at: http://www.emea.europa.eu/docs_en_GB/document_library/Position_statement/2009/1/2/WC500018033.pdf
<i>Piperadenia peregrina</i> Berth. (Piper adenitum Ric.)	Leguminosae (Fabaceae)	Root	Rotenoids: e.g. rotenone, miltetone, isomiltetone		Michel J.L. et al.: 2010. Estrogenic and serotonergic butenolides from the leaves of <i>Piper hispidum</i> Swingle. (Piperaceae). <i>J Ethnopharmacol.</i> 129: 220-226.
<i>Piscidia piscipola</i> (L.) Sarg.	Leguminosae (Fabaceae)	Root			Neturkar P.V. et al.: 2004. In vitro toxicity of kava alkaloids, pipermethyline, in HepG2 cells compared to xavacolones. <i>Toxicology Science</i> 79 (1): 106-111.
<i>Plochea sagittalis</i> (Lam.) Calverea	Compositae (Asteraceae)	Aerial part	Essential oil: monoterpane etheroxide: 1,8-cineole and bicyclic monoterpenes: e.g. camphor. Reported to contain methyl Eugenol in unspecified quantities.	Hepatotoxicity has been reported.	PDR for Herbal Medicines: 2004. Thomson ed. ISBN: 1-56383-5125-7

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<i>Podophyllum</i> spp.	Berberidaceae	Rhizome	Genus in which species may contain the resin podophyllin (3-6%) composed of cyclooligomers, e.g. podophylotoxin (20%), alpha and beta peltains and their derivatives.		Burtoner J. 2009. Pharmacognosie. (Phytochimie, Plantes médicinales). Ed. Tec & Doc Lavoisier. Paris, 4ème édition. ISBN: 978-2-7430-1188-8 Sullivan P. et al. 2008. Assessment of diversity in <i>Podophyllum</i> heurandium by genetic and phytochemical markers. <i>Scientia Horticulturae</i> , 115(4): 399-408 Barnes J., Anderson A., Phillipson J.D. 2007. <i>Herbal Medicines</i> . 3rd ed. Ed. Pharmaceutical Press. ISBN 978-0-55969-623-0 Rosenstein G. et al. 1976. <i>Podophyllum - a dangerous laxative</i> . <i>Pediatrics</i> , 57: 419-421.
<i>Polygonia</i> spp.	Polygonaceae	Rhizome			Burtoner J. 2009. Pharmacognosie. (Phytochimie, Plantes médicinales). Ed. Tec & Doc Lavoisier. Paris, 4ème édition. ISBN: 978-2-7430-1188-8 Barnes J., Anderson A., Phillipson J.D. 2007. <i>Herbal Medicines</i> . 3rd ed. Ed. Pharmaceutical Press. ISBN 978-0-55969-623-0
<i>Polygonatum</i> spp.	Asparagaceae	Whole plant			Frome D., Pfander H.J. and Anton R. 2009. <i>Plantes à risques</i> . Ed. Tec et Doc Lavoisier, ISBN: 978-2-7430-0907-1
<i>Polygonum multiflorum</i> Thunb.	Polygonaceae	Root	Anthraquinones : e.g. emodin, chrysophanol		Australian Government/CMC/58. Complementary Medicines Evaluation Committee. Extracted Rawlfed Minutes. Fifty-eighth Meeting. 18 August 2006 Yu J. et al. 2011. Hepatotoxicity of major constituents and extractions of <i>Radix Polygoni Multiflori</i> and <i>Radix Polygoni Multiflori</i> . <i>Pharmacoparata. J. Ethnopharmacol.</i> 137(3): 1291-9.
<i>Polygonum lilik-nas</i> L. <i>See Dryopteris lilik-nas</i> (L.) Sautt.					
<i>Poncirus trifoliata</i> (L.) Raf.	Rutaceae	Fruit	Acidone alkaloids, e.g. 5-hydroxy- <i>isoradionyne</i>		Wu T-S. et al. 1986. The first isolation of an acidone alkaloid from <i>Poncirus trifoliata</i> . <i>J Nat Prod.</i> 49(6): 1154-1155.
<i>Populus nigra</i> L.	Salicaceae	Bark and bud		Buds: benzoylsalicylic acid, salicylalcohol glycoside: salicin (2.4%), salicorin and their benzoyle derivatives, e.g. populin, tremulidin	Barnes J., Anderson A., Phillipson J.D. 2007. <i>Herbal Medicines</i> . 3rd ed. Ed. Pharmaceutical Press. ISBN 978-0-55969-623-0
<i>Populus tremula</i> L.	Salicaceae	Bark and bud		Buds: benzoylsalicylic acid, salicylalcohol glycoside: salicin (2.4%), salicorin and their benzoyle derivatives, e.g. populin, tremulidin	Barnes J., Anderson A., Phillipson J.D. 2007. <i>Herbal Medicines</i> . 3rd ed. Ed. Pharmaceutical Press. ISBN 978-0-55969-623-0
<i>Potentilla erecta</i> (L.) Haussch.	Rosaceae	Whole plant		Rich in tannins (15 to 20%). Hydrolysable tannins used at high doses over long periods may have a negative impact on liver function.	Council of Europe. 2007. <i>Neutral sources of flavonoids</i> . Report No. 2. Council of Europe Publishing. ISBN 978-92-817-8156-7. Botanical Safety Handbook. Mc Guinn M (Ed). ISBN: 0-89925-1875-8
<i>Potentilla reptans</i> L.	Rosaceae	Whole plant		Tannins from 6 to 12%. Hydrolysable tannins used at high doses over long periods may have a negative impact on liver function.	PDR for Herbal Medicines. 2004. Thomson ed. ISBN: 1-56363-5125-7
<i>Prunella vulgaris</i> L.	Lamiaceae (Labiatae)	Flowerhead		Antiestrogenic activity but compounds not identified	Collins N.H. et al. 2009. Characterization of antiestrogenic activity of the Chinese herb, <i>Prunella vulgaris</i> . <i>Using in vitro</i> and <i>in vivo</i> (Mouse Xenograft) models. <i>Biol Reprod.</i> 80(2):375-383. (<i>Uterinum in Biol Reprod.</i> 2009 Jun;80(6):1306).
<i>Prunus</i> spp.	Rosaceae	Fruit, leaf and seed	Genus in which species may contain cyanogenic glycosides, e.g. amygdalin, prunasin	Teratogenic effects of <i>Prunus serotina</i> (leaves and bark) have been reported in swine	Natural sources of flavonoids (Rep No 3). Council of Europe. (2008) PDR for Herbal Medicines. 2004 Thomson ed. ISBN: 1-56363-5125-7 Frome D. and Pfander J. A Colour Atlas of Poisonous Plants. Wolfe 1984 ISBN 0723408394 Sally L.A. et al. 1971. Outbreak of swine malformation associated with the wild black cherry, <i>Prunus serotina</i> Ehrh. <i>Zool.</i> 22(4): 498-501. Fitzgerald T.D. 2008. Larvae of the fall webworm, <i>Hyalobates cunea</i> , inhibit cyanogenesis in <i>Prunus</i> . <i>J Exp Biol.</i> 211: 671-677. Zhou J. et al. 2002. Investigation of the microheterogeneity and aglycone specificity concerning residues of black cherry <i>Prunus</i> hydrolases. <i>Plant Physiol.</i> 129(3): 1253-1264
<i>Pseudocaryophyllus gullii</i> (Speg)	Myricaceae	Fruit and leaf	Phenylpropanoids, e.g. methyl Eugenol (5%)		De Frenk J.S. et al. 1972. Essential oil of <i>Pseudocaryophyllus gullii</i> . <i>Ann Acad Bras Cien.</i> 44(Suppl. 1):175-180.
<i>Psoralea</i> spp.	Leguminosae (Fabaceae)	Fruit and seed	Genus in which species may contain furanocoumarins: e.g. psoralen		Frome D., Pfander H.J. and Anton R. 2009. « <i>Plantes à risques</i> ». Ed. Tec et Doc Lavoisier. ISBN: 978-2-7430-0907-1
<i>Psychotria viridis</i> Ruiz et Pav.	Rubiaceae	Whole plant	Indole alkaloids (tryptamine derivatives) : e.g. N,N-dimethyltryptamine		Reyna Prieto V. et al. 1994. Isolation of the alkaloid N,N-dimethyltryptamine from <i>Psychotria viridis</i> R. & P. <i>Bol Soc Quim Peru.</i> 60(1): 21-23. Blackledge R. et al. 2003. <i>Psychotria viridis</i> - a botanical source of dimethyltryptamine (DMT). <i>Microgram Journal.</i> 1(1): 21-18-22.
<i>Psidium aquilinum</i> (L.) Kuhn.	Derris/daillaceae (Myrtaceae)	Whole plant	Noresquiterpene glycosides : e.g. Marquissite. Presence of thiaminase and cyanogenic glycoside: prunasin	The biotransformation of the carinogenic flavonoids gives rise to the neurotoxic marquissite B	Frome D., Pfander H.J. and Anton R. 2009. « <i>Plantes à risques</i> ». Ed. Tec et Doc Lavoisier. ISBN: 978-2-7430-0907-1
<i>Pueraria candollei</i> Benth. var. <i>mitrifida</i> (Ary Shaw & Suval.) Niyomdham (<i>Pueraria mitrifida</i> Ary Shaw & Suval.)	Leguminosae (Fabaceae)	Tuber	Isolavones: miconestrol, deoximiconestrol, daidzein, genistein ...	Extracts of <i>P. mitrifida</i> induced higher frequencies of micronuclei	Saengphet K. et al. 2005. Mutagenicity of <i>Pueraria mitrifida</i> Ary Shaw & Suvalatubaru and antimutagenicity of <i>Tridaxia laurifolia</i> Lam. <i>Southeast Asian J Trop Med Public Health.</i> 36 (Suppl 4): 238-241. Jaisornpon S. et al. 2007. Assessment of fertility and reproductive toxicity in adult female mice after long-term exposure to <i>Pueraria mitrifida</i> herb. <i>J Reprod Dev.</i> 53(5): 995-1005.
<i>Pueraria mitrifida</i> Ary Shaw & Suval. See <i>Pueraria candollei</i> Benth. var. <i>mitrifida</i> (Ary Shaw & Suval.) Niyomdham					
<i>Pulsatilla officinalis</i> L.	Borraginaceae	Root	Possible presence of pyridizidine alkaloids	Novel Food Catalogue: not authorised for food or food supplement use	Frome D., Pfander H.J. and Anton R. 2009. « <i>Plantes à risques</i> ». Ed. Tec et Doc Lavoisier. ISBN: 978-2-7430-0907-1
<i>Pulsatilla pratensis</i> Mill.	Ranunculaceae	Aerial part	Unsaturated lactone : protoanemonin	Protoanemonin only present in fresh herb	Frome D., Pfander H.J. and Anton R. 2009. « <i>Plantes à risques</i> ». Ed. Tec et Doc Lavoisier. ISBN: 978-2-7430-0907-1

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<i>Passiflora vitifera</i> Mill. (<i>Adenocaulon vulgare</i> L.)	Ranunculaceae	Aerial part	Unsaturated lactone : protoanemonin	Protoanemonin only present in fresh herb	Frohne D., Pfander H. J. and Anton R. 2009. « Planins a resques ». Ed. Tec et Doc-Lavoisier, ISBN: 978-2-7430-0907-1
<i>Punica granatum</i> L.	Lythraceae (Punicaceae)	Fruit: root cortex and tree bark	Piperidine alkaloids (0.5-0.7%) e.g. pelletierine, iso-pelletierine, methylscopelleterine and tropane alkaloids; e.g. pseudopelletierine	The fruit hydroalcoholic extract is genotoxic <i>in vitro</i> and <i>in vivo</i> .	Sanchez-Lamar A. et al. 2008. Assessment of the genotoxic risk of <i>Punica granatum</i> L. (Punicaceae) whole fruit extracts. J Ethnopharmacol. 121:115(9):416-422. Malik, A. et al 2005. Proinflammatory, Punicic granatum and its potential for chemoprevention and chemotherapeutic of prostate cancer. Proc. Natl. Acad. Sci. USA. 102: 14813-14818 Schmidt A et al. 2005. Investigation of a betaine alkaloid from <i>Punica granatum</i> . Nat Prod Res. (5), 541-546
<i>Puraria roxburghii</i> Wall.	Purariaceae	Leaf and seed	Seed: proteic trypsin inhibitor	Leaf extract of <i>Puraria roxburghii</i> at 0.5, 1.0 or 2.0 g/kg body weight/day given orally for seven days to mice induced mitosis-disruptive chromosomal changes in bone marrow cells.	Awasthy K. S. et al. 2000. Cytogenetic toxicity of leaf extract of <i>Puraria roxburghii</i> , a medicinal plant. J Toxicol Sci. 25(5):3, 177-180. Chandrapo N. S. et al. 2008. Purification and characterization of a trypsin inhibitor from <i>Puraria roxburghii</i> seeds. Ethnobotany. 69(1):1, 2120-2126
<i>Pyrola pubera</i> Michx.	Santalaceae	Fruit and seed	Polypeptides e.g. purothion, viscoxin, phoratoxin, cranbin and thionin		Russell AB et al. 1997. Purgative Plants of North Carolina. North Carolina State University. Oserio e Castro VR. et al. 2001. Binding to and hemolysis of human erythrocytes by pyrolane thionin and Naja naja kaouthia cardiotoxin: inhibition by prothrombin. J Nat Toxins.10(3). 285-288
<i>Quassia</i> spp.	Simarubaceae	Wood	Genus in which species may contain quassinoids (nortriterpenoids); e.g. quassin and/or iridoid alkaloids; e.g. beta-carboline, camphir-6-one	Reproductive toxicity in animals	Council of Europe. 2008. Natural sources of Flavonurins. Report No. 3. Council of Europe Publishing. ISBN 978-92-871-6422-3 Buretton J. 2009. Pharmacognosie, (Phytochimie, Plantes médicinales), Ed. Tec & Doc-Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1188-8 Woo G.H. et al. 2007. Promoting potential of a Jamaica quassia extract in a rat medium-term hepatocarcinogenesis bioassay. Food Chem Toxicol. 45(7), 1160-1164. Parveen S. et al. 2003. A comprehensive evaluation of the reproductive toxicity of <i>Quassia amara</i> in male rats. Reprod Toxicol. 17(1):1,45-50.
<i>Quercus</i> spp.	Fagaceae	Bark, fruit and leaf	Calcumoxalate (11%) Triterpenoid saponins (quillaja saponins)	Genus in which species may contain high levels of tannins. Hydrolysable tannins used at high doses over long periods may have a negative impact on liver function. Inhibits trypsin-like proteases and alpha-amylases. Animal intoxication in a number of different countries and with different <i>Quercus</i> species	Frome, D.S., Pfander H.J. and Anton R. 2009. Planins a resques. Lavoisier ISBN 978-2-7430-0907-1. Medvedkov A.A. and Ivanisov A.V. 1996. Seasonal dynamics of the content of proteins inhibiting trypsin-like proteases in the leaves of common oak (<i>Quercus petraea</i> Liebl.). Ukr Biokhim Zh. 18(6):6, 44-50. Zajack A. et al. 2007. Appgo tannin: structural analysis and salivary amylase inhibition. Carbohydr. Res. 342(5), 717-723 Perez V. et al. 2011. Oak leaf (<i>Quercus pyrenaica</i>) poisoning in cattle. Res. Vet. Sci. 91(2), 269-277
<i>Quillaja saponaria</i> Molina	Quillajaaceae (Rosaceae)	Bark	Calcumoxalate (11%) Triterpenoid saponins (quillaja saponins)		FAO/WHO JECFA. 2005. Quillaja extracts Type 1 and Type 2. Chemical and technical assessment 65th JECFA. Hu K. et al. 2010. Nanoparticulate <i>Quillaja</i> saponin induces apoptosis in human leukemia cell lines with a high therapeutic index. Int J Nanomedicine. 5:514-522.
<i>Ranunculus</i> spp.	Ranunculaceae	Whole plant	Genus in which species may contain an unsaturated lactone : protoanemonin	Protoanemonin only present in fresh herb. In <i>Ranunculus fernatius</i> Thunb., two new iridoid/polydioxazinolide alkaloidal glycosides. In the root of <i>Ranunculus repens</i> L., two potent inhibitors of urease activity have been identified.	Frome D., Pfander H.J. and Anton R. 2009. Planins a resques. Ed. Tec et Doc-Lavoisier, ISBN: 978-2-7430-0907-1 Zhang L. et al. 2007. Two new iridoid/polydioxazinolide alkaloidal glycosides from <i>Ranunculus fernatius</i> Chem Pharm Bull (Tokyo). 55(8):1267-1269. Khan W. N. et al. 2006. New natural urease inhibitors from <i>Ranunculus repens</i> J Enzyme Inhib Med Chem. 21(1):1,17-19.
<i>Rauvolfia</i> spp.	Apocynaceae (Rubiaceae)	Whole plant	Genus in which species may contain iridoid alkaloids; e.g. reserpine, serpentine, yohimbine, almalicine		Burton J. 2009. Pharmacognosie, (Phytochimie, Plantes médicinales), Ed. Tec & Doc-Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1188-8
<i>Rauvolfia aromatica</i> Som. (<i>Azalea</i> <i>hybridum</i> <i>aromaticum</i> Willd.)	Lauraceae	Leaf	Essential oil: phenylpropanoids; e.g. methylchavicol (79.7%), methyl Eugenol (8.5%)	The fruits of <i>Rhamnus humboldtiana</i> Willd. ex Scull. contain neurotoxic substances (mainly diester-resorcinols of some anthracene-dimeric derivatives or of anthracenes linked to a naphthalenic derivative).	Ramanoelina P. A. R. et al. 2006. Chemical composition of <i>Rauvolfia aromatica</i> Som. leaf essential oils from Madagascar. Journal of Essential Oil Research. 18 (2):215-217
<i>Rhamnus</i> spp.	Rhamnaceae	Bark and fruit	Genus in which species may contain hydroxyanthracene derivatives.		Demmelte L. and Demner K. 2010. Anthraquinones in plants. Source, safety and applications in gastrointestinal health. Nottingham University Press. ISBN: 978-1-897676-32-5
<i>Rhus</i> spp.	Polygonaceae	Whole plant	Genus in which species may contain oxalates and hydroxyanthracene derivatives		Frome D., Pfander H. J. and Anton R. 2009. « Planins a resques ». Ed. Tec et Doc-Lavoisier, ISBN: 978-2-7430-0907-1
<i>Rhododendron</i> spp.	Ethnaceae	Flower and leaf	Genus in which species may contain diterpenes: grayanotoxins; e.g. andromedotoxin		Hazards in the wet tropics N°31 November 1995. Wet Tropics Management Authority - Queensland Department of Environment and Heritage. http://www.dem.qld.gov.au/regisler/pdf/0820bc.pdf Trippelt S. 1957. Toxic constituents of the Australian finger cherry, <i>Rhodomyrtus macrocarpa</i> Benth. J. Chem. Soc. 1957: 414-419
<i>Rhodomyrtus</i> spp.	Myrtaceae	Fruit		<i>R. macrocarpa</i> berries reported to cause permanent blindness in children but may be due to turgid toxins	
<i>Rhus</i> spp.	Anacardiaceae	Aerial part	Genus in which species may contain urushols	Fresh fruits may contain high level of tannins. Hydrolysable tannins used at high doses over long periods may have a negative impact on liver function.	Frome, Pfander and Anton. 2009. Frohne D., Pfander H. J. and Anton R. 2009. « Planins a resques ». Ed. Tec et Doc-Lavoisier, ISBN: 978-2-7430-0907-1
<i>Rhynchosia</i> spp.	Leguminosae (Fabaceae)	Root			Wang J. G. et al. 2007. Comparison of the anti-fertility effects of four extracts from the roots of <i>Rhynchosia volubilis</i> Lour. <i>Zhonghua Nan Ke Xue</i> . 31(10), 871-5.
<i>Rhinus communis</i> L.	Euphorbiaceae	Seed	Toxalbumin, ricin	Impact on rats and mice: pregnancy and reproduction.	EFSA Panel on Contaminants in the Food Chain (CONTAM). 2008. Scientific opinion on ricin (from <i>Rhinus communis</i>) as undesirable substances in animal feed. The EFSA Journal. 726, 1-38

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Botanical name	Family	parts of plants of possible concern	Chemical of concern	Remarks on toxic/adverse effect(s) not known to be related to the identified chemical(s) of concern	Specific References
<i>Rhus corymbosa</i> (L.) Haller: f.	Convolvaceae	Aerial part and seed	Indole alkaloids (ysergic acid derivatives), e.g. ysergranide (ergine)		Taber W. A. and Hancock R. A. 1982. Location of ergot alkaloid and fungi in the seed of <i>Rhus corymbosa</i> (L.) Haller. "Phytotherapy". Can J Microbiol. 8. 137-143. Frohne D., Pfander H. J. et Anton R. «Plantes à risques ». Ed. Tec et Doc-Lavoisier (2009). ISBN: 978-2-7430-0907-1
<i>Rubia pseudocastanea</i> L.	Leguminosae (Fabaceae)	Whole plant	Toxalbumins: robin (1.6% in bark), phaseol		Hui A. et al. 2004. A rare ingestion of the black locust tree. J Toxicol Clin Toxicol. 42 (1):93-95.
<i>Roenertia hybrida</i> (L.) DC.	Papaveraceae	Whole plant	Beta-carboline alkaloids: e.g. roescalbine, norroescalbine, roehamine		Gozler B. and Shamma M. 1990. Four Beta Carboline Alkaloids from <i>Roenertia-hybrida</i> J. Nat Prod (Lloyd) 53: 740-3 Gozler B. 1990. Labradore a New Pentacyclic Protoporphine Alkaloid from <i>Roenertia hybrida</i> . Heterocycles (Tokyo) 31: 149-152
<i>Rovidea japonica</i> (Thunb.) Roth	Asparagaceae (Liliaceae)	Whole plant	Cardenolides: e.g. rodekin A		Masuda T. et al. 2003. Cytotoxic screening of medicinal and edible plants in Okinawa, Japan, and identification of the main toxic constituent of <i>Rovidea japonica</i> (Omeloid). Biosci Biotech Biochem 67(6):1401-1404.
<i>Rosmarinus officinalis</i> L.	Lamiaceae (Labiatae)	Aerial part	Essential oil from the herb: bicyclic monoterpenes: e.g. camphor and monoterpene etheroxide: 1,8-cineole (13 to 31%) Essential oil from the leaf: monoterpene etheroxide: 1,8-cineole (11.2-47%) and bicyclic monoterpenes: e.g. camphor (13.31%) and monocyclic monoterpene ketone: pulegone (0.98%)		Council of Europe. 2005. Active principles (constituents of chemical concern) contained in natural sources of flavonoids (Rap No. 3). Council of Europe. (2008)
<i>Rubia cordifolia</i> L.	Rubiaceae	Root	1,3-Dihydroxy-2-hydroxymethyl-9,10-anthraquinone: lucidin		Westendorf J. et al. 1998. The genotoxicity of lucidin, a natural component of <i>Rubia inctorum</i> L., and lucidinmethylether, a component of ethanolic <i>Rubia</i> extracts. Cell. Biol. Toxicol. 4(2): 225-239
<i>Rubia inctorum</i> L.	Rubiaceae	Root	1,3-Dihydroxy-2-hydroxymethyl-9,10-anthraquinone: lucidin		Rubiee inctorum radix / Krappwurzel Bartz. Nr. 162 vom 29.08.1992. Monographien der E-Kommission. Bundesanzeiger. Verlagsanstalt. Köln.
<i>Rubus idaeus</i> L.	Rosaceae	Leaf		Oral administration to rats since the start of the gestation until parturition showed an increase of the gestation length. Female offspring (F1) showed precocious puberty age and a significant proportion of their offspring (F2) were growth retarded.	Johnson J. R. et al. 2006. Effect of maternal raspberry leaf consumption in rats on pregnancy outcome and the fertility of the female offspring. Reprod. Sci. DOI: 10.1177/1053971908328223
<i>Rumex spp.</i>	Polygonaceae	Whole plant	Genus in which species may contain hydroxyanthracene derivatives and oxalates		Frohne D., Pfander H. J. and Anton R. 2009. « Plantes à risques ». Ed. Tec et Doc-Lavoisier. ISBN: 978-2-7430-0907-1
<i>Ruscus aculeatus</i> L.	Asparagaceae	Rhizome		Contains steroidal saponins: e.g. ruscogenin, neoruscogenin	Frohne, Pfander and Anton. 2009. Frohne D., Pfander H. J. and Anton R. 2009. « Plantes à risques ». Ed. Tec et Doc-Lavoisier. ISBN: 978-2-7430-0907-1
<i>Rula spp.</i>	Rutaceae	Whole plant	Genus in which species may contain furquinoline alkaloids: e.g. diaminine and furcoumarins: e.g. bergapten	Essential oil of the aerial part of <i>Rula graveolens</i> shows abortifacient properties (probably due to the presence of methylmethylketone)	Frohne D., Pfander H. J. and Anton R. 2009. « Plantes à risques ». Ed. Tec et Doc-Lavoisier. ISBN: 978-2-7430-0907-1
<i>Salacia reticulata</i> Wight	Celastraceae	Root		Adverse effect on pregnancy	Ratnasooniga W. D. et al. 2003. Adverse pregnancy outcome in rats following exposure to a <i>Salacia reticulata</i> (Celastraceae) root extract. Braz J Med Biol Res. 36(7): 931-935.
<i>Salix spp.</i>	Salicaceae	Bark, bud, inflorescence and leaf	Genus in which species contain high concentrations of tannins (up to 20%) and the amentum may contain phytoestrogens	Salicylic glycosides (salicin, salicin, saligenin, salipreoside, picein, tannin and tremulacin) in concentration from 0.04 to 12.06%. Salix intake possibly associated with Reye's syndrome. May induce or increase pre-natal jaundice	Kerastatierne P. et al. 2009. Application of high-performance liquid chromatography for the research of salicin in bark of different varieties of <i>Salix</i> . Medicina (Kaunas). 45(6):644-51 Liguiere A., et al. 2008. Reye's and Reye-like syndromes. Cell Biochem Funct. 26: 741-746 Sengupta, E., Minelli, E., Crescini, G. and Garzanti, S. (2007) Fitoterapia. Ed. Casa Editrice Ambrosiana. ISBN: 978-8808-19269-1
<i>Salvia divinorum</i> Epling et Jativa	Lamiaceae (Labiatae)	Whole plant	Neoclerodane diterpene: e.g. salivonin A		Brau R. Jr. et al. 2008. Opioid receptors and legal highs: <i>Salvia divinorum</i> and Kratom. Chin Toxicol (Phila)49(12): 146-52. Gundramm O. et al. 2007. <i>Salvia divinorum</i> and salivonin A: an update on pharmacology and analytical methodology. Phanta Med. 73 (10): 1039-46. (Eralum in Phanta Med. 2007. 73(10): 1139).
<i>Salvia frutescens</i> Mill.	Lamiaceae (Labiatae)	Leaf		Ingestion of aqueous and ethanolic extracts of <i>Salvia frutescens</i> leaves by female rat and male rats (between 200 and 500 mg/kg) resulted in adverse effects on fertility of male and female rats	Ebeltoña, A. et al. 1998. Reproductive toxicity potentials of <i>Salvia frutescens</i> (Labiatae) in rats. J. Ethnopharmacol. 61: 67-74.
<i>Salvia lavandulifolia</i> Vahl (<i>Salvia officinalis</i> ssp. <i>lavandulifolia</i> (Vahl) Gams)	Lamiaceae (Labiatae)	Aerial part	Essential oil: monoterpene etheroxide: 1,8-cineole (11.8- 41.2%) and bicyclic monoterpenes: e.g. camphor (10-39%)		Council of Europe. 2007. Natural sources of Flavonoids. Report No. 2. Council of Europe Publishing. ISBN: 978-92-871-6156-7
<i>Salvia officinalis</i> L.	Lamiaceae (Labiatae)	Aerial part	Essential oil from leaf: bicyclic monoterpenes: e.g. alpha-thuyone (12-65%), beta-thuyone (1.2-35.6%) (total thuyone content 30-60%), camphor (4.4-30%) and monoterpene etheroxide: 1,8-cineole (8-22.5%). phenylpropanoids: e.g. methylchavicol		Council of Europe. 2008. Natural sources of Flavonoids. Report No. 3. Council of Europe Publishing. ISBN: 978-92-871-6622-3 Reid et al. 2007. The essential oil of <i>Salvia officinalis</i> L. from various European countries. J. Ess. Oil Res. 21(6):108-111.
<i>Salvia sclarea</i> L.	Lamiaceae (Labiatae)	Aerial part	Essential oil from the herb: monoterpene etheroxide: 1,8-cineole (3.23%) and bicyclic monoterpenes: e.g. camphor (1%) Essential oil from the flower: 1,8-cineole (traces), camphor.		Council of Europe. 2007. Natural sources of Flavonoids. Report No. 2. Council of Europe Publishing. ISBN: 978-92-871-6156-7 Kumar L. et al. 2009. Chemical composition and biological activities of essential oil from <i>Salvia sclarea</i> Benth. (Labiatae) in vitro. Medicines. 14: 1338-1447
<i>Sambucus canadensis</i> L.	Adoxaceae (Caprifoliaceae)	Whole plant	Possible presence of cyanogenic glycosides (S)-sambunigin	Branches, unripe berries or seeds of a number of different <i>Sambucus</i> species are characterized by the presence of substances able to induce gastrointestinal disorders	Baumstuster R. A. et al. 2000. Sambunigin and cyanogenic variability in populations of <i>Sambucus canadensis</i> L. (Caprifoliaceae). Biochim Syst Ecol. 28(7): 689-695. Frohne D., Pfander H. J. and Anton R. 2009. « Plantes à risques ». Ed. Tec et Doc-Lavoisier. ISBN: 978-2-7430-0907-1
<i>Sambucus obulus</i> L.	Adoxaceae (Caprifoliaceae)	Whole plant	Cyanogenic glycoside: S-sambunigin	Ethyliacate extract showing high toxicity in mice Presence of lectins in branches. Branches, unripe berries or seeds of a number of different <i>Sambucus</i> species are characterized by the presence of substances able to induce gastrointestinal disorders	Chaves L. et al. 1998. Presence of polymeric and free forms of the non-toxic type 2 ribosome-inactivating protein abulin and a structurally related non-homodimeric lectin in fruits of <i>Sambucus obulus</i> L. Planta. 204 (3): 310-319. Ehrlichmann M. A. et al. 2007. Separation of active and toxic portions in <i>Sambucus obulus</i> . Pak J Biol Sci. 10(22):1471-1473.

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<i>Sambucus nigra</i> L.	Adoxaceae (Caprifoliaceae)	Whole plant	Cyanogenic glycoside: S-sambunigin (3 to 17 mg HCN /100 g fresh weight in leaf and 3 mg HCN / 100g of fruit)	Presence of lectins in branches. Branches, unripe berries or seeds of a number of different <i>Sambucus</i> species are characterised by the presence of substances able to induce gastrointestinal disorders	Sangiorgi, E., Minelli, E., Crescini, G. and Garzanti, S. (2007) Fitoterapia, (ed. Casa Editrice Ambrosiana), ISBN: 978-88-808-18266-1 Frohe D., Pfander H. J. and Anton R. 2009. « Pantès à risques ». Ed. Tec et Doc-Lavoisier, ISBN: 978-2-7430-0907-1 EMEA/HMPC. 2007. <i>Sambucus nigra</i> L., (toxicity assessment) report for the development of community monographs and for inclusion of herbal substances(s), preparation(s) or combinations thereof in the list. EMEA/HMPC/253170/2007/Corr.
<i>Sanguinaria canadensis</i> L.	Papaveraceae	Rhizome and root	Benzylisoquinoline alkaloids (protoberberines): e.g. sanguinarine, cheberaline, berbamine, protopine		Frohe D., Pfander H. J. et Anton R. « Pantès à risques ». Ed. Tec et Doc-Lavoisier (2009). ISBN: 978-2-7430-0907-1
<i>Sanicula europaea</i> L.	Apiaceae (Umbelliferae)	Leaf	Tripenoid saponins: sancicloside R-1, sancicloside N	Tripenoid saponins: sancicloside A and B (major) and others. These saponins are also called saponolins as they are among the most irritating saponins.	Schioppe T. et al. 1998. Saponoside R-1: a new tripenoid saponin from <i>Sanicula europaea</i> . <i>Planta Med</i> 64(1):83-85. Arda et al. 1997. Sancicloside N from <i>Sanicula europaea</i> L. <i>J. Nat. Prod</i> 60(11):1170-1173. Mimaki Y. et al. 1996. Steroidal saponins from <i>Sansevieria trifasciata</i> . <i>Phytochemistry</i> 43(6): 1325-1331
<i>Sansevieria</i> spp.	Asparagaceae (Agavaceae)	Leaf	Genus in which species may contain steroidal saponins		Gribes T. and Ferreres JM. 2004. Description, distribution, activity and phytochemical relationship of niosome inactivating proteins in plants, fungi and bacteria. <i>Mol. Cell. Biochem.</i> 264: 45-51. Flavel D. J. 1999. Saponin immunotoxins. <i>Curr. Top. Microbiol. Immunol.</i> 234: 57-61. Kohle K. et al. 1999. New tripenoid saponins and saponolins from <i>Saponaria officinalis</i> . <i>J. Nat. Prod</i> 62(12): 1655-1659 Teuchter E., Anton R. et Losten A. « Pantès à risques ». Ed. Tec et Doc-Lavoisier (2005). ISBN: 2-7430-0720-6 Council of Europe. 2008. Natural sources of flavonoids. Report No. 3. Council of Europe Publishing. ISBN: 978-92-871-6422-3
<i>Saponaria officinalis</i> L.	Caryophyllaceae	Whole plant	Genus in which species may contain in their essential oil phenylpropanoids: e.g. safrole, isosafrole, methyl Eugenol		Council of Europe. 2008. Natural sources of flavonoids. Report No. 3. Council of Europe Publishing. ISBN: 978-92-871-6422-3
<i>Sassafras</i> spp.	Lauraceae	Whole plant	Genus in which species may contain in their essential oil phenylpropanoids: e.g. safrole, isosafrole, methyl Eugenol		Kao C.H. et al. 1999. Using 99mTc-DTPA radionuclide inhalation lung scintigraphy to detect the lung injury induced by consuming <i>Sassafras androgynus</i> vegetable and comparison with conventional pulmonary function tests. <i>Respiration</i> , 66: 48-51
<i>Saururus androgynus</i> (L.) Merr.	Phyllanthaceae	Leaf	Benzylisoquinoline alkaloids: e.g. papaverine (0.5%)	In Taiwan, lung problems seen with high intake of leaves.	PDR for Herbal Medicines. 2004 Thomson ed. ISBN: 1-56363-5125-7 Riazuddin S. et al. 1997. Mutagenicity testing of some medicinal herbs. <i>Environ. Mol. Mutagen.</i> 10(2): 141-148
<i>Saussurea</i> spp.	Compositae (Asteraceae)	Whole plant	Genus in which species may contain calcium oxalate raphides	<i>Saussurea lappa</i> reported to be mutagenic (Ames test)	Frohe D., Pfander H. J. et Anton R. « Pantès à risques ». Ed. Tec et Doc-Lavoisier (2009). ISBN: 978-2-7430-0907-1
<i>Scutellum</i> spp.	See <i>Mesembryanthemum</i> spp.				
<i>Scorifera</i> spp.	Araliaceae	Aerial part	Genus in which species may contain calcium oxalate raphides		Nelson L. S., Shin R. D. and Bailk M. J. 2007. Handbook of poisonous and injurious plants. 2nd edition. Springer. ISBN: 0-387-31268-4
<i>Scorinus teretifolius</i> Raddi	Anacardiaceae	Bark and stem	Genus in which species may contain oxalate raphides and inflammatory protein derivatives	Stem bark decoction showing mutagenic properties (Ames test)	Nelson L. S., Shin R. D. and Bailk M. J. 2007. Handbook of poisonous and injurious plants. 2nd edition. Springer. ISBN: 0-387-31268-4
<i>Scorocaulon officinale</i> Gray (<i>Schizalia officinum</i> Brandt et Garzani)	Malvaceae	Seed	Steroidal alkaloids: e.g. veratrine (mixture of cevadine, veratridine)		Nelson L. S., Shin R. D. and Bailk M. J. 2007. Handbook of poisonous and injurious plants. 2nd edition. Springer. ISBN: 0-387-31268-4
<i>Scodopisus</i> spp.	Araceae	Aerial part	Genus in which species may contain oxalate raphides and inflammatory protein derivatives		Nelson L. S., Shin R. D. and Bailk M. J. 2007. Handbook of poisonous and injurious plants. 2nd edition. Springer. ISBN: 0-387-31268-4
<i>Scopolia</i> spp.	Solanaceae	Whole plant	Genus in which species may contain tropane alkaloids: e.g. hyoscyamine, atropine, scopolamine, and tetrahydro- and tropane alkaloids: calystegines	The fresh plant contains hyoscyamine, twice more active than atropine (racemic mixture)	Bruneau J. 2005. <i>Pantès à risques (Végétaux dangereux pour l'homme et les animaux)</i> . Ed. Tec et Doc-Lavoisier, Paris, 8ème édition, ISBN: 2-7430-0806-7 Cheng, S.W. et al. 2002. Anticholinergic Poisoning from a large dose of <i>Scopolia</i> Extract. <i>Vel. Hum. Tokco</i> , 44: 222-223 Asano N. et al. 1996. Calystegine B4, a novel teretrolase inhibitor from <i>Scopolia japonica</i> . <i>Carbohydr. Res</i> 293(2):195-204
<i>Scutellaria baicalensis</i> Georgi	Lamiaceae	Leaf and stem	Genus in which species may contain oxalate raphides and inflammatory protein derivatives	O-methylated flavone: wogonin; long term administration of 120 mg of wogonin per kg to rats resulted in heart injury	Qi et al. 2009. Toxicological studies of wogonin in experimental animals. <i>Phytother. Res.</i> 23(3): 417-422
<i>Sedum acre</i> L.	Crossulaceae	Flower and leaf	Alpha substituted piperidine alkaloids: e.g. sedacrine		Frohe D., Pfander H. J. et Anton R. 2009. « Pantès à risques ». Ed. Tec et Doc-Lavoisier, ISBN: 978-2-7430-0907-1
<i>Semecarpus anacardium</i> L.f.	Anacardiaceae	Fruit	Phenolic acids: e.g. anacardic acid, urushiol III		Kesava Rao K. V. et al. 1979. Toxicological study of <i>Semecarpus anacardium</i> nut extract. <i>Indian J. Physiol. Pharmacol.</i> 23(2):115-120 International Agency for Research in Cancer (IARC)(2002) <i>Semecarpus</i> species and rhotifoline. Bruneau J. 2005. <i>Pantès à risques (Végétaux dangereux pour l'homme et les animaux)</i> . Ed. Tec et Doc-Lavoisier, Paris, 8ème édition, ISBN: 2-7430-0806-7
<i>Senecio</i> spp.	Compositae (Asteraceae)	Whole plant	Genus in which species may contain unsaturated pyridizidine alkaloids: e.g. senecionine, riddelline		Sangiorgi, E., Minelli, E., Crescini, G. and Garzanti, S. (2007) <i>Fitoterapia</i> , (ed. Casa Editrice Ambrosiana, ISBN: 978-88-808-18266-1 Frenzilli F. (2009) <i>Fitoterapia Quarta Edizione</i> . (editore Elsevier S.r.l. Milano) ISBN 978-88-214-5981-1. P. pages 346-352. Teddlid J. Macdonald R., Ruklis I., Wilk T.J. <i>Senecio</i> repens for benign prostatic hyperplasia. <i>Cochrane Database of Systematic Reviews</i> 2009, Issue 2. Art. No.: CD001423. DOI: 10.1002/14651858.CD001423.pub2
<i>Senecio repens</i> (N. Bartram) Small	Asteraceae	Fruit	Genus in which species may contain toxic amino acids: e.g. L-canavanine	Lipid-sterolic fraction: reported anti-androgenic and anti-estrogenic activity.	Frohe D., Pfander H. J. et Anton R. « Pantès à risques ». Ed. Tec et Doc-Lavoisier (2009). ISBN: 978-2-7430-0907-1
<i>Sebania</i> spp.	Leguminosae (Fabaceae)	Whole plant	Genus in which species may contain toxic amino acids: e.g. L-canavanine	Sebanamide A (glutamide derivative) causing diarrhea and CNS depression	Khanlou S. et al. 2005. HPTC method for chemical standardization of <i>Stida</i> species and estimation of the alkaloid ephedrine. <i>Journal of Planar Chromatography</i> 18: 364-367
<i>Stida acuta</i> Burm. f.	Maliaceae	Whole plant	Phenylethylnamines: e.g. ephedrine (0.006% in dried root, 0.04% in aerial part)		

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<i>Sida cordifolia</i> L.	Malvaceae	Whole plant	Polyphydroxyzolidine alkaloids and derivatives, e.g. swainsonine Indolizidine alkaloids, e.g. cyclopamine Phenylethylamines, e.g. ephedrine (0.007% in dried root, 0.112% in aerial part) and pseudoephedrine		Matsui T.A. et al. 2007. The plant alkaloid cryptolepine induces p21WAF1/CIP1 and cell cycle arrest in a human osteosarcoma cell line. <i>Int. J. Oncol.</i> 31(4):915-922. Marchel E. et al. 2006. A rapid and simple procedure for the determination of ephedrine alkaloids in dietary supplements by gas chromatography-mass spectrometry. <i>J. Pharm Biomed.</i> 41(5):1633-1641. Khatoun S. et al. 2005. HPLC method for chemical standardization of <i>Sida</i> species and estimation of the alkaloid ephedrine. <i>Journal of Planar Chromatography</i> 18: 364-367.
<i>Sida rhombifolia</i> L.	Malvaceae	Whole plant	Phenethylamines, e.g. ephedrine (0.031% in dried root, 0.017% in aerial part), quinoxalines and carboxylated tryptamines		Matsui T.A. et al. 2007. The plant alkaloid cryptolepine induces p21WAF1/CIP1 and cell cycle arrest in a human osteosarcoma cell line. <i>Int. J. Oncol.</i> 31(4):915-922. Marchel E. et al. 2006. A rapid and simple procedure for the determination of ephedrine alkaloids in dietary supplements by gas chromatography-mass spectrometry. <i>J. Pharm Biomed.</i> 41(5):1633-1641. Prakash A. et al. 1981. Alkaloid constituent of <i>Sida acuta</i> . <i>S. humilis</i> , <i>S. rhombifolia</i> and <i>S. spinosa</i> . <i>Planta Med.</i> 43(4): 384-388 Khatoun S. et al. 2005. HPLC method for chemical standardization of <i>Sida</i> species and estimation of the alkaloid ephedrine. <i>Journal of Planar Chromatography</i> 18: 364-367.
<i>Sinomenium acutum</i> (Thunb.) Rehdler & E.H. Wilson	Menispermaceae	Whole plant	Isquinoline alkaloids (morphinanes), sinomenine	Convulsive central excitation at high doses in animals. Presence of steroidal saponins, e.g. oenillin G, asparagostole E, asparagostole B that are poorly absorbed	Yamashiki H. 1976. Pharmacology of sinomenine, an anti-rheumatic alkaloid from <i>Sinomenium acutum</i> . <i>Acta Med. Okayama.</i> 30(1), 1-20. Belhoche Z. et al. 2008. Steroidal saponins from the roots of <i>Sinlix aspera</i> subsp. <i>malabarica</i> . <i>Chem. Pharm. Bull. (Tokyo)</i> . 56(9): 1324-1327
<i>Sinlix aspera</i> L.	Smilacaceae	Root			British Herbal Pharmacopoeia, Edition 1993
<i>Smilax officinalis</i> Kunth (<i>Smilax tordill</i> Aq., <i>Smilax varilliflora</i> Aq.)	Smilacaceae (Liliaceae)	Root		Steroidal saponins: sarsasapoin parvalli, sarsasapogenin, neoglygenin that are poorly absorbed	Burton J. 2009. Pharmacognosy. (Phytochemistry, Plantae medicinales). Ed. Tec & Doc Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1888-8 Berardo R. R. et al. 1996. Steroidal saponins from <i>Smilax officinalis</i> . <i>Phytochemistry</i> , 43(2), 465-469
<i>Solandra</i> spp.	Solanaceae	Whole plant	Genus in which species may contain tropane alkaloids, e.g. L-lyscovamine, scopolamine	The fresh plant contains hyoscyamine, twice more active than atropine (racemic mixture)	Wang Ch. 2006. Medicinal plants of the Asia-Pacific. World Scientific Publishing Co. Pte. Ltd. ISBN 981-256-341-5
<i>Solanum</i> spp.	Solanaceae	Whole plant	Genus in which species may contain glycosidic steroidal alkaloids, e.g. solandrine, tomatidine, ...		Keeler R. F. et al. 1980. Strychnosamine-containing <i>Solanum</i> species and induction of congenital cardiac malformations. <i>Toxicol.</i> 28(6), 873-874. Frome D. Pfander, HJ and Anton R. 2009. Plantae à l'usage. Ed. Tec & Doc-Lavoisier, ISBN: 978-2-7430-0907-1
<i>Solenostemma nigel</i> (Dallé) Hayne	Apocynaceae	Leaf and stem	Leaf: pregrane ester glycosides; e.g. stemmosides a & b	Latex known to induce purgative effect	Pérez A. et al. 2005. New unusual pregrane glycosides with antiproliferative activity from <i>Solenostemma nigel</i> . <i>Steroids</i> 70(9), 594-603 Araël L. Hamard. 2001. New steroids from <i>Solenostemma nigel</i> leaves. <i>Fitoterapia</i> . 72(7), 747-755 Hassan HA. et al. 2001. Pregnane derivatives from <i>Solenostemma nigel</i> . <i>Phytochemistry</i> . 57, 507-511
<i>Sophora japonica</i> L. (<i>Symphoricarpon japonicum</i> (L.) Schott.)	Leguminosae (Fabaceae)	Fruit and seed	Seed: quinozidine alkaloids, e.g. cystine, N-methyl cystine, matrine, sophorine	Fruit: abortifacient effect reported.	Evans WC. (2009). <i>Tease and Evans Pharmacognosy</i> . Elsevier. ISBN: 978-0-7-7020-2933-2 Hempen CH. and Fischer T. 2009. A Materia Medica for Chinese Medicine. Churchill Livingstone. Elsevier. ISBN: 978-0-443-10094-9 PDR for Herbal Medicines. 2004. Thomson ed. ISBN: 1-56363-5125-7
<i>Sophora secundiflora</i> (Ortega) Lag. ex DC.	Leguminosae (Fabaceae)	Seed	Quinozidine alkaloids, e.g. cystine (0.25%), N-methylcystine, anagyrine, epi-hupnine, delia-5-dehydrolupanine		Hatfield et al. 1977. An investigation of <i>Sophora secundiflora</i> seeds (Messebaeans). <i>Lloydia</i> 40(4), 374-383. Izadocot et al. 1976. Structure and toxicity of alkaloids and amino acids of <i>Sophora secundiflora</i> . <i>J. Pharm. Sci.</i> 65(3), 382-384.
<i>Sophora tonkinensis</i> Gagnepain	Leguminosae (Fabaceae)	Root	Quinozidine alkaloids, e.g. cystine, methylcystine, tonkinensines A and B		Ding P.L. et al. 2005. Determination of quinozidine alkaloids in <i>Sophora tonkinensis</i> by HPLC. <i>Phytochem. Analysis</i> . 16(4), 257-263
<i>Spartium junceum</i> L.	Leguminosae (Fabaceae)	Whole plant	Quinozidine alkaloids, e.g. cystine, sparteine		Burton J. 2005. <i>Plantae toxiques (Végétaux dangereux pour l'homme et les animaux)</i> . Ed. Tec & Doc. Lavoisier, Paris, 3ème édition, ISBN: 2-7430-0806-7
<i>Spathiphyllum</i> spp.	Araceae	Whole plant	Genus in which species may contain calcium oxalate raphides and proteolytic enzymes		Burton J. 2005. <i>Plantae toxiques (Végétaux dangereux pour l'homme et les animaux)</i> . Ed. Tec & Doc. Lavoisier, Paris, 3ème édition, ISBN: 2-7430-0806-7
<i>Spigelia</i> spp.	Loganiaceae	Aerial part	Genus in which species may contain acetylcholine-type monoterpene alkaloids and dieneene alkaloids (ryanodines, e.g. spigantine, spigeline)		Morais S.M. et al. 2002. Chemical investigation of <i>Spigelia anthelmia</i> , used in Brazilian folk medicine as anthelmintic. <i>Rev. Bras. Farmacogn.</i> 12, supl., 81-82. Hubner H. et al. 2001. Minor constituents of <i>Spigelia anthelmia</i> and their cardiac activities. <i>Phytochemistry</i> 57, 285-296. Achenbach H. et al. 1995. Spigantine, the cardioactive principle of <i>Spigelia anthelmia</i> . <i>J Natl Prod.</i> 58(7):1092-1096
<i>Spheckelia</i> spp.	Amaryllidaceae	Bulb	Genus in which species may contain isoquinoline alkaloids (Amaryllidaceae alkaloids), e.g. yconine, pseudoyconine, ismine		Hohmann J. et al. 2002. Antiproliferative Amaryllidaceae alkaloids isolated from the bulbs of <i>Spheckelia formosissima</i> and <i>Hybanthus vestalis</i> . <i>Planta Med.</i> 68, 454-457. Roth, D. 2000. <i>Konkurrenz: Giftpflanzen Pflanzenstoffe, Conen Verlagsgesellschaft</i> , 4th ed. 1994. ISBN 3-609-61810-4
<i>Steganotaenia araliacea</i> Hochst.	Apiaceae (Umbelliferae)	Bark and stem	Lignans, e.g. steganach, dibenzocyclooctadene lactone, 10-demethoxyselegane, steganone, prestegane B.		Agunu A. et al. 2005. Diuretic activity of the stem-bark extracts of <i>Steganotaenia araliacea</i> hook. (Apiaceae). <i>J. Ethnopharmacol.</i> 96(3), 471-475. Evans WC. (2009). <i>Tease and Evans Pharmacognosy</i> . Elsevier. ISBN: 978-0-7-7020-2933-2 Mergelhan K.M. et al. 2001. 10-Demethoxyselegane, a new lignan from <i>Steganotaenia araliacea</i> . <i>J Natl Prod.</i> 64 (11), 1480-1482.

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<i>Stephania</i> spp.	Menispermaceae	Root	Genus in which species may contain bisbenzyltetrahydroisoquinoline alkaloids: e.g. tetrandrine, fangchinoline, and/or hasubanan alkaloids, e.g. rumanine and cepharanthine.		Zhi-Da M. et al., 1995. Alkaloids of <i>Stephania sinica</i> . <i>Phytochemistry</i> , 24(12), 3084-3095. Bruneton J., 2009. Pharmacognosie. (Phytochimie, Plantes médicinales). Ed. Tec & Doc Lavoisier, Paris, 4ème édition. ISBN: 978-2-7430-1188-8 Bruneton J., 2005. Plantes toxiques (Végétaux dangereux pour l'homme et les animaux). Ed. Tec & Doc, Lavoisier, Paris, 3ème édition. ISBN: 2-7430-0806-7
<i>Sternbergia</i> spp.	Amaryllidaceae	Whole plant	Genus in which species may contain isoquinoline alkaloids (Amaryllidaceae alkaloids); e.g. lycorine, galanthamine, sternbergine, hippamine, ...		Umer N. et al., 2005. Antimicrobial activity of <i>Sternbergia sicula</i> and <i>Sternbergia lutea</i> . <i>Fitoterapia</i> 76, 226-229. Evidente A., 1986. Isolation and structural characterization of lutescine, a new alkaloid from bulbs of <i>Sternbergia lutea</i> . <i>J Nat Prod</i> , 49, 90-94. Padungoung V. et al., 1989. Four New Chinese-Type Alkaloids from <i>Sternbergia</i> Species. <i>J Nat Prod</i> , 52 (4), 785-791. Keya G. et al., 2010. HPLC - DAD analysis of lycorine in Amaryllidaceae species. <i>Nat Prod Commun</i> , 5(6), 873-6. American Herbal Products Association. 1997. Botanical Safety Handbook. Mc Guffin M (Ed). ISBN: 0-8493-1675-8
<i>Stillingia sylvatica</i> L.	Eliphorbiaceae	Root	diterpenes, cyanogenic glycosides		Fresh root: caustic latex
<i>Strobilus asper</i> (Retz.) Lour. (<i>Tropis aspera</i> Retz.)	Moraceae	Root bark	Cardiac glycosides: e.g. asperosid, strobiloside, mansoni.		Rastogi S. et al., 2006. <i>Strobilus asper</i> Lour. (Shaknotakia): a review of its chemical pharmacological and ethnomedical properties. <i>Advances Access publication</i> , 11, 216-221
<i>Strophanthus</i> spp.	Apocynaceae	Seed	Genus in which species may contain cardenolide glycosides: e.g. ouabain and ajlycones: e.g. strophanthidin		Frome D., Pfander HJ and Anton R., 2009. Plantes à risques. Ed. Tec et Doc-Lavoisier. ISBN: 978-2-7430-0907-1
<i>Strychnos</i> spp.	Loganiaceae	Fruit and seed	Genus in which species may contain indole alkaloids (e.g. strychnine) and/or bisbenzylisoquinoline alkaloids (e.g. tubocurarine)		Frome D., Pfander HJ and Anton R., 2009. Plantes à risques. Ed. Tec et Doc-Lavoisier. ISBN: 978-2-7430-0907-1
<i>Symplocobium japonicum</i> (L.) Schott. See <i>Sophora labillardieri</i> L.	Caprifoliaceae	Fruit		Reported gastrointestinal irritating effect. Substance not defined	Roth, Daurere, Komann, Giftpflanzen Pflanzenzergle. Comed Verlagsgesellschaft, 4th ed., 1994. ISBN 3-609-61810-4 Wicini M. and Anton R., 2003. Plantes thérapeutiques (Tradition, pratique officinale, science et thérapeutique). Ed. Tec & Doc, Lavoisier, Paris, 2ème édition, 692 pages. ISBN : 2-7430-0631-5
<i>Symphylum</i> spp.	Boraginaceae	Whole plant	Genus in which species may contain unsaturated pyridizidine alkaloids		Teuscher E., Anton R. et Lohstein A., 2005. Plantes aromatiques (Épices, aromatisants, condiments et huiles essentielles). Ed. Tec et Doc-Lavoisier. ISBN : 2-7430-0720-6
<i>Syzygium aromaticum</i> (L.) Merr. & L.M. Perry (<i>Caryophyllus aromaticus</i> L., <i>Eugenia caryophyllata</i> Thunb. (nom. illeg.) Marshall)	Myrtaceae	Flower bud (bove)	Essential oil: phenylpropanoids: e.g. methylchavicol (89.33%), methyl Eugenol (310-340 ppm)		Elkin N.L., 1986. Plants in indigenous medicine & diet: biobiochemical approaches. Volume 1. Routledge. ISBN: 0913178020, 9780913178027
<i>Tabeaia</i> spp.	Bignoniaceae	Bark	Genus in which species may contain naphthoquinones : e.g. lapachol and beta-lapachone		
<i>Tabeaia heptaphylla</i> (Vahl.) Toledo See <i>Handroanthus heptaphyllus</i>					
<i>Tabeaia</i> sp. (K. Schum.) Standl. See <i>Handroanthus heptaphyllus</i>					
<i>Tabeaia</i> spp.	Apocynaceae	Whole plant	Indole alkaloids: e.g. ibogaine.		
<i>Tagesis</i> spp.	Compositae (Asteraceae)	Whole plant	Genus in which the essential oil of some species may contain the phenylpropanoid methylchavicol		Bruneton J., 2009. Pharmacognosie. (Phytochimie, Plantes médicinales). Ed. Tec & Doc, Lavoisier, Paris, 4ème édition. ISBN: 978-2-7430-1188-8 Roth, Daurere, Komann, Giftpflanzen Pflanzenzergle. Comed Verlagsgesellschaft, 4th ed., 1994. ISBN 3-609-61810-4
<i>Tamus communis</i> L. See <i>Dioscorea</i> spp.					
<i>Tanaecium balsamita</i> L.	Compositae (Asteraceae)	Aerial part	Essential oil from the aerial parts at full flowering stage: monocyclic monoterpenes ketone: camone (61%), bicyclic monoterpenes: beta-thujone (20.8%), alpha-thujone (3.2%), monoderpene etheroxide: 1-8-cineole (4.4%)		Jainrand, K.; Rezaee, M.B., 2005. Chemical constituents of essential oils from <i>Tanaecium balsamita</i> L. spp. balsamitoides (Schulz-Simp.) Gertson. <i>Journal of Essential Oil Research</i> , 17(6): 565-566 Yousefzadi M. et al., 2009. Cytotoxic, antimicrobial activity and composition of essential oil from <i>Tanaecium balsamita</i> L. susp. balsamita. <i>Nat. Prod. Commun.</i> 4(1):119-122.
<i>Tanaecium chierataefolium</i> (Trevin.) Sch.Bip. (<i>Chrysanthemum chierataefolium</i> (Trevin.) Vis., <i>C. chierataefolium</i> (Trevin.) Vis., <i>Tanaecium chierataefolium</i> (Trevin.) Sch.Bip.)	Compositae (Asteraceae)	Aerial part	Leaf : monoterpenes, pyrethrins		Matsuda K. et al., 2005. Biosynthesis of pyrethrin I in seedlings of <i>Chrysanthemum chierataefolium</i> . <i>Phytochemistry</i> 60(13), 1529-1535.
<i>Tanaecium parthenium</i> (L.) Sch.Bip. (<i>Chrysanthemum parthenium</i> (L.) Benth.)	Compositae (Asteraceae)	Aerial part	Sesquiterpene lactone: parthenolide Essential oil: bicyclic monoterpenes: e.g. camphor (42.64%)		Council of Europe. 2005. Active principles (constituents of chemical concern) contained in natural sources of flavonoids. Ed. Council of Europe Publishing. http://www.coe.int/t/e/social_cooperation/doc_ssp/active_principles_active%20principles.pdf Heprinstal S. et al., 1992. Parthenolide content and bioactivity of feverfew (<i>Tanaecium parthenium</i> (L.) Schulz-Simp.). Estimation of commercial and adulterated feverfew products. <i>J. Pharm. Pharmacol.</i> 44(3):351-355. Hogopman M. et al., 1987. A study on tansy chemotypes. <i>Planta Medica</i> , 53 (3): 284-287. Council of Europe. 2005. Active principles (constituents of chemical concern) contained in natural sources of flavonoids. Ed. Council of Europe Publishing. http://www.coe.int/t/e/social_cooperation/doc_ssp/active_principles_active%20principles.pdf
<i>Taraxacum vulgare</i> L. (<i>Chrysanthemum vulgare</i> (L.) Benth.)	Compositae (Asteraceae)	Aerial part	Essential oil (0.12-0.18%); bicyclic monoterpenes: camphor (up to 90%), thujones (up to 80%) and monoderpene etheroxide: 1-8-cineole.		Bruneton J., 2009. Pharmacognosie. (Phytochimie, Plantes médicinales). Ed. Tec & Doc, Lavoisier, Paris, 4ème édition. ISBN: 978-2-7430-1188-8
<i>Taxus</i> spp.	Taxaceae	Whole plant except aril	Genus in which species may contain diterpene pseudoalkaloids (taxoids): e.g. taxine, taxol, cephalomannine		

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<i>Tephrosia</i> spp.	Leguminosae (Fabaceae)	Whole plant	Genus in which species may contain rotenoids: e.g. rotenone		Irvine JIE, and Frey RH. 1959. Source materials for rotenone, occurrence of rotenoids in some species of the genus <i>Tephrosia</i> . J. Agril. Food Chem. 7(2): 106-107. Sulman HB, et al. 1982. The toxic effects of <i>Tephrosia apollinea</i> on goats. Journal of Comparative Pathology. 92 (2): 309-315.
<i>Teucrium</i> spp.	Lamiaceae (Labiatae)	Aerial part	Genus in which species may contain turanone/diterpene dipeptides: e.g. teucriin		Mosler-Kara et al. 1992. Fatal hepatitis after herbal tea. Lancet 340: 674 Fau D, et al. 1997. Diterpenoids from germanium, an herbal medicine, induce apoptosis in isolated rat hepatocytes. Gastroenterology. 113(4): 1324-1346 Rodriguez MC, et al. 1984. Isoloflidiin, a neo-decane dipeptide from <i>Teucrium chamaedrys</i> , and revised structures of teucriin F and G. Phytochemistry 23: 1465-1469
<i>Thapsia</i> spp.	Apiaceae (Umbelliferae)	Fruit	Genus in which the essential oil of some species may contain the phenylpropanoid methyl Eugenol		EMEA-HMPC. 2005. Public statement on the use of herbal medicinal products containing methyl Eugenol. EMEA/HMPC/138362/2005. Avato P, et al. 1998. Effect of <i>Thapsia</i> Essential Oils on Bile Composition in Rats. Pharmaceutical Biology (Formerly International Journal of Pharmacognosy). 36 (5): 335-340. Avato P, et al. 1996. Essential oils from fruits of three types of <i>Thapsia villosa</i> . Phytochemistry. 43(3): 609-612.
<i>Thermopsis lanceolata</i> R.Br.	Leguminosae (Fabaceae)	Flower and seed	Quinolizidine alkaloids: e.g. cystine, thermopsis, anagyrene.		Paaner KE, and Keeler RF. 1993. Quinolizidine and piperidine alkaloid teratogens from poisonous plants and their mechanism of action in animals. Vet. Clin. North Am. Food Anim. Pract. 9(1): 33-40. Keeler RF, and Baker DC. 1990. Mopahity in cattle induced by alkaloid extracts from <i>Thermopsis montana</i> , <i>Laburnum anagyroides</i> and <i>Lupinus</i> sp. J. Comp. Pathol. 103(2): 169-182. Burgeon J. 2005. Plantes toxiques (Végétaux dangereux pour l'homme et les animaux). Ed. Tec & Doc. Lavoisier, Paris, 3ème édition. ISBN : 2-7430-0806-7 Vignatova V, et al. An investigation of the alkaloids of <i>thermopsis inosoides</i> . Chemistry of Natural Compounds. 7(4): 440-442.
<i>Theselia</i> spp.	Apocynaceae	Whole plant	Genus in which species may contain cardiac glycosides and their aglycons: e.g. thevetoside...		Roth, Daurener, Komann, Giftpflanzen Pflanzenstoffe, Cornet Verlagsgesellschaft, 4th ed. 1994. ISBN 3-609-61810-4 Burgeon J. 2005. Plantes toxiques (Végétaux dangereux pour l'homme et les animaux). Ed. Tec & Doc. Lavoisier, Paris, 3ème édition. ISBN: 2-7430-0806-7 Frome D, Pfander HU and Anton R. 2009. Plantes à risques. Ed. Tec et Doc-Lavoisier. ISBN: 978-2-7430-0907-1
<i>Thuja</i> spp.	Cupressaceae	Whole plant	Genus in which the essential oil of some species may contain the bicyclic monoterpenes thujones		Frome D, Pfander HU and Anton R. 2009. Plantes à risques. Ed. Tec et Doc-Lavoisier. ISBN: 978-2-7430-0907-1
<i>Thymus</i> spp.	Lamiaceae (Labiatae)	Aerial part	Genus in which the essential oil of the species contain a variety of constituents including the monoterpane etheroxide 1-8-cineole	Many species show different chemotypes.	Teuscher E, Anton R, et Losten A. 2005. Plantes aromatiques (Eucies, aromates, condiments et huiles essentielles). Ed. Tec et Doc-Lavoisier. ISBN : 2-7430-0720-6 Bourke CA, et al. 1992. Locomotor effects in sheep of alkaloids identified in Australian <i>Thibulus terrestris</i> . Aust. Vet. J. 69: 163-165 Dinchev D, et al. 2008. Distribution of steroidal saponins in <i>Thibulus terrestris</i> from different geographical regions. Phytochemistry. 69: 176-186. Gauthaman K, et al. 2002. Aphanistic properties of <i>Thibulus terrestris</i> extract (protodeson) in normal and castrated rats. Life. Sci. 71: 1385-1396. Paula-Lopes TRV, et al. 2006. Hepatotoxicity of medicinal plants. XXXIII. Action of <i>Thibulus terrestris</i> L. in rats. Rev Bras Fisiol Med 8: 4: 150-156 Kellerman TS et al. 1990. Photosensitivity in South Africa. II. The experimental production of the ovine hepatogenous photosensitivity disease geelklopp (<i>Thibulus</i> ovis) by the simultaneous ingestion of <i>Thibulus terrestris</i> plants and cultures of <i>Phytomyces chlararum</i> containing the mycotoxin sporidesmin. Onderstepoort J Vet Res. 47(4):231-61.
<i>Tribulus terrestris</i> L.	Zygophyllaceae	Whole plant	β-carboline alkaloids (40-80 mg/kg dry matter), e.g. harmaline and norharmaline Lithogenic steroidal saponins: e.g. protodeson Mycotoxin: sporidesmin	Central Nervous System toxicity observed in sheep. Hepatotoxicity observed in male rats after oral administration of the fruit. Reported effect on testosterone levels and prostate weight following administration of a fruit extract with high protodeson level to castrated male rats	Frome D, Pfander HU and Anton R. 2009. Plantes à risques. Ed. Tec et Doc-Lavoisier. ISBN: 978-2-7430-0907-1 Copper RA, et al. 1996. Preparative separation of pyrrolizidine alkaloids by high-speed counter-current chromatography. J Chromatogr A. 732(1): 43-50. Burgeon J. 2009. Pharmacognosie, (Phytochimie, Plantes médicinales). Ed. Tec & Doc Lavoisier, Paris, 4ème édition. ISBN: 978-2-7430-1189-8 Méranger JM, Et al. 1987. Purification and characterisation of <i>Trichosanthin</i> . J Biol Chem. 262(24): 11623-11633
<i>Trichocereus</i> spp.	Cactaceae	Whole plant	Genus in which species may contain phenylethylamine alkaloids: e.g. mescaline, ...		Jürgens A, and Doherr S. 2004. Chemical composition of arther volatiles in Ranunculaceae genera-specific profiles in Armonio, Aquilaga, Cairna, Pissallia, Ranunculus, and <i>Trollius</i> species. American Journal of Botany. 91: 1989-1990.
<i>Trichodesma incanum</i> Bunge	Borraginaceae	Aerial part	Unsaturated pyrrolizidine alkaloid: trichodesmine		
<i>Trichosanthes kirilowii</i> Maxim.	Cucurbitaceae	Root	Polypeptide: trichosanthin		
<i>Trollius europaeus</i> L.	Ranunculaceae	Whole plant	Unsaturated lactone : protoanemonin	Protoanemonin only present in fresh plant	
<i>Tropis aspera</i> Retz.					
<i>Tropis asper</i> (Retz.) Lour.					
<i>Tulipa</i> spp.	Liliaceae	Whole part	Phytoalexins: e.g. tulipain		Irritation of the bulbs have given rise to acute symptoms in humans like sweating, increased salivation, difficult breathing and vomiting. Feeding of large amounts of tulip bulbs to cows resulted in 14/50 deaths within 6 weeks.
<i>Turbinaria corymbosa</i> (L.) Raf. (Alveolata: Rhodophyta)	Convolvulaceae	Leaf and seed	Indole alkaloids (ergoline alkaloids, lysergic acid derivatives). Dried leaf: 0.016-0.027 ergoline alkaloids (egmin and espinin). Dried stem: 0.010-0.012 ergoline alkaloids (ergine and espinine).		Hagers Handbuch der Pharmazeutischen Praxis. Springer Verlag. 1998. ISBN: 3-540-52688-9

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<i>Thyphora aspinifolia</i> Wight & Arn. See <i>Thyphora indica</i> Merr.					
<i>Thyphora indica</i> Merr. (<i>T. aspinifolia</i> Wight & Arn., <i>Cynanchum indicum</i> Burtt.)	Apocynaceae	Leaf and root	Indolizidine alkaloids: e.g. lybopherine, lycosidine, lybophorinine		Daniel M. 2005. Medicinal Plants, Chemistry & Properties. Oxford & IBH Publishing Company Pvt. Ltd. ISBN 8120416899, 9788120416895. Singra A. et al. 2011. <i>Thyphora aspinifolia</i> Wight & Arn. - Review. Nature of Pharmaceutical Technology, 11(1), 1-4.
<i>Urginea</i> spp.	Asparagaceae	Bulb	Genus in which species may contain butadienolide glycosides and their aglycones: e.g. glucoscleraine, scleraine, scleraverine,....		Hagers Handbuch der Pharmazeutischen Praxis, Springer Verlag, 1998. ISBN: 3-540-52688-9 EMEA/CMP. 1998. <i>Urginea maritima</i> - summary report. EMEA/ANL/603/99.
<i>Usnea</i> spp.	Parmeliaceae	Lichen	Genus in which species may contain the dibenzodipyrone usnic acid		Burleton J. 2009. Pharmacognosie, (Phytochimie, Plantes médicinales), Ed. Tec & Doc Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1188-8 Gastronardot Hepato, 20(7), 1138-1139. Sanchez W. et al. 2006. Severe hepatotoxicity associated with use of a dietary supplement containing usnic acid. Mayo Clin Proc. 81(4):541-4. Guo L. et al. 2008. Review of usnic acid and <i>Usnea barbata</i> toxicity. J Environ Sci Health C (Toxicol Environ Carcinog Ecotoxicol Rev.) 28(4):317-38. Heath C. (Ed.) 2008. Pharmacognosie, (Phytochimie, Plantes médicinales), Ed. Tec & Doc Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1188-8
<i>Valerianopsis arborae</i> (Gardner) Baker (<i>Eriarthus arborae</i> (Gardner) MacLisih)	Asteraceae	Leaf and wood bark	Essential oil from wood bark: phenylpropanoids: methylchavicol (3.6 %), methyl Eugenol (5.9%), elemicin (2.7%) Essential oil from leaf: phenylpropanoid: safrole (0.74%),		Hagers Handbuch der Pharmazeutischen Praxis, Springer Verlag, 1998. ISBN: 3-540-52688-9 Santos NKA. 2011. Chemical characterization and synergistic antibiotic activity of volatile compounds from the essential oil of <i>Valerianopsis arborae</i> . Med. Chem. Res. 20(5), 637-641.
<i>Valeriana araroba</i> (Aquilari) Ducke (<i>Andira araroba</i> Agilar)	Leguminosae (Fabaceae)	Wood	Hydroxyanthracene: chrysarbinol (1,8-hydroxy-3-methylanthranol)		Burleton J. 2009. Pharmacognosie, (Phytochimie, Plantes médicinales), Ed. Tec & Doc Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1188-8
<i>Veratrum</i> spp.	Melasthaceae	Whole plant	Genus in which species may contain steroid alkaloids: e.g. protoveratrine, and alkaline esters such as jerine derivatives (furanopyridine), e.g. cyclopamine.		Burleton J. 2009. Pharmacognosie, (Phytochimie, Plantes médicinales), Ed. Tec & Doc Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1188-8 Frohne D, Pfänder HJ and Anton R. 2009. Plantes à risques, Ed. Tec et Doc-Lavoisier, ISBN: 978-2-7430-0907-1
<i>Vicia</i> spp.	Apocynaceae	Whole plant	Genus in which species may contain iridoid alkaloids: e.g. vincamine		Burleton J. 2009. Pharmacognosie, (Phytochimie, Plantes médicinales), Ed. Tec & Doc Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1188-8
<i>Vincetoxicum hircundaria</i> Medik. (<i>V. officinale</i> Moench, <i>Cynanchum vincetoxicum</i> (L.) Pers.)	Apocynaceae	Whole plant	Iscourolidic alkaloids: e.g. asclepiadine (vinetoxin analogs). Root: vinetoxin analogs: heterosides of oxosteroids (thuridicoside B, C and D, Cyanatoside C and E). Aerial part: Phenanthroindolizidine alkaloids (10 beta-anilino-N-oxide, 10 beta,13 alpha,14 beta-hydroxyanilino-N-oxide, 10 beta,13alpha-secoanilino-N-oxide, also 13alpha-secoanilino, 13alpha-6-O-desmethylsecoanilino).		Stærk D. et al. 2002. In Vitro Cytotoxic Activity of Phenanthroindolizidine Alkaloids from <i>Cynanchum vincetoxicum</i> and <i>Thyphora tanaka</i> against Drug-Resistant and Multidrug-Resistant Cancer Cells. J. Nat. Prod. 65, 1289-1302. Burleton J. 2009. Pharmacognosie, (Phytochimie, Plantes médicinales), Ed. Tec & Doc Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1188-8 Frohne D, Pfänder HJ and Anton R. 2009. Plantes à risques, Ed. Tec et Doc-Lavoisier, ISBN: 978-2-7430-0907-1
<i>Vincetoxicum nigrum</i> Moench. (<i>Asclepias vincetoxicum</i> L.)	Apocynaceae	Whole plant	Phenanthroindolizidine alkaloids: e.g. (-)-santidine		Gibson DM et al. 2011. Phytotoxicity of aniridine from invasive swallow-worts. J. Chem. Ecol. 37(8), 871-9.
<i>Viscum album</i> L.	Santalaceae	Whole plant	Peptides: viscotoxins (I, II, III) and glycoproteins: viscum lectins		Hagers Handbuch der Pharmazeutischen Praxis, Springer Verlag, 1998. ISBN: 3-540-52688-9 Council of Europe. 2008. Natural sources of Flavonings. Report No. 3. Council of Europe Publishing. ISBN 978-92-871-6422-3 Burleton J. 2005. Plantes toxiques (Végétaux dangereux pour l'homme et les animaux), Ed. Tec & Doc, Lavoisier, Paris, 3ème édition, ISBN: 2-7430-0806-7
<i>Vitex agnus-castus</i> L.	Lamiaceae (Labiatae)	Aerial part	The yield of essential oil in dried plant material is 0.76% from urtipe fruits, 0.72% from the fruits, 0.56% from aerial parts. Essential oil from fruit: monoterpane etheroxide: 1,8-dineol (16-18%) and bicyclic monoterpenes: e.g. sabinene (7-17%) Essential oil from leaf: monoterpenes: e.g. 1,8-dineol (22-33%) and sabinene (2-18%) Essential oil from flower: monoterpenes: e.g. 1,8-dineol (13.5%) and sabinene (5.7%)	Synges of liver toxicity were observed in two new repeat-dose toxicity studies on extracts of the fruit. Administration of powdered seeds resulted in a slight reduction in the number of foetuses when administered to pregnant rats in doses of 1 or 2 mg/kg from days 1-10 of pregnancy. A lactation inhibiting effect (decrease of prolactin) was seen in lactating female rats dosed with a <i>Vitex agnus-castus</i> preparation. <i>In Vitro</i> studies with cells from rat pituitaries showed that an extract had a dose-dependent lowering effect of prolactin.	EMA. 2010. Community herbal monograph on <i>Vitex agnus-castus</i> L., fructus. www.ema.europa.eu Lal R. et al. 1985. Antifebrile and oxytocic activity of <i>Vitex agnus-castus</i> seeds in female albino rats. Bulletin of Postgraduate Institute of Medical Education and Research, Chandigarh 19, 44-47 Stojkovic D. et al. 2011. Chemical composition and antimicrobial activity of <i>Vitex agnus-castus</i> L. fruits and leaves essential oils. Food Chem. 128, 1017-1022. Winterhoff et al. 1991. Die Hemmung der Laktation bei Ratten als indirekter Beweis für die Senkung von Prolaktin durch <i>Agnus castus</i> . Z. Phytotherapie 12, 175-179. Zogbi M.D.C.B. et al. 1999. The essential oil of <i>Vitex agnus-castus</i> growing in the Amazon region. Flavour-Frag. J. 14, 211-213.
<i>Vocanga</i> spp.	Apocynaceae	Bark and root	Genus in which species may contain iridoid alkaloids: e.g. coronaridine, vocangine, conopharyngine.		Julian HR. et al. eds. 2009. African Natural Plant Products: New Discoveries and Challenges in Chemistry and Quality. Vol 1021. American Chemical Society, ISBN: 9780941268973. Burleton J. 2009. Pharmacognosie, (Phytochimie, Plantes médicinales), Ed. Tec & Doc Lavoisier, Paris, 4ème édition, ISBN: 978-2-7430-1188-8 Council of Europe. 2008. Natural sources of Flavonings. Report No. 3. Council of Europe Publishing. ISBN 978-92-871-6422-3.
<i>Wistaria orobancha</i> (Willd.) DC.	Thymelaeaceae	Whole plant	Genus in which species may contain diterpenes: daphnane orthoesters: e.g. hurakoxine, simplekxine		Sakai S. et al. 1997. A Self-Adjusting Carboxyvirane Ligand for Galinac Specific Lectins. Tetrahedron Letters, 38(47), 8145-8148.
<i>Wistaria floribunda</i> (Willd.) DC.	Leguminosae (Fabaceae)	Whole plant	Lectins		Nasir A. et al. 2009. Proteomic approaches to study structure, functions and toxicity of legume seeds lectins. Perspectives for the assessment of food quality and safety. Journal of Proteomics, 72, 527-538.
<i>Wistaria sinensis</i> (Sims) DC.	Leguminosae (Fabaceae)	Whole plant	Lectins		

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<i>Withania somnifera</i> (L.) Dunal	Solanaceae	Whole plant	In leaf: steroidal lactones, withanolides in root: piperidine alkaloids, artemierine, anahygrine and various alkaloids including withanine, somniferine, somnibe, tropine		Burton J. 2009. Pharmacognosy, (Phytochemistry, Plantae medicinales), Ed. Tec & Doc. Loviser, Paris, 4ème édition, ISBN : 978-2-7430-1188-8 Kulkarni SK, Dhir A. 2008. Withania somnifera: an Indian ginseng. Prog Neuro-psychopharmacol Biol Psychiatry, 32(5):1093-105 Misra LN. 2008. Selective reactivity of 2-mecapoteinanol with betaia beta-epoxide in sterols from Withania somnifera. Steroids, 73(3):242-251
<i>Xanthium</i> spp.	Compositae (Asteraceae)	Flowering top	Genus in which species may contain diterpenes, e.g. carboxyatractyloside	Toxicosis usually associated with the consumption of the seedlings in the cotyledon stage which contain a high concentration of carboxyatractyloside. Seeds also known to contain the toxin	Witte ST, et al. 1990. Cockspur toxicosis in cattle associated with the consumption of mature Xanthium strumarium. J Vet Diagn Invest. 2(4):283-287 Ghorbani M. 1997. Phytochemical reinvestigation of <i>Xysmalobium undulatum</i> roots (Uzara), <i>Phlaria Mendica</i> B3(4), 342-346
<i>Xysmalobium undulatum</i> (L.) R.Br.	Apocynaceae	Root	Cardenolides (saponoside types) e.g. uzarin, xysmaloin,		
<i>Yucca filamentosa</i> L.	Asparagaceae	Whole plant	Steroid saponins: e.g. sarsasapogenin, tigogenin (14% in leaves)		Dragalin IP, and Kriml P K. 1975. Steroidal saponins of <i>Yucca filamentosa</i> 'Yuccoside C' and protyucoside C. Phytochemistry 14 (8), 1817-1820. Knyla PK. 1972. Steroid saponins II. Glycosides A and B from <i>Yucca filamentosa</i> . Chemistry of Natural Compounds, 8(5), 584-586.
<i>Zanthoxylum alatum</i> Robb.	Rutaceae	Bark and seed	Furocoumarins: e.g. bergapten, umbelliferone, Benzofuranthrine alkaloids, e.g. chelerythrine and derivatives		Akhtar N. 2009. Chemical constituents from the seeds of <i>Zanthoxylum alatum</i> . Journal of Asian Natural Products Research, 11(1), 91-95.
<i>Zanthoxylum americanum</i> Mill.	Rutaceae	Bark and seed	Benzylisoquinoline alkaloids: e.g. magnolone, benzofuranthrine alkaloids, e.g. chelerythrine,	Lignans: asarinin and sesamin	Burton J. 2009. Pharmacognosy, (Phytochemistry, Plantae medicinales), Ed. Tec & Doc. Loviser, Paris, 4ème édition, ISBN : 978-2-7430-1188-8 American Herbal Products Association. 1997. Botanical Safety Handbook. Mc Guffin M (Ed). ISBN: 0-8893-1675-8
<i>Zanthoxylum ciliv-herculis</i> L.	Rutaceae	Bark and seed	Benzylisoquinoline alkaloids: e.g. magnolone, benzofuranthrine alkaloids, e.g. chelerythrine,	Lignans: asarinin and sesamin	Burton J. 2009. Pharmacognosy, (Phytochemistry, Plantae medicinales), Ed. Tec & Doc. Loviser, Paris, 4ème édition, ISBN : 978-2-7430-1188-8 American Herbal Products Association. 1997. Botanical Safety Handbook. Mc Guffin M (Ed). ISBN: 0-8893-1675-8
<i>Zigadenus</i> spp.	Melanthaceae	Whole plant	Genus in which species may contain steroid alkaloids, e.g. zygadenine, zygacine		Moerman DE. 1996. An analysis of the food plants and drug plants of native North America. Journal of Ethnopharmacology, 52, 1-22. EMA. 2011. Assessment report on Zingiber officinale Roscoe, rhizoma. www.ema.europa.eu Wilkinson JM. 2000. Effect of ginger tea on the fetal development of Sprague-Dawley rats. Reprod. Toxicol. 14, 507-512.
<i>Zingiber officinale</i> Roscoe	Zingiberaceae	Rhizome		An increased percentage of resorbed implantations were found in both dosed groups of pregnant rats receiving an infusion made from 20 g/l or 50 g/l freshly grated ginger compared to a control group (p<0.05). No signs of maternal toxicity were observed, neither were any gross morphological malformations in the treated foetuses.	
FUNGI					
<i>Amanita</i> spp.	Amanitaceae	Fruiting body	Genus in which species may contain trypamine: e.g. biforoline, cyclic peptides; e.g. phallotoxins and amatoxins; isoxazole alkaloids, e.g. ibotenic acid and xerodermy ammonium alkaloids, e.g. muscarine.		Evens W. 2009. Pharmacognosy, 16th edition, Saunders, Ltd. ISBN: 978-0-7020-2932-2
<i>Boletus satanas</i> Lenz	Boletaceae	Fruiting body	Monomeric glycoprotein, bolesatine.		Emmamy R et al. 1995. Mode of action of bolesatine, a cytotoxic glycoprotein from <i>Boletus satanas</i> Lenz. Mechanistic approaches. Toxicology, 100(1-3), 51-55 Kretz O et al. 1992. Properties of bolesatine, a transitional inhibitor from <i>Boletus satanas</i> Lenz. Amino-terminal sequence determination and inhibition of rat mitochondrial protein synthesis. Toxicology Letters, 64-65, 763-766. Emmamy R et al. 1994. Effect of bolesatine, a glycoprotein from <i>Boletus satanas</i> , on rat thymus in vivo. Toxicology, 89(2), 113-118
<i>Clicocybe</i> spp.	Trichomonataceae	Fruiting body	Genus in which species may contain muscarine (<i>C. dealbata</i> , <i>C. involuta</i> ...), some riche-like lectins and ciliocybins (indolines)		Ian R. Hall et al. 2003. Edible and poisonous mushrooms of the world. Timber Press USA, ISBN: 0-88192-586-1 Gensel K, et al. 1988. Muscarine in <i>Clicocybe</i> species. J Pharm Sci. 57(2):331-3. Svaliger U et al. 2011. CNL, a ricin B-like lectin from mushroom <i>Clicocybe nebularis</i> , induces maturation and activation of dendritic cells via the toll-like receptor 4 pathway. Immunology, 134(4), 409-18
<i>Cornucybe</i> spp.	Bolbitaceae	Fruiting body	Genus in which species may contain inside alkaloids (tryptamine derivatives), e.g. psilocin, psilocybin...	Some species are known to contain proteoglycans such as phalloidin inducing gastrointestinal disorders and anxiety	Hall R et al. 2003. Processing of the phalloidin protein by prolyl oligopeptidase from the mushroom <i>Cornucybe</i> albidus. J Biol Chem. 284(27), 18070-18077 Burton J. 2009. Pharmacognosy, (Phytochemistry, Plantae medicinales), Ed. Tec & Doc. Loviser, Paris, 4ème édition, ISBN: 978-2-7430-1188-8
<i>Copelandia</i> spp.	Agaricomycetidae	Fruiting body	Genus in which species may contain inside alkaloids (tryptamine derivatives), e.g. psilocin, psilocybin		Merin MD, et al. 1993. Species identification and chemical analysis of psychotrope fungi in the Hawaiian Islands. J Ethnopharmacol. 40(1), 21-40. Hall R. 2003. Edible and Poisonous Mushrooms of the World. Timber Press, p. 103. ISBN: 0881925861 Helm R, et al. 1966. On a group poisoning with psilocybin syndrome caused in France by a <i>Copelandia</i> . C R Acad Sci. Heild Sciences Acad Sci D. 262(4), 519-523 Gomori K, et al. 2009. Acute encephalopathy caused by psilocybin, a magic mushroom, in 2004, and magic mushroom regulation in Japan. Chudoku Kenkyu. 22(1), 61-69.
<i>Cortinaurus orellanus</i> Fr.	Cortinariaceae	Fruiting body	Pyridine N-oxide alkaloids: e.g. orellanine and derivatives Bipyridine alkaloids		Duvc C et al. 2003. Acute renal failure following ingestion of <i>Cortinaurus orellanus</i> in 12 patients. Initial presentation and progress over a period of 13 Years. Presse Med. 32(6), 249-253 Oudraim H et al. 1997. Novel methods for identification and quantification of the mushroom nephrotoxin orellanine. Thin-layer chromatography and electrophoresis screening of mushrooms with electron spin resonance determination of the toxin. J Chromatogr. 758(1), 145-157 Judge BS et al. 2010. Ingestion of a newly described North American mushroom species from Michigan resulting in chronic renal failure. <i>Cortinaurus orellanus</i> . Clin Toxicol (Phila). 48(6):545-549

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<i>Cortinarius rubellus</i> Cooke (Cortinarius speciosissimus Kuhnér & Romagn.) <i>Cortinarius speciosissimus</i> Kuhnér & Romagn. <i>See Cortinarius rubellus</i> Cooke	Cortinariaceae	Fruiting body	Pyridine N-oxide alkaloids: e.g. orsellanine and derivatives Bicyclic alkaloids		Koller GE et al. 2002. The presence of orsellanine in spores and basidiocarp from <i>Cortinarius orellanus</i> and <i>Cortinarius rubellus</i> . Mycologia 94(5): 752-756.
<i>Galerina marginata</i> (Batsch) Kuhnér	Galeriniaceae	Fruiting body and mycelium	Bicyclic octapeptide derivatives: amatoxins (alpha-, beta- and gamma amanitins)		Erlanger F., et al. 2004. Amatoxins in wood-rotting <i>Galerina marginata</i> . Mycologia 96(4): 720-729
<i>Gyromitra esculenta</i> (Pers.) Fr.	Discinaceae	Fruiting body	Hydrazones: gyromitrin (acetaldehyde N-methyl-N-(methylhydrazono) approximately 50 mg/kg fresh weight)		Andersson C. et al. 1995. Hydrazones in the false morel, <i>Trematium</i> 561. Nordic Council of Ministers. ISBN 92 9120 681 4.
<i>Hevelia</i> spp.	Hebeliaceae	Fruiting body		Intoxication may be caused by <i>Hevelia</i> species. The species are often confused with <i>Gyromitra</i> species, the latter being known to contain toxic hydrazones (gyromitrin).	Beug MW, et al. Thirty-plus years of mushroom poisoning: summary of the approximately 2,000 reports in the NIAID case registry. <i>Mycetozoa</i> 16(2): 41-68. http://mycozoo2.org/mushres/pdf_files/03mush30year.pdf
<i>Hevelia</i> spp.	Hebeliaceae	Fruiting body		Valeral thought to be responsible for toxicity in humans.	Lurie Y. et al. 2009. Mushroom poisoning from species of genus <i>Hevelia</i> (fiber head mushroom): a case series with exact species identified. <i>Chem. Toxicol.</i> 47: 562-565.
<i>Hevelia</i> spp.	Hebeliaceae	Fruiting body		Valeral thought to be responsible for toxicity in humans.	Slyve T. 1992. The occurrence of muscarine and muscimol in various fungi. <i>Coelia</i> 25(6): 94-100.
<i>Hevelia</i> spp.	Hebeliaceae	Fruiting body		Valeral thought to be responsible for toxicity in humans.	Camraxe S. and Lupo A. T. 1994. Labile toxic compounds of the tricarbit, the role of the lacticidous hyphae as a storage depot for precursors of purgative diacylglycerols. <i>Mycologia</i> 102(2): 355-358.
<i>Hevelia</i> spp.	Hebeliaceae	Fruiting body		Valeral thought to be responsible for toxicity in humans.	Rindunen J. and von Wright A. 1992. The mutagenicity of <i>Lactarius</i> mushrooms. <i>Mut. Res.</i> 103: 115-118.
<i>Hevelia</i> spp.	Hebeliaceae	Fruiting body		Valeral thought to be responsible for toxicity in humans.	Pyysalo H. et al. 1990. Application of gas chromatography to the analysis of sesquiterpene lactones from <i>Lactarius</i> (Russulaceae) mushrooms. <i>J. Chromatogr.</i> 190: 469-470.
<i>Hevelia</i> spp.	Hebeliaceae	Fruiting body		Valeral thought to be responsible for toxicity in humans.	von Wright A. et al. 1992. The mutagenicity of some edible mushrooms in the Ames test. <i>Food Chem. Toxicol.</i> 20: 285-297.
<i>Hevelia</i> spp.	Hebeliaceae	Fruiting body		Valeral thought to be responsible for toxicity in humans.	Rennet F. et al. 1993. Fungal liver failure after <i>Lepidia</i> mushroom poisoning. <i>J. Hepatol.</i> 19: 374-376.
<i>Lepidia</i> spp.	Agaricaceae	Fruiting body	Genus in which species may contain cyclopeptide toxins (amatoxins), e.g. amanitine A and B		Andersson C., Kristianson J., Gyr J. 2008. Occurrence and use of halitogenic mushrooms containing psilocybin alkaloids. <i>Tenthredo</i> 2008/006. ISBN: 978-92-983-6393-3.
<i>Lepidia</i> spp.	Agaricaceae	Fruiting body	Genus in which species may contain cyclopeptide toxins (amatoxins), e.g. amanitine A and B		Andersson C., Kristianson J., Gyr J. 2008. Occurrence and use of halitogenic mushrooms containing psilocybin alkaloids. <i>Tenthredo</i> 2008/006. ISBN: 978-92-983-6393-3.
<i>Paranodus</i> spp.	Agaricomycetidae	Fruiting body	Genus in which species may contain indole alkaloids (tryptamine derivatives), e.g. psilocin, psilocybin		Andersson C., Kristianson J., Gyr J. 2008. Occurrence and use of halitogenic mushrooms containing psilocybin alkaloids. <i>Tenthredo</i> 2008/006. ISBN: 978-92-983-6393-3.
<i>Pluteus</i> spp.	Pileateae	Fruiting body	Genus in which species may contain indole alkaloids (tryptamine derivatives), e.g. psilocin, psilocybin		Strömberg J. et al. 2003. Levels of psilocybin and psilocin in various types of mushrooms. <i>Scand. J. Lab. Diagnostics</i> 44(5): 45-46.
<i>Pluteus</i> spp.	Pileateae	Fruiting body	Genus in which species may contain indole alkaloids (tryptamine derivatives), e.g. psilocin, psilocybin		Reigfänger D. et al. 2005. Halitogenic mushrooms. <i>Medicina</i> (Kaisers), 41(12): 1007-1070.
<i>Pluteus</i> spp.	Pileateae	Fruiting body	Genus in which species may contain indole alkaloids (tryptamine derivatives), e.g. psilocin, psilocybin		Andersson C. et al. 2007. Halitogenic fungi (psilocyba). Part II. Identification of <i>Psilocybe</i> spp. (partially by PCR). <i>Acta Societatis Scientiarum Fennicarum</i> 57(3): 295-298.
<i>Psilocybe</i> spp.	Strophariaceae	Fruiting body	Genus in which species may contain indole alkaloids (tryptamine derivatives), e.g. psilocin and psilocybin		Edwards VC. (2009) <i>Trace and Excess Pharmacology</i> . Elsevier. ISBN: 978-0-7020-2932-2
<i>Stropharia</i> spp.	Strophariaceae	Fruiting body	Genus in which species may contain indole alkaloids (tryptamine derivatives), e.g. psilocin, psilocybin		
ALGAE					
<i>Ascophyllum nodosum</i> (L.) Le Jolis	Fucales	Thallus		Known to contain high levels of iodine (on average 482 µg/g dry weight)	Pfander D et al. 1999. Evaluation of the contamination of marine algae (seaweed) from the St. Lawrence River and likely to be consumed by humans. <i>Environ. Res. Section A</i> 80: S175-S182.
<i>Fucus</i> spp.	Fucales	Thallus		Genus in which species may contain different amounts of iodine depending on growth conditions and environment, the algae may concentrate heavy metals (e.g. Pb, Cd)	Tears J. et al. 2004. Variability of Iodine Content in Common Commercially Available Edible Seaweeds. <i>Thyroid</i> 14(10): 836-841
<i>Fucus</i> spp.	Fucales	Thallus		Genus in which species may contain different amounts of iodine depending on growth conditions and environment, the algae may concentrate heavy metals (e.g. Pb, Cd)	Bruteton J. 2009. <i>Pharmacognosy: Phytochemistry, Planes médicinales</i> . Ed. Tec & Doc. Elsevier. Paris. 4ème édition. ISBN: 978-2-7430-1188-8
<i>Macrocystis pyritera</i> (L.) C.Ag.	Marsipposaceae (Laminariaceae)	Thallus		May contain high levels of iodine. Depending on growth conditions and environment, the algae may concentrate heavy metals (e.g. Pb, Cd)	Barnes J., Anderson A., Phillips J.D. 2007. <i>Herbal Medicines</i> . 3rd ed. Ed. Pharmaceutical Press. ISBN: 978-0-85369-623-0
					Seki H., Anzaki A. 1998. Biosorption of heavy metal ions to brown algae, <i>Macrocystis pyritera</i> , <i>Keikimantella cassiniana</i> , and <i>Undaria pinnatifida</i> . <i>J. Colloid and Interface Science</i> 206: 297-301.

Botanicals appearing on a negative list or subject to restricted use in at least one European Member State but for which not enough information on possible substances of concern or adverse effect could be found, or for which the information present could not be verified.

Botanical name	Family	Part of plants indicated in the national list(s)
<i>Acanthea virilis</i> Pharm. ex Wehmer	Acanthaceae	
<i>Aletris farinosa</i> L.	Nartheciaceae	Root
<i>Anemarrhena asphodeloides</i> Bunge	Asparagaceae	Rhizome
<i>Anemopaegma arvense</i> (Vell.) Stellfeld	Bignoniaceae	Bark
<i>Aniba parviflora</i> (Meisn.) Mez	Lauraceae	
<i>Anthyllis vulneraria</i> L.	Leguminosae (Fabaceae)	
<i>Aquilaria malaccensis</i> Lam. (<i>Aquilaria agollocha</i> Roxb.)	Thymelaeaceae	Bark and leaf
<i>Artemisia apiacea</i> Hance See <i>Artemisia carvifolia</i> Roxb.		
<i>Artemisia carvifolia</i> Roxb. (<i>Artemisia caruifolia</i> Roxb., <i>Artemisia apiacea</i> Hance)	Compositae (Asteraceae)	Aerial part
<i>Artemisia nitida</i> Bertol.	Compositae (Asteraceae)	
<i>Asplenium scolopendrium</i> L.	Aspleniaceae	
<i>Atractylodes macrocephala</i> Koidz.	Compositae (Asteraceae)	Rhizome
<i>Atractylodes ovata</i> DC.	Compositae (Asteraceae)	Rhizome
<i>Betula alleghaniensis</i> Britton	Betulaceae	
<i>Blepharis capensis</i> Pers.	Acanthaceae	Leaf and root
<i>Cachrys pabularia</i> (Lindl.) Herrnstadt & Heyn (<i>Prangos pabularia</i> Lindl.)	Apiaceae (Umbelliferae)	Seed
<i>Calendula arvensis</i> L.	Compositae (Asteraceae)	Flower
<i>Calystegia soldanella</i> R.Br.	Convolvulaceae	Whole plant
<i>Catalpa bignonioides</i> Walter	Bignoniaceae	Bark and fruit
<i>Ceanothus americanus</i> L.	Rhamnaceae	Root bark
<i>Cedrela toona</i> Roxb. See <i>Toona ciliata</i> M.Roem.		Bark
<i>Cedrus deodara</i> (D.Don) G.Don	Pinaceae	Wood
<i>Cedrus libani</i> A.Rich.	Pinaceae	Aerial part
<i>Chamaemelum mixtum</i> All.	Compositae (Asteraceae)	Whole plant
<i>Chlorocodon whitei</i> Hook.f. (<i>Mondia whitei</i> (Hook.f.) Skeels)	Apocyanaceae	Root
<i>Chrozophora tinctoria</i> (L.) A.Juss.	Euphorbiaceae	Aerial part
<i>Chrysanthemum sinense</i> Sweet (<i>Chrysanthemum morifolium</i> Ramat., <i>Tanacetum morifolium</i> Kitam.)	Compositae (Asteraceae)	Flower
<i>Chrysopogon zizanioides</i> (L.) Roberty	Poaceae (Gramineae)	Root
<i>Cineraria</i> spp.	Compositae (Asteraceae)	Aerial part
<i>Cinnamomum micranthum</i> (Hayata) Hayata	Lauraceae	Wood
<i>Citrus junos</i> (Makino) Tanaka	Rutaceae	Fruit

Botanical name	Family	Part of plants indicated in the national list(s)
Citrus medica L. (<i>C. medica</i> (L.) var. <i>macrocarpa</i> Risso; <i>C. medica</i> (L.) var. <i>vulgaris</i> Risso; <i>C. medica</i> L. var. <i>cedrata</i> Risso)	Rutaceae	
Cnidium dubium (Schkuhr) Schmeil & Fitschen	Apiaceae (Umbelliferae)	
Cnidium officinale Makino	Apiaceae (Umbelliferae)	Rhizome
Cola ballayi Cornu ex Heckel	Malvaceae	
Comandra spp.	Santalaceae	
Conioselinum univittatum Kar. & Kir.	Apiaceae (Umbelliferae)	
Convolvulus scoparius L.f.	Convolvulaceae	
Copaifera martii Hayne	Leguminosae (Fabaceae)	
Dicoma anomala Sond.	Compositae (Asteraceae)	Root
Dirca palustris L.	Thymelaeaceae	
Dorema ammoniacum D.Don.	Apiaceae (Umbelliferae)	
Eclipta prostrata (L.) L.	Compositae (Asteraceae)	Whole plant
Erythroxylum catuaba Raym.-Hamet	Erythroxylaceae	
Eupatorium perfoliatum L.	Compositae (Asteraceae)	Aerial part
Eupatorium triplinerve Vahl (<i>Ayapana triplinervis</i> (Vahl) R.M.King & H.Rob.)	Compositae (Asteraceae)	Whole plant
Ferula gummosa Boiss. (<i>F. galbaniflua</i> Boiss. & Bushe)	Apiaceae (Umbelliferae)	Aerial part
Forestiera spp.	Oleaceae	
Fortunella japonica (Thunb.) Swingle (<i>Citrus japonica</i> Thunb.)	Rutaceae	Bark
Geranium maculatum L.	Geraniaceae	
Gnaphalium uliginosum L.	Asteraceae (Compositae)	
Grindelia camporum Greene	Asteraceae (Compositae)	
Grindelia hirsutula Hook. & Arn.	Asteraceae (Compositae)	
Guaicum sanctum L.	Zygophyllaceae	
Gymnema aurantiacum Hook.f.	Apocyanaceae	
<i>Gymnema sylvestre</i> (Retz.) Schult. See Marsdenia sylvestris (Retz.) P.I.Forst		
Gypsophila paniculata L.	Caryophyllaceae	
Haematoxylum campechianum L.	Leguminosae (Fabaceae)	Bark and wood
Harrisonia abyssinica Oliv.	Rutaceae	Bark and root
Helichrysum nudifolium Less.	Asteraceae (Compositae)	
Herniaria hirsuta L.	Caryophyllaceae	Aerial part
Homalomena spp.	Araceae	
Hydrangea arborescens L.	Hydrangeaceae	Root
Indigofera tinctoria L.	Leguminosae (Fabaceae)	Unspecified
Ionidium ipecacuanha Vent.	Violaceae	Root
Iris versicolor L.	Iridaceae	Rhizome
Jacaranda caroba (Vell.) DC.	Bignoniaceae	
Juniperus ashei J.Buchholz	Cupressaceae	

Botanical name	Family	Part of plants indicated in the national list(s)
<i>Juniperus procera</i> Hochst. ex Endl.	Cupressaceae	Wood
<i>Kunzea ambigua</i> (Sm.) Druce	Myrtaceae	
<i>Lavandula burnati</i> Briq. See <i>Lavandula intermedia</i> nothosp. <i>intermedia</i>		
<i>Lavandula heterophylla</i> Viv. (<i>Lavandula hybrida</i> Ging.)	Lamiaceae (Labiatae)	
<i>Lavandula hybrida</i> Ging. See <i>Lavandula heterophylla</i> Viv.		
<i>Lavandula intermedia</i> nothosp. <i>intermedia</i> (<i>Lavandula x burnati</i> Briq.)	Lamiaceae (Labiatae)	
<i>Lebeckia contaminata</i> (L.) Thunb. (<i>Asphaltus contaminatus</i> (L.) Druce, <i>Spartium contaminatum</i> L.)	Leguminosae (Fabaceae)	
<i>Leopoldia comosa</i> (L.) Parl. (<i>Muscari comosum</i> (L.) Mill.)	Asparagaceae	Bulb
<i>Leucophae</i> spp. See <i>Sideritis</i> spp		
<i>Ligusticum porteri</i> J.M.Coult. & Rose	Apiaceae (Umbelliferae)	Root
<i>Ligustrum lucidum</i> Aiton	Oleaceae	Fruit
<i>Linzia nigriflora</i> (Oliv. & Hiern) C.Jeffrey (<i>Vernonia nigriflora</i> Oliv. & Hiern)	Asteraceae (Compositae)	
<i>Liriope</i> spp.	Asparagaceae	Fruit
<i>Lomatium dissectum</i> (Nutt.) Mathias & Constance	Apiaceae (Umbelliferae)	
<i>Lysimachia vulgaris</i> L.	Primulaceae	
<i>Lythrum</i> spp.	Lythraceae	Aerial part
<i>Malpighia glabra</i> L. (<i>Malpighia puniceifolia</i> L.)	Malpighiaceae	Bark
<i>Mangifera indica</i> L.	Anacardiaceae	Bark
<i>Marsdenia sylvestris</i> (Retz.) P.I.Forst (<i>Gymnema sylvestre</i> (Retz.) R.Br., <i>Gymnema sylvestre</i> (Retz.) Schult.)	Apocynaceae	Leaf
<i>Mentzelia cordifolia</i> Urb. & Gigl.	Loasaceae	Aerial part
<i>Mitchella repens</i> L.	Rubiaceae	Fruit and leaf
<i>Morella cerifera</i> (L.) Small.	Myricaceae	
<i>Moringa aptera</i> Gaertn.	Moringaceae	Fruit
<i>Moringa peregrina</i> (Forssk.) Fiori	Moringaceae	
<i>Muscari comosum</i> (L.) Mill. See <i>Leopoldia comosa</i> (L.) Parl.		
<i>Nardostachys grandiflora</i> DC. (<i>N. jatamansi</i> DC.)	Caprifoliaceae	Root
<i>Naregamia alata</i> Wight. & Arn.	Meliaceae	Whole plant
<i>Nectandra coto</i> Rusby	Lauraceae	Bark
<i>Nectandra puchury-major</i> Nees & Mart. ex Nees	Lauraceae	Seed
<i>Nectandra rodioei</i> Hook.	Lauraceae	Bark
<i>Nilgirianthus ciliatus</i> (Nees) Bremek.	Acanthaceae	
<i>Oenanthe phellandrium</i> Lam.	Apiaceae (Umbelliferae)	Fruit

Botanical name	Family	Part of plants indicated in the national list(s)
<i>Oplopanax elatus</i> (Nakai) Nakai	Araliaceae	Bark, leaf and root.
<i>Palicourea densiflora</i> Mart. See <i>Rudgea viburnoides</i> ssp. <i>viburnoides</i>		
<i>Palicourea densiflora</i> Wawra See <i>Psychotria longipedunculata</i> (Gardner) Müll.		
<i>Parietaria judaica</i> L.	Urticaceae	
<i>Pelargonium graveolens</i> L'Hér.	Geraniaceae	Aerial part
<i>Phyllanthus fraternus</i> G.L.Webster	Phyllanthaceae	Aerial part
<i>Picea glauca</i> (Moench) Voss	Pinaceae	
<i>Picrorhiza kurrooa</i> Benth. See <i>Picrorhiza lindleyana</i> (Wall.) Steud.		
<i>Picrorhiza lindleyana</i> (Wall.) Steud. (<i>Picrorhiza kurrooa</i> Benth.)	Scrophulariaceae	Rhizome and root
<i>Polemonium caeruleum</i> L.	Polemoniaceae	
<i>Polemonium reptans</i> L.	Polemoniaceae	
<i>Polygonatum sibiricum</i> F.Delaroche	Asparagaceae	Rhizome
<i>Polygonatum tegundir</i>	Species could not be identified	
<i>Polygonum aviculare</i> L.	Polygonaceae	Whole plant
<i>Pseudowintera colorata</i> (Raoul) Dandy	Winteraceae	Leaf
<i>Psychotria longipedunculata</i> (Gardner) Müll.Arg.	Rubiaceae	
<i>Pterocarpus santalinus</i> L.f.	Leguminosae (Fabaceae)	Bark and wood
<i>Ptychopetalum olacoides</i> Benth.	Olacaceae	Root
<i>Ptychopetalum uncinatum</i> Anselmino	Olacaceae	
<i>Rhus toxicodendron</i> L. See <i>Toxicodendron pubescens</i> Mill.		
<i>Richardia scabra</i> L.	Rubiaceae	
<i>Rivina humilis</i> L.	Phytolaccaceae	Whole plant
<i>Rosa moschata</i> Herrm.	Rosaceae	
<i>Rosa rubiginosa</i> L.	Rosaceae	
<i>Rudgea viburnoides</i> ssp. <i>viburnoides</i>	Rubiaceae	
<i>Salvia columbariae</i> Benth.	Lamiaceae (Labiatae)	
<i>Santalum austrocaledonicum</i> Vieill.	Santalaceae	
<i>Santalum spicatum</i> (R.Br.) A.DC.	Santalaceae	
<i>Santolina chamaecyparissus</i> L.	Asteraceae (Compositae)	
<i>Saponaria rubra</i> Lam.	Caryophyllaceae	Seed
<i>Saraca indica</i> L.	Leguminosae (Fabaceae)	Seed
<i>Scrophularia umbrosa</i> Dumort.	Scrophulariaceae	
<i>Scutellaria lateriflora</i> L.	Lamiaceae (Labiatae)	Aerial part
<i>Sedum reflexum</i> L. See <i>Sedum rupestre</i> L.		
<i>Sedum rupestre</i> L. (<i>Sedum reflexum</i> L.)	Crassulaceae	
<i>Sedum telephium</i> L. (<i>Hylotelephium telephium</i> (L.) H.Ohba.)	Crassulaceae	

Botanical name	Family	Part of plants indicated in the national list(s)
<i>Sida spodochroma</i> F.Muell.	Malvaceae	
Sideritis spp. (<i>Leucophaea</i> spp.)	Lamiaceae (Labiatae)	
<i>Smilax aristolochiifolia</i> Mill. (<i>Smilax kerberi</i> F.W.Apt.)	Smilacaceae	Root
<i>Smilax bona-nox</i> L.	Smilacaceae	
<i>Smilax cordata-ovata</i> Rich.	Smilacaceae	
<i>Smilax febrifuga</i> Kunth See <i>Smilax purhampuy</i> Ruiz		
<i>Smilax kerberi</i> F.W.Apt. See <i>Smilax aristolochiifolia</i> Mill.		
<i>Smilax longifolia</i> Rich. (<i>Smilax papyracea</i> Duhamel)	Smilacaceae	
<i>Smilax papyracea</i> Duhamel See <i>Smilax longifolia</i> Rich.	Smilacaceae	
<i>Smilax purhampuy</i> Ruiz (<i>Smilax febrifuga</i> Kunth)	Smilacaceae	
<i>Smilax tamnoides</i> L.	Smilacaceae	
<i>Solanandra grandiflora</i> Sw.	Solanaceae	
<i>Solenostemma argel</i> Hayne	Asclepiadaceae	Leaf
<i>Soymida febrifuga</i> Juss.	Meliaceae	Fruit
<i>Stephanotis</i> spp.	Apocyanaceae	
<i>Stevia salicifolia</i> Cav.	Asteraceae (Compositae)	
<i>Tanacetum morifolium</i> Kitam. See <i>Chrysanthemum sinense</i> Sweet		
<i>Tecoma</i> spp.	Bignoniaceae	
<i>Toona ciliata</i> M.Roem. (<i>Cedrela toona</i> Roxb.)	Meliaceae	
<i>Torilis japonica</i> (Houtt.) DC.	Apiaceae (Umbelliferae)	
<i>Toxicodendron pubescens</i> Mill. (<i>Rhus toxicodendron</i> L.)	Anacardiaceae	Whole plant
<i>Toxicodendron radicans</i> (L.) Kuntze	Anacardiaceae	Whole plant
<i>Toxicodendron vernix</i> (L.) Kuntze	Anacardiaceae	Whole plant
<i>Veronicastrum virginicum</i> (L.) Farw.	Plantagenaceae	
<i>Vernonia nigritana</i> Oliv. & Hiern See <i>Linzia nigritana</i> (Oliv. & Hiern) C.Jeffrey		
<i>Vincetoxicum versicolor</i> (Bunge) Dechne (<i>Cynanchum versicolor</i> Bunge)	Apocynaceae	
<i>Vitex negundo</i> L.	Lamiaceae (Labiatae)	
<i>Vitex trifolia</i> L.	Lamiaceae (Labiatae)	
<i>Vladimiria souliei</i> (Franch.) Y.Ling	Asteraceae (Compositae)	
<i>Withania coagulans</i> (Stocks) Dunal	Solanaceae	Whole plant
<i>Ziziphus oenoplia</i> (L.) Mill.	Rhamnaceae	
FUNGI		
<i>Elaphomyces granulatus</i> Fr.	Elaphomycetaceae	Fruiting body
<i>Trametes versicolor</i> (L.) Lloyd	Polyporaceae	Fruiting body
<i>Volvaria</i> spp.	Pluteaceae	Fruiting body

Botanicals appearing on a negative list or subject to restricted use in at least one European Member State but for which the Scientific Committee, through the analysis of the data found, could not identify substances of concern, or other data for the inclusion in the compendium. A systematic literature search should be performed for these species

Botanical name	Family	part of plants indicated in the national list(s)
<i>Abelmoschus moschatus</i> Medik.	Malvaceae	
<i>Abies alba</i> Mill.	Pinaceae	Bud and cone
<i>Abies balsamea</i> (L.) Mill.	Pinaceae	Bud and cone
<i>Abies sibirica</i> Ledeb.	Pinaceae	Bud and cone
<i>Abies spectabilis</i> (D.Don.) Mirb.	Pinaceae	Bud and cone
<i>Abutilon theophrasti</i> Medik. (<i>Abutilon avicennae</i> Gaertn.)	Malvaceae	Fruit and seed
<i>Achyrocline satureioides</i> (Lam.) DC. (<i>A. satureioides</i> (Lam.) DC.)	Asteraceae (Compositae)	
<i>Adenophora stricta</i> Miq.	Campanulaceae	
<i>Adiantum capillus-veneris</i> L.	Adiantaceae	
<i>Aesculus hippocastanum</i> L.	Sapindaceae	Bark and seed
<i>Agrimonia odorata</i> Mill. See <i>Agrimonia repens</i> L.		
<i>Agrimonia eupatoria</i> L.	Rosaceae	Aerial part
<i>Agrimonia repens</i> L. (<i>Agrimonia odorata</i> Mill.)	Rosaceae	
<i>Agropyron repens</i> (L.) P.Beauv. See <i>Elymus repens</i> (L.) Gould		
<i>Akebia quinata</i> (Houtt.) Decne.	Lardizabalaceae	Whole plant
<i>Akebia trifoliata</i> (Thunb.) Koidz.	Lardizabalaceae	Whole plant
<i>Alchemilla alpina</i> L.	Rosaceae	
<i>Alchemilla arvensis</i> (L.) Scop. See <i>Aphanes arvensis</i> L.		
<i>Alchemilla vulgaris</i> auct. pl. See <i>Alchemilla xanthochlora</i> Rothm.		Whole plant
<i>Alchemilla xanthochlora</i> Rothm. (<i>Alchemilla vulgaris</i> auct. pl.)	Rosaceae	Aerial part
<i>Alnus glutinosa</i> (L.) Gaertn.	Betulaceae	Bark and leaf
<i>Aloysia citriodora</i> Palau (<i>A. citrodora</i> Palau, <i>A. triphylla</i> (L'Hér.) Britton, <i>Lippia triphylla</i> (L'Hérit.) Kuntze, <i>L. citriodora</i> (Lam.) Kunth, <i>L. citrodora</i> Kunth)	Verbenaceae	
<i>Althaea officinalis</i> L.	Malvaceae	
<i>Amyris balsamifera</i> L.	Rutaceae	
<i>Ananas comosus</i> (L.) Merr. (<i>Ananas sativus</i> (Lindl.) Schult.f.)	Bromeliaceae	
<i>Antennaria dioica</i> (L.) Gaertn. (<i>Gnaphalium dioicum</i> L.)	Asteraceae (Compositae)	
<i>Anthemis nobilis</i> L. See <i>Chamaemelum nobile</i> (L.) All.		
<i>Aphanes arvensis</i> L. (<i>Alchemilla arvensis</i> (L.) Scop.)	Rosaceae	Aerial part
<i>Aralia racemosa</i> L.	Araliaceae	Rhizome and root
<i>Arbutus unedo</i> L.	Ericaceae	Leaf

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<i>Arctium lappa</i> L. (<i>Arctium majus</i> Bernh.)	Asteraceae (Compositae)	
<i>Avena sativa</i> L.	Poaceae (Gramineae)	
<i>Baccharis coridifolia</i> DC.	Asteraceae (Compositae)	Flower and seed
<i>Bacopa monnieri</i> (L.) Pennell (<i>Bacopa monniera</i> (L.) Wettst.)	Plantaginaceae	Leaf
<i>Betula nigra</i> L.	Betulaceae	Leaf
<i>Betula pendula</i> Roth.	Betulaceae	Leaf
<i>Betula pubescens</i> Ehrh.	Betulaceae	Leaf
<i>Bidens tripartita</i> L.	Asteraceae (Compositae)	Aerial part
<i>Bistorta officinalis</i> Delabre (<i>Polygonum bistorta</i> L.)	Polygonaceae	Rhizome
<i>Bupleurum falcatum</i> L.	Apiaceae (Umbelliferae)	Root
<i>Callitris introtropica</i> R.T.Baker & H.G.Sm.	Cupressaceae	
<i>Calophyllum inophyllum</i> L.	Calophyllaceae	Fruit and resin from the trunk
<i>Carex arenaria</i> L.	Cyperaceae	
<i>Carlina acaulis</i> L.	Asteraceae (Compositae)	Root
<i>Catalpa bignonioides</i> Walter (<i>C. syringifolia</i> Sims)	Bignoniaceae	leaf, pod and seed
<i>Catoferia spicata</i> (Benth.) Benth. (<i>Orthosiphon spicatus</i> Benth.)	Lamiaceae (Labiatae)	
<i>Cedrus atlantica</i> (Endl.) Carrière	Pinaceae	Bud and wood
<i>Centaurium erythraea</i> Raf.	Gentianaceae	Flowering top
<i>Chamaemelum nobile</i> (L.) All. (<i>Anthemis nobilis</i> L.)	Asteraceae (Compositae)	Whole plant
<i>Chelone glabra</i> L.	Plantaginaceae	Leaf
<i>Cnicus benedictus</i> L.	Asteraceae (Compositae)	Whole plant
<i>Collinsonia canadensis</i> L.	Lamiaceae (Labiatae)	Leaf, root and shoot
<i>Conyza canadensis</i> (L.) Cronquist	Asteraceae (Compositae)	Whole plant
<i>Coridothymus capitatus</i> (L.) Rchb.f.	Lamiaceae (Labiatae)	Aerial part
<i>Cornus officinalis</i> Siebold & Zucc.	Cornaceae	Fruit
<i>Corylus avellana</i> L.	Betulaceae	Leaf and nut
<i>Corymbia citriodora</i> (Hook.) K.D.Hill. & L.A.S.Johnson	Myrtaceae	Leaf
<i>Crataegus azarolus</i> L.	Rosaceae	
<i>Crataegus laevigata</i> (Poiret) DC.	Rosaceae	
<i>Crataegus monogyna</i> Jacq.	Rosaceae	
<i>Crataegus nigra</i> Waldst. & Kit.	Rosaceae	
<i>Crataegus pentagyna</i> Willd.	Rosaceae	
<i>Crataegus rhipidophylla</i> Gand. var. <i>rhipidophylla</i>	Rosaceae	
<i>Cryptomeria japonica</i> (L.f.) D.Don.	Taxodiaceae	Wood dust
<i>Cupressus sempervirens</i> L.	Cupressaceae	Cone
<i>Cyanotis vaga</i> (Lour.) Schult. & Schult.f.	Commelinaceae	Root
<i>Cydonia oblonga</i> P.Mill.	Rosaceae	Seed
<i>Cynara cardunculus</i> L.	Asteraceae (Compositae)	
<i>Cyperus scariosus</i> R.Br.	Cyperaceae	Rhizome and root
<i>Cystoseira canariensis</i> Sauvageau (<i>C. humilis</i> Schousboe ex Kützing)	Sargassaceae	Thallus
<i>Dipterocarpus retusus</i> Blume	Dipterocarpaceae	

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<i>Dittrichia graveolens</i> (L.) Greuter	Asteraceae (Compositae)	Leaf
<i>Echinacea angustifolia</i> DC.	Asteraceae (Compositae)	
<i>Echinacea pallida</i> (Nutt.) Nutt.	Asteraceae (Compositae)	
<i>Echinacea purpurea</i> (L.) Moench	Asteraceae (Compositae)	
<i>Eleutherococcus senticosus</i> (Rupr. & Maxim.) Maxim.	Araliaceae	
<i>Eleutherococcus sessiliflorus</i> (Rupr. & Maxim.) S.Y.Hu	Araliaceae	
<i>Elymus repens</i> (L.) Gould (<i>Agropyron repens</i> (L.) P.Beauv., <i>Elytrigia repens</i> (L.) Nevski)	Poaceae (Gramineae)	
<i>Elytrigia repens</i> (L.) Nevski See <i>Elymus repens</i> (L.) Gould		
<i>Ephedra nevadensis</i> Wats.	Ephedraceae	
<i>Equisetum arvense</i> L.	Equisetaceae	Aerial part
<i>Eriodictyon californicum</i> (Hook. & Arn.) Torr.	Boraginaceae	Aerial part
<i>Eucommia ulmoides</i> Oliv.	Eucommiaceae	Bark and leaf
<i>Euphorbia hirta</i> L. (<i>Chamaesyce hirta</i> (L.) Millesp.)	Euphorbiaceae	Aerial part
<i>Euphrasia officinalis</i> L.	Orobanchaceae	
<i>Filipendula ulmaria</i> (L.) Maxim.	Rosaceae	Aerial part
<i>Filipendula vulgaris</i> Moench	Rosaceae	Aerial part
<i>Fragaria vesca</i> L.	Rosaceae	Aerial part
<i>Fraxinus excelsior</i> L.	Oleaceae	Bark
<i>Galium verum</i> L.	Rubiaceae	Aerial part
<i>Gentiana cruciata</i> L.	Gentianaceae	
<i>Gentiana lutea</i> L.	Gentianaceae	
<i>Geranium robertianum</i> L.	Geraniaceae	
<i>Geum rivale</i> L.	Rosaceae	Rhizome and root
<i>Geum urbanum</i> L.	Rosaceae	Rhizome and root
<i>Gevuina avellana</i> Molina	Proteaceae	
<i>Globularia vulgaris</i> L.	Plantaginaceae	Aerial part
<i>Harpagophytum procumbens</i> Meisn.	Pedaliaceae	Root
<i>Harpagophytum zeyheri</i> Decne.	Pedaliaceae	Root
<i>Helichrysum arenarium</i> (L.) Moench	Asteraceae (Compositae)	
<i>Hieracium pilosella</i> L. (<i>Pilosella officinarum</i> F.W.Schultz & Sch.Bip.)	Asteraceae (Compositae)	Aerial part
<i>Hordeum vulgare</i> L.	Poaceae (Gramineae)	Seed
<i>Inula helenium</i> L.	Asteraceae (Compositae)	Root
<i>Ipomoea aquatica</i> Forssk.	Convolvulaceae	
<i>Iris germanica</i> L.	Iridaceae	Rhizome and root
<i>Iris pallida</i> Lam.	Iridaceae	Rhizome and root
<i>Jasminum officinale</i> L.	Oleaceae	Flower
<i>Lactuca serriola</i> L.	Asteraceae (Compositae)	
<i>Lamium album</i> L.	Lamiaceae (Labiatae)	
<i>Lespedeza capitata</i> Michx.	Fabaceae (Leguminosae)	Aerial part
<i>Leuzea carthamoides</i> (Willd.) DC. See <i>Rhaponticum carthamoides</i> (Willd.) Iljin		
<i>Lippia triphylla</i> (L'Hérit.) Kuntze See <i>Aloysia citrodora</i> Palau		
<i>Lobaria pulmonaria</i> (L.) Hoffm.	Lobariaceae	Thallus

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<i>Lythrum salicaria</i> L.	Lythraceae	Aerial part
<i>Magnolia fargesii</i> (Finet & Gagnep.) W.C.Cheng (<i>M. biondii</i> Pamp.)	Magnoliaceae	
<i>Malva sylvestris</i> L.	Malvaceae	Aerial part
<i>Marrubium vulgare</i> L.	Lamiaceae (Labiatae)	Aerial part
<i>Matricaria recutita</i> L. (<i>Chamomilla recutita</i> (L.) Rauschert)	Asteraceae (Compositae)	Flower
<i>Melissa officinalis</i> L.	Lamiaceae (Labiatae)	Aerial part
<i>Myroxylon balsamum</i> (L.) Harms	Fabaceae (Leguminosae)	Trunk balsam
<i>Oenothera biennis</i> L.	Onagraceae	Seed
<i>Olea europaea</i> L.	Oleaceae	Aerial part
<i>Ononis spinosa</i> L.	Fabaceae (Leguminosae)	Whole plant
<i>Opuntia ficus-indica</i> (L.) Mill.	Cactaceae	
<i>Oroxylum indicum</i> (L.) Kurz	Bignoniaceae	Fruit and seed
<i>Orthosiphon aristatus</i> (Blume) Miq. <i>Orthosiphon spicatus</i> Benth. See <i>Catoferia spicata</i> (Benth.) Benth.	Lamiaceae (Labiatae)	Aerial part
<i>Paeonia lactiflora</i> Pall.	Paeoniaceae	
<i>Paeonia officinalis</i> L.	Paeoniaceae	
<i>Paeonia suffruticosa</i> Andr.	Paeoniaceae	
<i>Panax ginseng</i> C.A.Mey.	Araliaceae	Root
<i>Panax quinquefolius</i> L.	Araliaceae	Root
<i>Papaver rhoeas</i> L.	Papaveraceae	Aerial part
<i>Parthenium integrifolium</i> L.	Asteraceae (Compositae)	
<i>Passiflora edulis</i> Sims	Passifloraceae	Whole plant
<i>Passiflora incarnata</i> L.	Passifloraceae	Whole plant
<i>Pinus sylvestris</i> L.	Pinaceae	Oleo-resin from the trunk
<i>Plantago afra</i> L.	Plantaginaceae	Leaf and seed
<i>Plantago arenaria</i> Waldst. & Kit.	Plantaginaceae	Leaf and seed
<i>Plantago lanceolata</i> L.	Plantaginaceae	Leaf and seed
<i>Plantago major</i> L.	Plantaginaceae	Leaf and seed
<i>Plantago media</i> L.	Plantaginaceae	Leaf and seed
<i>Plantago ovata</i> Forssk.	Plantaginaceae	Leaf and seed
<i>Platycodon grandiflorus</i> (Jacq.) A.DC.	Campanulaceae	
<i>Pogostemon cablin</i> Benth.	Lamiaceae (Labiatae)	
<i>Polianthes tuberosa</i> L. <i>Polygonum bistorta</i> L. See <i>Bistorta officinalis</i> Delabre	Asparagaceae	Aerial part
<i>Polypodium vulgare</i> L.	Polypodiaceae	Rhizome and root
<i>Potentilla anserina</i> L.	Rosaceae	Root
<i>Primula elatior</i> (L.) Hill.	Primulaceae	Whole plant
<i>Primula veris</i> L.	Primulaceae	Whole plant
<i>Primula vulgaris</i> Huds.	Primulaceae	
<i>Prunus africana</i> (Hook.f.) Kalkman	Rosaceae	
<i>Prunus cerasus</i> L.	Rosaceae	
<i>Pseudotsuga menziesii</i> (Mirb.) Franco	Pinaceae	
<i>Pterocarpus marsupium</i> Roxb.	Fabaceae (Leguminosae)	Bark and wood

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<i>Raphanus sativus</i> var. <i>niger</i> J.Kern (<i>Raphanus sativus</i> L. convar. <i>sativus</i> Radish group)	Brassicaceae	
<i>Rhaponticum carthamoides</i> (Willd.) Iljin (<i>Stemmacantha carthamoides</i> (Willd.) Dittrich, <i>Leuzea carthamoides</i> (Willd.) DC.)	Asteraceae (Compositae)	
<i>Rhodiola rosea</i> L.	Crassulaceae	
<i>Ribes nigrum</i> L.	Grossulariaceae	
<i>Rosa canina</i> L.	Rosaceae	
<i>Rosa damascena</i> Mill.	Rosaceae	Flower and leaf
<i>Rubus fruticosus</i> L. s.l.	Rosaceae	
<i>Sanguisorba officinalis</i> L.	Rosaceae	Root
<i>Santalum album</i> L.	Santalaceae	
<i>Schinus molle</i> L.	Anacardiaceae	Fruit and leaf
<i>Schisandra chinensis</i> (Turcz.) Baill.	Schisandraceae	Fruit
<i>Schisandra sphenanthera</i> Rehd. et Wills.	Schisandraceae	Fruit
<i>Scrophularia nodosa</i> L.	Scrophulariaceae	Whole plant
<i>Selenicereus grandiflorus</i> Britton & Rose (<i>Cactus grandiflorus</i> L.)	Cactaceae	Aerial part
<i>Silybum marianum</i> (L.) Gaertn.	Asteraceae (Compositae)	Flowering top and seed
<i>Solidago canadensis</i> L.	Asteraceae (Compositae)	
<i>Solidago virgaurea</i> L.	Asteraceae (Compositae)	
<i>Sorbus aucuparia</i> L.	Rosaceae	Fruit
<i>Sorbus domestica</i> L.	Rosaceae	Fruit
<i>Spergularia rubra</i> (L.) J.Presl. & C.Presl.	Caryophyllaceae	
<i>Stachys officinalis</i> (L.) Trevis.	Lamiaceae (Labiatae)	
<i>Stellaria media</i> (L.) Vill.	Caryophyllaceae	Whole plant
<i>Stellaria dichotoma</i> L.	Caryophyllaceae	Whole plant
<i>Stemmacantha carthamoides</i> (Willd.) Dittrich See <i>Rhaponticum carthamoides</i> (Willd.) Iljin		
<i>Styrax benzoides</i> Craib.	Styracaceae	Resin from the trunk
<i>Swertia chirayita</i> (Roxb.) H.Karst.	Gentianaceae	Whole plant
<i>Tamarindus indica</i> L.	Fabaceae (Leguminosae)	
<i>Taraxacum officinale</i> F.H.Wigg., s.l.	Asteraceae (Compositae)	
<i>Terminalia arjuna</i> (Roxb.) Wight & Arn.	Combretaceae	
<i>Terminalia bellirica</i> (Gaertn.) Roxb.	Combretaceae	
<i>Tilia cordata</i> Mill.	Malvaceae	
<i>Tilia europaea</i> L.	Malvaceae	
<i>Tilia platyphyllos</i> Scop.	Malvaceae	
<i>Tilia tomentosa</i> Moench	Malvaceae	
<i>Trigonella foenum-graecum</i> L.	Fabaceae (Leguminosae)	
<i>Triticum aestivum</i> L. subsp. <i>aestivum</i>	Poaceae (Gramineae)	
<i>Turnera diffusa</i> Schult.	Passifloraceae	
<i>Uncaria tomentosa</i> (Schult.) DC.	Rubiaceae	Aerial part
<i>Urtica dioica</i> L.	Urticaceae	Aerial part
<i>Vaccinium myrtillus</i> L.	Ericaceae	Leaf
<i>Vaccinium vitis-idaea</i> L.	Ericaceae	Leaf
<i>Valeriana procera</i> Kunth	Caprifoliaceae	
<i>Valeriana officinalis</i> L.	Caprifoliaceae	
<i>Valeriana repens</i> Host	Caprifoliaceae	
<i>Verbascum densiflorum</i> Bertol.	Scrophulariaceae	
<i>Verbascum phlomoides</i> L.	Scrophulariaceae	
<i>Verbena officinalis</i> L.	Verbenaceae	

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<i>Veronica officinalis</i> L.	Plantaginaceae	
<i>Viburnum lantana</i> L.	Caprifoliaceae	
<i>Viburnum opulus</i> L.	Caprifoliaceae	
<i>Viburnum prunifolium</i> L.	Caprifoliaceae	Bark
<i>Viola arvensis</i> Murray	Violaceae	
<i>Viola odorata</i> L.	Violaceae	Flower and leaf
<i>Viola tricolor</i> L.	Violaceae	Flower and leaf
<i>Vitis vinifera</i> L.	Vitaceae	Fruit, leaf and seed
<i>Ziziphus jujuba</i> Mill.	Rhamnaceae	

FUNGI

<i>Cordyceps sinensis</i> (Berk.) Sacc. See <i>Ophiocordyceps sinensis</i> (Berk.) G.H.Sung, J.M.Sung, Hywel-Jones & Spatafora		
<i>Ganoderma lucidum</i> (Curtis) P.Karst.	Ganodermataceae	Fruiting body
<i>Ophiocordyceps sinensis</i> (Berk.) G.H.Sung, J.M.Sung, Hywel-Jones & Spatafora (<i>Cordyceps sinensis</i> (Berk.) Sacc.)	Ophiocordycipitaceae	Mycelium
<i>Poria cocos</i> F.A.Wolf See <i>Wolfiporia extensa</i> (Peck) Ginns.		
<i>Wolfiporia cocos</i> (F.A. Wolf) Ryvarden & Gilb. See <i>Wolfiporia extensa</i> (Peck) Ginns.		
<i>Wolfiporia extensa</i> (Peck) Ginns. (<i>Wolfiporia cocos</i> (F.A. Wolf) Ryvarden & Gilb., <i>Poria cocos</i> F.A.Wolf)	Polyporaceae	Fruiting body

ALGAE

<i>Aphanizomenon flos-aquae</i> Bornet & Flauhault (<i>Byssus flos-aquae</i> L.)	Nostocaceae	
<i>Spirulina maxima</i> (Setchell & Gardner) Geitler	Pseudanabaenaceae	